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| International Union for the Protection of New Varieties of Plants |  |

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| Technical Working Party for Vegetables  Fifty-Third Session Seoul, Republic of Korea, May 20 to 24, 2019 | TWV/53/7  Original: English  Date: April 11, 2019 |

Partial revision of the Test Guidelines for TOMATO

Document prepared by experts from the Netherlands

Disclaimer: this document does not represent UPOV policies or guidance

The purpose of this document is to present a proposal for a partial revision of the Test Guidelines for Tomato (document TG/44/11 Rev.).

The Technical Working Party for Vegetables (TWV), at its fifty-second session, held in Beijing, China, from September 17 to 21, 2018, agreed that the Test Guidelines for Tomato (document TG/44/11 Rev.) be partially revised for Characteristics 48 “Resistance to *Fusarium oxysporum* f. sp. *lycopersici* (Fol)” and 53 “Resistance to *Pyrenochaeta lycopersici* (Pl)” (see document TWV/52/20 “Report”, Annex V).

The TWV, at its fifty-second session, considered documents TWV/52/11 and TWV/52/19 and agreed that Characteristic and Ad. 48 “Resistance to *Fusarium oxysporum* f. sp. *lycopersici* (Fol)” be excluded from the partial revision presented in document TWV/52/11 “Matters to be resolved concerning Test Guidelines adopted by the Technical Committee: partial revision of the test guidelines for tomato” as research was ongoing and agreed that this characteristic should be reconsidered by the TWV at its fifty-third session (see document TWV/52/20 “Report”, paragraph 62).

4. The following changes are proposed:

1. To change the example varieties and to change the denomination of the races of Characteristics 48.1, 48.2 and 48.3 “Resistance to *Fusarium oxysporum* f. sp. *lycopersici* (Fol)”;
2. To change the explanation Ad. 48 by adding an alternative method to observe the resistance and by minor changes in the current method in Chapter 8.2 “Explanations for individual characteristics”;
3. To change the example varieties of Characteristic 53 “Resistance to *Pyrenochaeta lycopersici* (Pl)”;
4. To change the explanation Ad. 53 in Chapter 8.2 “Explanations for individual characteristics”

5. The proposed changes are presented below in highlight and underline (insertion) and ~~strikethrough~~ (deletion).

Proposal to change the example varieties and to change the denomination of the races of Characteristics 48.1, 48.2 and 48.3 “Resistance to *Fusarium oxysporum* f. sp. *lycopersici* (Fol)”

*Current wording*

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
|  |  | English | français | deutsch | español | Example Varieties Exemples Beispielssorten Variedades ejemplo | Note/ Nota |
| 48.   (+) | VG | Resistance to  *Fusarium oxysporum* f. sp. *lycopersici* (Fol) | Résistance à *Fusarium oxysporum* f. sp. *lycopersici* (Fol) | Resistenz gegen *Fusarium oxysporum* f. sp. *lycopersici* (Fol) | Resistencia a  *Fusarium oxysporum* f. sp. *lycopersici* (Fol) |  |  |
| **48.1  (\*)** | **VG** | **– Race 0 (ex 1)** | **– Pathotype 0 (ex 1)** | **– Pathotyp 0 (ex 1)** | **– Raza 0 (ex 1)** |  |  |
| **QL** |  | absent | absente | fehlend | ausente | Marmande verte | 1 |
|  |  | present | présente | vorhanden | presente | Anabel, Marporum, Marsol | 9 |
| **48.2  (\*)** | **VG** | **– Race 1 (ex 2)** | **– Pathotype 1 (ex 2)** | **– Pathotyp 1 (ex 2)** | **– Raza 1 (ex 2)** |  |  |
| **QL** |  | absent | absente | fehlend | ausente | Marmande verte | 1 |
|  |  | present | présente | vorhanden | presente | Motelle, Walter | 9 |
| **48.3** | **VG** | **– Race 2 (ex 3)** | **– Pathotype 2 (ex 3)** | **– Pathotyp 2 (ex 3)** | **– Raza 2 (ex 3)** |  |  |
| **QL** |  | absent | absente | fehlend | ausente | Marmande verte, Motelle | 1 |
|  |  | present | présente | vorhanden | presente | Alliance, Florida, Ivanhoé, Tributes | 9 |

