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INTERNATIONAL UNION FOR THE PROTECTION OF NEW VARIETIES OF PLANTS

Geneva

DRAFT

TURNIP

UPOV Code(s): BRASS_RAP_RAP

Brassica rapa L. var. rapa (L.) Thell.

GUIDELINES

FOR THE CONDUCT OF TESTS

FOR DISTINCTNESS, UNIFORMITY AND STABILITY

prepared by experts from France to be considered by the Technical Working Party for Vegetables at its fifty-first session, to be held in Roelofarendsveen, Netherlands, from 2017-07-03 to 2017-07-07

Disclaimer: this document does not represent UPOV policies or guidance

Alternative names:*

Botanical name	English	French	German	Spanish
<i>Brassica rapa</i> L. var. <i>rapa</i> (L.) Thell., <i>Brassica rapa</i> L. ssp. <i>rapa</i> ; var. <i>rapifera</i> Metzg.	Turnip	Navet	Herbstrübe, Mairübe	Nabo

The purpose of these guidelines ("Test Guidelines") is to elaborate the principles contained in the General Introduction (document TG/1/3), and its associated TGP documents, into detailed practical guidance for the harmonized examination of distinctness, uniformity and stability (DUS) and, in particular, to identify appropriate characteristics for the examination of DUS and production of harmonized variety descriptions.

ASSOCIATED DOCUMENTS

These Test Guidelines should be read in conjunction with the General Introduction and its associated TGP documents.

Other associated UPOV documents: TG 185 Turnip rape

These names were correct at the time of the introduction of these Test Guidelines but may be revised or updated. [Readers are advised to consult the UPOV Code, which can be found on the UPOV Website (www.upov.int), for the latest information.]

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1. <u>Subject of these Test Guidelines</u>

These Test Guidelines apply to all varieties of Brassica rapa L. var. rapa (L.) Thell.

2. <u>Material Required</u>

- 2.1 The competent authorities decide on the quantity and quality of the plant material required for testing the variety and when and where it is to be delivered. Applicants submitting material from a State other than that in which the testing takes place must ensure that all customs formalities and phytosanitary requirements are complied with.
- 2.2 The material is to be supplied in the form of seed.
- 2.3 The minimum quantity of plant material, to be supplied by the applicant, should be:

20 g or 10,000 seeds

The seed should meet the minimum requirements for germination, species and analytical purity, health and moisture content, specified by the competent authority. In cases where the seed is to be stored, the germination capacity should be as high as possible and should, be stated by the applicant.

- 2.4 The plant material supplied should be visibly healthy, not lacking in vigor, nor affected by any important pest or disease.
- 2.5 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If it has been treated, full details of the treatment must be given.
- 3. <u>Method of Examination</u>
- 3.1 Number of Growing Cycles
- 3.1.1 The minimum duration of tests should normally be two independent growing cycles.
- 3.1.2 The two independent growing cycles should be in the form of two separate plantings.
- 3.2 Testing Place

Tests are normally conducted at one place. In the case of tests conducted at more than one place, guidance is provided in TGP/9 "Examining Distinctness".

3.3 Conditions for Conducting the Examination

The tests should be carried out under conditions ensuring satisfactory growth for the expression of the relevant characteristics of the variety and for the conduct of the examination.

- 3.4 Test Design
- 3.4.1 Each test should be designed to result in a total of at least 60 Plants, which should be divided between at least 2 replicates.
- 3.4.2 The design of the tests should be such that plants or parts of plants may be removed for measurement or counting without prejudice to the observations which must be made up to the end of the growing cycle.
- 3.5 Additional Tests

Additional tests, for examining relevant characteristics, may be established.

4. Assessment of Distinctness, Uniformity and Stability

4.1 Distinctness

4.1.1 General Recommendations

It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding distinctness. However, the following points are provided for elaboration or emphasis in these Test Guidelines.

4.1.2 Consistent Differences

The differences observed between varieties may be so clear that more than one growing cycle is not necessary. In addition, in some circumstances, the influence of the environment is not such that more than a single growing cycle is required to provide assurance that the differences observed between varieties are sufficiently consistent. One means of ensuring that a difference in a characteristic, observed in a growing trial, is sufficiently consistent is to examine the characteristic in at least two independent growing cycles.

4.1.3 Clear Differences

Determining whether a difference between two varieties is clear depends on many factors, and should consider, in particular, the type of expression of the characteristic being examined, i.e. whether it is expressed in a qualitative, quantitative, or pseudo-qualitative manner. Therefore, it is important that users of these Test Guidelines are familiar with the recommendations contained in the General Introduction prior to making decisions regarding distinctness.

4.1.4 Number of plants or parts of plants to be Examined

Unless otherwise indicated, for the purposes of distinctness, all observations on single plants should be made on 40 plants or parts of plants taken from each of 40 plants and any other observations made on all plants in the test, disregarding any off-type plants.

4.1.5 Method of Observation

The recommended method of observing the characteristic for the purposes of distinctness is indicated by the following key in the second column of the Table of Characteristics (see document TGP/9 "Examining Distinctness", Section 4 "Observation of characteristics"):

MG: single measurement of a group of plants or parts of plants

MS: measurement of a number of individual plants or parts of plants

VG: visual assessment by a single observation of a group of plants or parts of plants

VS: visual assessment by observation of individual plants or parts of plants

Type of observation: visual (V) or measurement (M)

"Visual" observation (V) is an observation made on the basis of the expert's judgment. For the purposes of this document, "visual" observation refers to the sensory observations of the experts and, therefore, also includes smell, taste and touch. Visual observation includes observations where the expert uses reference points (e.g. diagrams, example varieties, side-by-side comparison) or non-linear charts (e.g. color charts). Measurement (M) is an objective observation against a calibrated, linear scale e.g. using a ruler, weighing scales, colorimeter, dates, counts, etc.

Type of record: for a group of plants (G) or for single, individual plants (S)

For the purposes of distinctness, observations may be recorded as a single record for a group of plants or parts of plants (G), or may be recorded as records for a number of single, individual plants or parts of plants (S). In most cases, "G" provides a single record per variety and it is not possible or necessary to apply statistical methods in a plant-by-plant analysis for the assessment of distinctness.

In cases where more than one method of observing the characteristic is indicated in the Table of Characteristics (e.g. VG/MG), guidance on selecting an appropriate method is provided in document TGP/9, Section 4.2.

4.2 Uniformity

- 4.2.1 It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding uniformity. However, the following points are provided for elaboration or emphasis in these Test Guidelines:
- 4.2.2 The assessment of uniformity for open-pollinated varieties should be according to the recommendations for cross-pollinated varieties in the General Introduction.
- 4.2.3 The assessment of uniformity for hybrid varieties depends on the type of hybrid and should be according to the recommendations for hybrid varieties in the General Introduction.
- 4.2.4 For the assessment of uniformity of hybrid varieties, a population standard of 1% and an acceptance probability of at least 95% should be applied. In the case of a sample size of 60 plants, 2 off-types are allowed.

4.3 Stability

- 4.3.1 In practice, it is not usual to perform tests of stability that produce results as certain as those of the testing of distinctness and uniformity. However, experience has demonstrated that, for many types of variety, when a variety has been shown to be uniform, it can also be considered to be stable.
- 4.3.2 Where appropriate, or in cases of doubt, stability may be further examined by testing a new seed stock to ensure that it exhibits the same characteristics as those shown by the initial material supplied.

5. <u>Grouping of Varieties and Organization of the Growing Trial</u>

- 5.1 The selection of varieties of common knowledge to be grown in the trial with the candidate varieties and the way in which these varieties are divided into groups to facilitate the assessment of distinctness are aided by the use of grouping characteristics.
- 5.2 Grouping characteristics are those in which the documented states of expression, even where produced at different locations, can be used, either individually or in combination with other such characteristics: (a) to select varieties of common knowledge that can be excluded from the growing trial used for examination of distinctness; and (b) to organize the growing trial so that similar varieties are grouped together.
- 5.3 The following have been agreed as useful grouping characteristics:
 - (a) Ploidy (characteristic 1)
 - (b) Petiole : anthocyanin coloration (characteristic 2)
 - (c) Leaf: type (characteristic 6)
 - (d) Swollen root (characteristic 16)
 - (e) Root: color of skin above soil (characteristic 18)
 - (f) Root: color of skin below soil (characteristic 20)
 - (g) Root: color of flesh (characteristic 21)
 - (h) Root: shape in longitudinal section (characteristic 23)
- 5.4 Guidance for the use of grouping characteristics, in the process of examining distinctness, is provided through the General Introduction and document TGP/9 "Examining Distinctness".

6. Introduction to the Table of Characteristics

- 6.1 Categories of Characteristics
- 6.1.1 Standard Test Guidelines Characteristics

Standard Test Guidelines characteristics are those which are approved by UPOV for examination of DUS and from which members of the Union can select those suitable for their particular circumstances.

6.1.2 Asterisked Characteristics

Asterisked characteristics (denoted by *) are those included in the Test Guidelines which are important for the international harmonization of variety descriptions and should always be examined for DUS and included in the variety description by all members of the Union, except when the state of expression of a preceding characteristic or regional environmental conditions render this inappropriate.