*Proposed new wording*

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
|  |  | English | français | deutsch | español | Example Varieties Exemples Beispielssorten Variedades ejemplo | Note/ Nota |
| 48.   (+) | VG | Resistance to  *Fusarium oxysporum* f. sp. *lycopersici* (Fol) | Résistance à *Fusarium oxysporum* f. sp. *lycopersici* (Fol) | Resistenz gegen *Fusarium oxysporum* f. sp. *lycopersici* (Fol) | Resistencia a  *Fusarium oxysporum* f. sp. *lycopersici* (Fol) |  |  |
| **48.1  (\*)** | **VG** | **– Race ~~0 (ex 1)~~ 0EU/1US** | **– Pathotype ~~0 (ex 1)~~ 0EU/1US** | **– Pathotyp ~~0 (ex 1)~~ 0EU/1US** | **– Raza ~~0 (ex 1)~~ 0 EU/1US** |  |  |
| **QL** |  | absent | absente | fehlend | ausente | Marmande verte, Moneymaker | 1 |
|  |  | present | présente | vorhanden | presente | ~~Anabel~~, Marporum, ~~Marsol~~Motelle, Tradiro | 9 |
| **48.2  (\*)** | **VG** | **– Race ~~1 (ex 2)~~ 1EU/2US** | **– Pathotype ~~1 (ex 2)~~ 1EU/2US** | **– Pathotyp ~~1 (ex 2)~~ 1EU/2US** | **– Raza ~~1 (ex 2)~~ 1EU/2US** |  |  |
| **QL** |  | absent | absente | fehlend | ausente | Marmande verte, Marporum, Moneymaker | 1 |
|  |  | present | présente | vorhanden | presente | Motelle, ~~Walter~~Tradiro | 9 |
| **48.3** | **VG** | **– Race ~~2 (ex 3)~~ 2EU/3US** | **– Pathotype ~~2 (ex 3)~~ 2EU/3US** | **– Pathotyp ~~2 (ex 3)~~ 2EU/3US** | **– Raza ~~2 (ex 3)~~ 2EU/3US** |  |  |
| **QL** |  | absent | absente | fehlend | ausente | Marmande verte, Motelle | 1 |
|  |  | present | présente | vorhanden | presente | Alliance, Florida, ~~Ivanhoé~~Ivanhoe, Tributes | 9 |

Proposal to change the explanation Ad. 48 by adding an alternative method to observe the resistance and by minor changes in the current method in Chapter 8.2 “Explanations for individual characteristics”

*Current wording*

Ad. 48: Resistance to *Fusarium oxysporum* f. sp. *lycopersici* (Fol)

1. Pathogen *Fusarium oxysporum* f. sp. *lycopersici*

3. Host species *Solanum lycopersicum*

4. Source of inoculum Naktuinbouw[[1]](#footnote-2) (NL) and GEVES[[2]](#footnote-3) (FR)

5. Isolate Race 0 (ex 1) (e.g. strains Orange 71 or PRI 20698 or Fol 071 1 (ex 2) (e.g. strains 4152 or PRI40698 or RAF 70 and 2 (ex 3)

individual strains may vary in pathogenicity

6. Establishment isolate identity use differential varieties (see 9.3)

7. Establishment pathogenicity on susceptible tomato varieties

8. Multiplication inoculum

8.1 Multiplication medium Potato Dextrose Agar, Medium “S” of Messiaen

8.4 Inoculation medium water for scraping agar plates or Czapek-Dox culture medium

(7 d-old aerated culture)

8.6 Harvest of inoculum filter through double muslin cloth

8.7 Check of harvested inoculum spore count; adjust to 106 per ml

8.8 Shelf-life/viability inoculum 4-8 h, keep cool to prevent spore germination

9. Format of the test

9.1 Number of plants per genotype at least 20 plants

9.2 Number of replicates 1 replicate

9.3 Control varieties for the test with

race 0 (ex 1)

Susceptible Marmande, Marmande verte, Resal

Resistant for race 0 only Marporum, Larissa, “Marporum x Marmande verte”, Marsol, Anabel

Resistant for race 0 and 1 Motelle, Gourmet, Mohawk

Control varieties for the test with

race 1 (ex 2)

Susceptible Marmande verte, Cherry Belle, Roma

Resistant for race 0 only Marporum, Ranco

Resistant for race 0 and 1 Tradiro, Odisea

Remark: Ranco is slightly less resistant than Tradiro

Control varieties for the test with

race 2 (ex 3)