- 6.2 States of Expression and Corresponding Notes
- 6.2.1 States of expression are given for each characteristic to define the characteristic and to harmonize descriptions. Each state of expression is allocated a corresponding numerical note for ease of recording of data and for the production and exchange of the description.
- 6.2.2 In the case of qualitative and pseudo-qualitative characteristics (see Chapter 6.3), all relevant states of expression are presented in the characteristic. However, in the case of quantitative characteristics with 5 or more states, an abbreviated scale may be used to minimize the size of the Table of Characteristics. For example, in the case of a quantitative characteristic with 9 states, the presentation of states of expression in the Test Guidelines may be abbreviated as follows:

State	Note
small	3
medium	5
large	7

However, it should be noted that all of the following 9 states of expression exist to describe varieties and should be used as appropriate:

State	Note
very small	1
very small to small	2
small	3
small to medium	4
medium	5
medium to large	6
large	7
large to very large	8
very large	9

- 6.2.3 Further explanation of the presentation of states of expression and notes is provided in document TGP/7 "Development of Test Guidelines".
- 6.3 Types of Expression

An explanation of the types of expression of characteristics (qualitative, quantitative and pseudoqualitative) is provided in the General Introduction.

6.4 Example Varieties

Where appropriate, example varieties are provided to clarify the states of expression of each characteristic.

6.5 Legend

	English		français		deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
1 2	3	4	5	6	7			
	Name of characteristics in English		Nom o caract frança	tère en	Name des Merkmals auf Deutsch	Nombre del carácter en español		
	states of expression		types	d'expression	Ausprägungsstufen	tipos de expresión		

1 Characteristic number

2	(*)	Asterisked characteristic	- see Chapter 6.1.2
3	Type of expression QL QN PQ	Qualitative characteristic Quantitative characteristic Pseudo-qualitative characteristic	– see Chapter 6.3 – see Chapter 6.3 – see Chapter 6.3
4	Method of observation (and type MG, MS, VG, VS	e of plot, if applicable)	– see Chapter 4.1.5
5	(+)	See Explanations on the Table of	f Characteristics in Chapter 8.1

- 6 Not applicable
- 7 Growth stage key See Explanations on the Table of Characteristics in Chapter 8

7. <u>Table of Characteristics/Tableau des caractères/Merkmalstabelle/Tabla de caracteres</u>

			English	English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
1.	(*)	QL	vs	(+)		12-70	·			
		Ploidy	/							
		diploid	1					Milan White	2	
		tetrapl	oid					Taronda	4	
2.	(*)	QL	VG			30-90	-			
		Petiol colora	e : anthocyanin ation		bigmentation cyanique	Trieb: Anthocyanfärbung	Tallo: pigmentación antociánica			
		absen	t	absent	е	fehlend	ausente	De Nancy à feuille entière	1	
		preser	nt	présen	te	vorhanden	presente	Scarlet Queen Red Stem, Blanc globe à collet violet, Hinona, Onobeni	9	
3.		QN	VG	(+)		70-130	·			
	•	Leaf:	attitude	Feuille	: port du pétiole	Blatt: Haltung des Stieles	Hoja: porte del peciolo			
		erect		dressé		aufrecht	erecto	Samson, Hinona	1	
		semi-e	erect	demi-d	ressé	halbaufrecht	semierecto	Agressa	3	
		horizo	ntal	horizor	ntal	waagerecht	horizontal	Teltower Kleine	5	
4.		QN	VG	(+)		100-130				
			degree of ving of the top							
		absen	t or very weak	1					1	
		weak		1				Fuku Komachi	3	
		mediu	m					Delilah	5	
		strong						Noir long	7	
		very s	trong						9	

		English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
5. (*)	QN	VG			100-130			1
	Leaf: green	intensity of color		;				
	very li	ght						1
	light						Leielander	3
	mediu	ım					Civasto R	5
	dark						Frisia	7
	very d	lark					Aberdeen Green Top Yellow	9
6. (*)	QL VG		(+)		100-130			
	Leaf:	type	Feuille	e: type	Blatt: Typ	Hoja: tipo		
	entire						Polybra	1
	lobed						Samson	2
	QN	MS/VG	(+)		100-130			-
	Lobed only: lobes	d-leaf varieties Leaf: number of						
	few						Токуо Тор	3
	mediu	ım					De Montesson	5
	many						Aberdeen Green Top Yellow	7
8.	QN	VG	(+)		100-130			1
	Entire only : incisi	e-leaf varieties Leaf : depth of ons of margin						
	very s	hallow						1
	shallo	w					Milan White	3
	mediu	Im					Delilah	5
	deep						Tokyo Market	7
	very d	leep					Polybra	9