Susceptible for race 0, 1 and 2 Marmande verte, Motelle, Marporum

Resistant for race 0, 1 and 2 Tributes, Murdoch, Marmande verte x Florida

9.4 Test design >20 plants; e.g. 35 seeds for 24 plants, including 2 blanks

9.5 Test facility glasshouse or climate room

9.6 Temperature 24-28°C (severe test, with mild isolate)

20-24°C (mild test, with severe isolate)

9.7 Light 12 hours per day or longer

9.8 Season all seasons

9.9 Special measures slightly acidic peat soil is optimal;

keep soil humid but avoid water stress

10. Inoculation

10.1 Preparation inoculums aerated Messiaen or PDA or Agar Medium S of Messiaen or

Czapek Dox culture or scraping of plates

10.2 Quantification inoculums pore count, adjust to 106 spores per ml,

lower concentration for a very aggressive isolate

10.3 Plant stage at inoculation 10-18 d, cotyledon to first leaf

10.4 Inoculation method roots and hypocotyls are immersed in spore suspension

for 5-15 min; trimming of roots is an option

10.7 Final observations 14-21 days after inoculation

11. Observations

11.1 Method visual

11.2 Observation scale symptoms:

growth retardation, wilting, yellowing,

vessel browning extending above cotyledon

11.3 Validation of test………………… evaluation of variety resistance should be calibrated with results of resistant and susceptible controls. Standards near borderline R/S will help to compare between labs.

12. Interpretation of test results in comparison with control varieties

absent [1] severe symptoms

present [9] mild or no symptoms

13. Critical control points

Test results may vary slightly in inoculum pressure due to differences in isolate, spore concentration, soil humidity and temperature.

*Proposed new wording*

Ad. 48: Resistance to *Fusarium oxysporum* f. sp. *lycopersici* (Fol)

Resistance to race 0EU/1US and race 1EU/2US to be tested in a bio-assay (method i) or in a DNA marker test (method ii), if appropriate. Resistance to race 2EU/3US to be tested in a bio-assay (method i).

1. Bio-assay

|  |  |  |
| --- | --- | --- |
| 1. | Pathogen | *Fusarium oxysporum* f. sp. *lycopersici* |
| 3. | Host species | *Solanum lycopersicum* |
| 4. | Source of inoculum | Naktuinbouw[[3]](#footnote-4) (NL), GEVES[[4]](#footnote-5) (FR) or INIA[[5]](#footnote-6) (ES) |
| 5. | Isolate | race ~~0 (ex 1)~~0EU/1US(e.g. strains Orange 71 or PRI 20698 or Fol 071)  race ~~1 (ex 2)~~1EU/2US(e.g. strains 4152 or PRI40698 or RAF 70)  race ~~2 (ex 3)~~2EU/3US (e.g. strain Fol029)  ~~individual strains may vary in pathogenicity~~ |
| 6. | Establishment isolate identity | use differential varieties (see 9.3) |
| 7. | Establishment pathogenicity | on susceptible tomato varieties |
| 8. | Multiplication inoculum |  |
| 8.1 | Multiplication medium | Potato Dextrose Agar, Medium “S” of Messiaen |
| 8.4 | Inoculation medium | water for scraping agar plates or Czapek-Dox culture medium (7 d-old aerated culture) |
| 8.6 | Harvest of inoculum | filter through double muslin cloth |
| 8.7 | Check of harvested inoculum | spore count; adjust to 106 per ml |
| 8.8 | Shelflife/viability inoculum | 4-8 h, keep cool to prevent spore germination |
| 9. | Format of the test |  |
| 9.1 | Number of plants per genotype | at least 20 plants |
| 9.2 | Number of replicates | 1 replicate |
| 9.3.1 | Control varieties for the test with race ~~0 (ex 1)~~0EU/1US |  |
|  | Susceptible | Marmande, Marmande verte, Resal, Moneymaker |
|  | Resistant ~~for race 0 only~~ | Marporum, Larissa, “Marporum x Marmande verte”, ~~Marsol, Anabel,~~ Motelle, Gourmet, Mohawk, Tradiro |
|  | ~~Resistant for race 0 and 1~~ | ~~Motelle, Gourmet, Mohawk~~ |
| 9.3.2 | Control varieties for the test with race ~~1 (ex 2)~~1EU/2US |  |
|  | Susceptible | Marmande verte, Cherry Belle, Roma, Marporum, Ranco, Moneymaker |
|  | ~~Resistant for race 0 only~~ | ~~Marporum, Ranco~~ |
|  | Resistant ~~for race 0 and 1~~ | Tradiro, Odisea, “Motelle x Marmande verte”, Motelle |
| 9.3.3 | Control varieties for the test with race ~~2 (ex 3)~~2EU/3US |  |
|  | Susceptible ~~for race 0, 1 and 2~~ | Marmande verte, Motelle, Marporum |
|  | Resistant ~~for race 0, 1 and 2~~ | Tributes, Murdoch, “Marmande verte x Florida”, Alliance, Florida, Ivanhoe |
| 9.4 | Test design | >20 plants; e.g. 35 seeds for 24 plants, including 2 blanks |
| 9.5 | Test facility | glasshouse or climate room |
| 9.6 | Temperature | 24-28°C (severe test, with mild isolate)  20-24°C (mild test, with severe isolate) |
| 9.7 | Light | 12 hours per day or longer |
| 9.8 | Season | all seasons |
| 9.9 | Special measures | slightly acidic peat soil is optimal;  keep soil humid but avoid water stress |
| 10. | Inoculation |  |
| 10.1 | Preparation inoculum | aerated Messiaen or PDA or Agar Medium S of Messiaen or Czapek Dox culture or scraping of plates |
| 10.2 | Quantification inoculum | spore count, adjust to 106 spores per ml, lower concentration for a very aggressive isolate |
| 10.3 | Plant stage at inoculation | 10-18 d, cotyledon to first leaf |
| 10.4 | Inoculation method | roots and hypocotyls are immersed in spore suspension for 5‑15 min; trimming of roots is an option |
| 10.7 | Final observations | 14-21 days after inoculation |
| 11. | Observations |  |
| 11.1 | Method | visual |
| 11.2 | Observation scale | symptoms:  growth retardation, wilting, yellowing,  vessel browning extending above cotyledon |
| 11.3 | Validation of test | evaluation of variety resistance should be calibrated with results of resistant and susceptible controls. Standards near borderline R/S will help to compare between labs. |
| 12. | Interpretation of test results in comparison with control varieties |  |
|  | absent | [1] severe symptoms |
|  | present | [9] mild or no symptoms |
| 13. | Critical control points | Test results may vary slightly in inoculum pressure due to differences in isolate, spore concentration, soil humidity and temperature. |

(ii) DNA marker test

Dominant resistance gene I2 is always associated with resistance to both race 0EU/1US and race 1EU/2US. The presence or absence of the resistance allele can be detected by the co-dominant marker as described in this method.

|  |  |  |
| --- | --- | --- |
| 1. | Pathogen | *Fusarium oxysporum* f. sp. *lycopersici* |
| 2. | Functional gene | I2 |
| 3. | Primers |  |
| 3.1 | Allele for susceptibility | Z1063-i2-F 5’-GTT TGA CAG CTT GGT TTT GT-3’  Z1063-i2-R 5’-CTC AAA CTC ACC ATC ATT GA-3’ |
| 3.2 | Allele for resistance | TFusF1 5’-CTG AAA CTC TCC GTA TTT C-3’  TFusRR1 5’-CGA AGA GTG ATT GGA GAT-3’ |
| 4. | Format of the test |  |
| 4.1 | Number of plants per genotype | at least 20 plants |
| 4.2 | Control varieties | homozygous allele for susceptibility present: Marmande verte, Marporum, Moneymaker  homozygous allele for resistance present: Motelle, Tradiro  add blanks as negative PCR control |
| 5. | Preparation |  |
| 5.1 | Preparation DNA | harvest per individual plant a part of a young leaf. Isolate total DNA of each individual plant separately with a standard DNA isolation protocol (CTAB/SDS based). Re-suspend in 100 µl T10E0,1 or another suitable buffer. Dilute total DNA to 1/10 (H2O) to obtain a DNA concentration between 1-10 ng/µl. |
| 5.2 | Preparation PCR | for example, use 3 µl of each diluted DNA sample into individuals PCR reactions. prepare the PCR master mix, 20µl reaction volume, for example:   * 3 µl of 10x diluted DNA * 2,5 µl of 10x reaction buffer * 2 mM MgCl2 * 0.1 µM of resistance primers each * 0.2 µM of susceptible primers each * 200 µM of each of the four dNTPs * 1 unit of Taq DNA polymerase |
| 6. | PCR conditions | for example:  1. initial denaturation step at 94°C for 3 minutes  2. 35 cycles at 94°C for 1 minute, 56°C for 1 minute, and 72°C for 2 minutes  3. final extension step of 72°C for 10 minutes |
| 7. | Observations |  |
| 7.1 | Method | visual, after detection on gel together with a suitable ladder |
| 7.2 | Observation scale |  |

|  |  |  |
| --- | --- | --- |
|  |  |  |
| amplicon of 940bp only | amplicon of 600bp only | amplicons of 940bp and 600bp |
| homozygous susceptible allele present | homozygous resistant allele present | susceptible and resistant allele present: heterozygous resistant |

|  |  |  |
| --- | --- | --- |
| 7.3 | Validation of test | control varieties should give the expected band(s). |
| 8. | Interpretation of test results |  |
|  | 48.1 Resistance to race 0EU/1US |  |
|  | absent | [1] can not be concluded from the DNA-test, a bio-assay should be performed. |
|  | present | [9] homozygous resistant or heterozygous resistant in DNA marker test.  In case the DNA marker test result does not confirm the morphological declaration in the TQ, a bio-assay should be performed to observe whether the variety is resistant e.g. on another mechanism like gene I2 without I. |
|  | 48.2 Resistance to race 1EU/2US |  |
|  | absent | [1] homozygous susceptible in DNA marker test |
|  | present | [9] homozygous or heterozygous resistant in DNA marker test.  In case the DNA marker test result does not confirm the morphological declaration in the TQ, a bio-assay should be performed to observe whether the variety is resistant e.g. on another mechanism like gene I3. |

Proposal to change the example varieties of Characteristic 53 “Resistance to *Pyrenochaeta lycopersici* (Pl)”

*Current wording*

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| 53.  (+) | VG | Resistance to *Pyrenochaeta lycopersici* (Pl) | Résistance à *Pyrenochaeta lycopersici* (Pl) | Resistenz gegen *Pyrenochaeta lycopersici* (Pl) | Resistencia a *Pyrenochaeta lycopersici* (Pl) |  |  |
| QL |  | absent | absente | fehlend | ausente | Montfavet H 63.5 | 1 |
|  |  | present | présente | vorhanden | presente | Kyndia, Moboglan, Pyrella | 9 |

*Proposed new wording*

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| 53.  (+) | VG | Resistance to *Pyrenochaeta lycopersici* (Pl) | Résistance à *Pyrenochaeta lycopersici* (Pl) | Resistenz gegen *Pyrenochaeta lycopersici* (Pl) | Resistencia a *Pyrenochaeta lycopersici* (Pl) |  |  |
| QL |  | absent | absente | fehlend | ausente | ~~Montfavet H 63.5~~ Marmande verte | 1 |
|  |  | present | présente | vorhanden | presente | ~~Kyndia, Moboglan, Pyrella~~ Garance | 9 |

Proposal to change the explanation Ad. 53 in Chapter 8.2 “Explanations for individual characteristics”

*Current wording*

Ad. 53: Resistance to *Pyrenochaeta lycopersici* (Pl)

1. Pathogen *Pyrenochaeta lycopersici*

3. Host species *Solanum lycopersicum*

4. Source of inoculum -

5. Isolate -

7. Establishment pathogenicity biotest

8. Multiplication inoculum

8.1 Multiplication medium V8 Agar

8.2 Multiplication variety susceptible tomato variety

8.3 Plant stage at inoculation seed

8.4 Inoculation medium mixture of soil, e.g. (70%), sand (20%) and inoculum (10.1) (10%) or soil mixed with diseased roots cut to small pieces

8.5 Inoculation method sowing, or transplanting at fruit maturity

8.6 Harvest of inoculum diseased roots are harvested after 2-4 months

8.7 Check of harvested inoculum visual inspection of lesions on roots

8.8 Shelf-life/viability inoculum the fungus will not die quickly, but may lose its pathogenicity within a week after isolation on an agar medium

9. Format of the test

9.1 Number of plants per genotype 20 plants

9.2 Number of replicates 1 replicate

9.3 Control varieties

Susceptible Montfavet H 63.5

Resistant Kyndia, Moboglan, Pyrella

9.5 Test facility greenhouse or climate cell

9.6 Temperature day 24°C, night 14°C

9.7 Light 12 h minimum

10. Inoculation

10.1 Preparation inoculum e.g. double-autoclaved mixture of soil with 10% oatmeal added e.g. Incubate for 10-14 d at 20°C with occasional, repeated turning

10.3 Plant stage at inoculation 6 weeks

10.4 Inoculation method transplanting into mixture of soil, sand and inoculum (8.4) or soil mixed with diseased roots cut to small pieces or naturally infected soil

10.7 Final observations 6-8 weeks after transplanting (flowering plant)

11. Observations

11.1 Method visual

11.2 Observation scale symptoms: brown lesions on roots

11.3 Validation of test evaluation of variety resistance should be calibrated with results of resistant and susceptible controls

12. Interpretation of test results in comparison with control varieties

absent [1] symptoms

present [9] no symptoms

13. Critical control points:

The fungus loses its pathogenicity quickly after isolation on an agar medium. It is advisable to keep the isolate alive on living plants.

*Proposed new wording*

Ad. 53: Resistance to *Pyrenochaeta lycopersici* (Pl)

|  |  |  |
| --- | --- | --- |
| 1. | Pathogen | *Pyrenochaeta lycopersici* |
| 2. | Quarantine status | No |
| 3. | Host species | *Solanum lycopersicum* |
| 4. | Source of inoculum | GEVES[[6]](#footnote-7) (FR) |
| 5. | Isolate | e.g. strain Pl 21 |
| 6. | Establishment isolate identity | On susceptible plant |
| 8. | Multiplication inoculum |  |
| 8.1 | Multiplication medium | Messiaen agar or synthetic medium |
| 8.4 | Inoculation medium | Autoclaved grains (e.g. barley) |
| 8.5 | Inoculation method | Mix of contaminated grains (e.g. 1 kg) with inoculum (e.g. medium from 2 Petri dishes with mycelium) |
| 8.6 | Harvest of inoculum | After 3 weeks |
| 9. | Format of the test |  |
| 9.1 | Number of plants per genotype | At least 20 |
| 9.2 | Number of replicates | 1 replicate |
| 9.3 | Control varieties | Susceptible : Marmande verte  Resistant : Garance and (*S. lycopersicum* x *habrochaites*) Emperador |
| 9.4 | Test design | add non inoculated plants |
| 9.5 | Test facility | Greenhouse or climatic chamber |
| 9.6 | Temperature | 20°C |
| 9.7 | Light | At least 12h |
| 10. | Inoculation |  |
| 10.1 | Preparation inoculum | Homogenize the contaminated grains |
| 10.3 | Plant stage at inoculation | 3-4 leaf stage |
| 10.4 | Inoculation method | Transplanting of plantlets in a mixture of soil (e.g. 3750 ml of soil with 750 ml of inoculum) |
| 10.7 | Final observations | 40 days post inoculation |
| 11. | Observations |  |
| 11.1 | Method | visual |
| 11.2 | Observation scale | Class 0: no necrosic lesion on roots  Class 1: few small and uncoloured necrotic lesions  Class 2: some brown necrotic lesions clearly visible (less than half the surface of the pivot)  Class 3: several brown necrotic lesions clearly visible (more than half the surface of the pivot)  Class 4: complete necrosis or destruction of the pivot |
| 11.3 | Validation of test | Evaluation of variety resistance should be calibrated with results of resistant and susceptible controls |
| 12. | Interpretation of data in terms of UPOV characteristic states | Any variety judged to be of the same resistance level or higher than Garance is judged as resistant.  Classes 0, 1 and 2 are commonly judged as resistant – Note 9  Classes 3 and 4 are commonly judged as susceptible – Note 1 |

[End of document]

1. Naktuinbouw: resistentie@naktuinbouw.nl [↑](#footnote-ref-2)
2. GEVES; Valerie.GRIMAULT@geves.fr [↑](#footnote-ref-3)
3. Naktuinbouw: resistentie@naktuinbouw.nl [↑](#footnote-ref-4)
4. GEVES: matref@geves.fr [↑](#footnote-ref-5)
5. INIA: resistencias@inia.es [↑](#footnote-ref-6)
6. GEVES: matref@geves.fr [↑](#footnote-ref-7)