		English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota	
9.	QN	VG			100-130				
:	Leaf: marg	undulation of in	Feuille : ondulation du bord			Hoja: ondulación del borde			
	abser	nt or very weak					Tokyo Cross	1	
	weak						Токуо Тор	3	
	mediu	medium					Frisia	5	
	strong							7	
	very s	strong					Imperial Green Globe	9	
10.	QN VG		(+)		100-130				
		dentation of in on the distal	Feuille bord	e: denture du	Blatt: Randzähnung	Hoja: dentado del borde			
	absent or very weak							1	
	weak						Milan White	3	
	mediu	ım					Polybra	5	
	strong)					Taronda	7	
	very s	strong					Appin	9	
11. (*)	QN	MS/VG	(+)		100-130				
	Leaf:	length	Feuille	e: longueur	Blatt: Länge	Hoja: longitud			
	short						Milan White	3	
	mediu	ım					Tokyo Cross	5	
	long						Tyfon	7	
12.	QN	MS/VG	(+)		100-130				
	Leaf:	width	Feuille	e : largeur	Blatt: Breite	Hoja: anchura			
	narrov	N					De Milan rouge extra hâtif à chassis	3	
	mediu	ım					Barkant	5	
	broad						Tyfon	7	

		English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
13.	QN	MS/VG	(+)		100-130			
:	only	<u>d-leaf varieties</u> : Leaf: length of inal lobe						
	short						Platte Witte Mei	3
	medium					Snowball	5	
	long						D'Auvergne hâtive	7
14.	QN MS/VG		(+)		100-130			
	only	d-leaf varieties : Leaf: width of nal lobe	Feuil foliol	le : largeur de la e terminale	Blatt: Breite der Endfieder	Hoja: anchura del foliolo terminal		
	narro	w					Platte Witte Mei	3
		medium					Civasto R	5
	broad						Massif	7
15.	QN	VG			100-130			
		hairiness of r side						
	abser	nt or very weak					Appin	1
	weak						Tokyo Market	3
	mediu	um					De Milan rouge extra hâtif à chassis	5
	stron	g					Blanc dur d'hiver	7
		strong						9
16. (*)	QL	VG			200			
	Swol	len root						
	weak	ly expressed						1
	stron	gly expressed					Noir long, De Croissy, Jaune boule d'or, Blanc globe à collet violet	9

		English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
17. (*)	QN	VG	(+)		260-290			
	Root	position in soil	Racine le sol	e: position dans	Rübe: Sitz im Boden	Raíz: posición en el suelo		
	very s	hallow					Milan White Forcing	1
	shallo	w					Oasis	3
	mediu	m					Agressa	5
	deep						Noir long	7
	very d	eep					Teltower Kleine	9
18. (*)	PQ	VG			240-260			-
	Root: color of skin above soil							
	white						Tokyo Cross	1
	green						Leielander, Rondo	2
	yellow	-orange					Jaune boule d'or	3
	red						Scarlet Queen Red Stem	4
	reddis	h purple					Falko, Hinona	5
	bluish	purple					Barkant	6
	black						Noir long	7
19.	QN	VG			240-260		- -	
		intensity of ation of skin soil		e: intensité de la ir principale de r	Rübe: Intensität der Hauptfarbe des Fleisches	Raíz: intensidad del color principal de la pulpa		
	light		claire		hell	claro		3
	mediu	m	moyen	ne	mittel	medio		5
	dark		foncée		dunkel	oscuro		7

		English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
20. (*)	PQ	VG		240-260			
	Root: below	color of skin soil					
	white					Taronda	1
	yellow					Goldana, Jaune boule d'or	2
	red					Scarlet Queen Red Stem	3
	purple						4
	black					Noir long	5
21. (*)	PQ	VG		240-280			
	Root:	color of flesh					
	white					Agressa	1
	light ye	ellow				Goldana	2
	dark y	ellow				Jaune boule d'or	3
22.	QL	VG		240-280			
	Root: colora	anthocyanin ation of flesh					
	absent	t				Marteau	1
	preser	nt				Scarlet Queen Red Stem	9

		English	fra	ançais	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
23. (*)	PQ	VG			260-280			
	Root: Iongi	shape in tudinal section						
	oblate	e narrow elliptic					Platte Witte Mei	1
		elliptic					Milan White	2
	circula	ar					Rondo	3
	squar	e					Champion Green Top, Yellow	4
	broad	oblong					Delilah, Barkant	5
	narro	w oblong					Long d'Alsace	6
	ovate							7
	narrov	w triangular					Grelos de Santiago	8
	obtria	ngular						9
24. (*)	QN	MS/VG			260-280	·	·	
	Root:	length	Racine: Io	ongueur	Rübe: Länge	Raíz: longitud		
	very s	hort					Milan White	1
	short						The Wallace	3
	mediu	ım					Dynamo	5
	long						Taronda	7
	very le	ong						9
25.	QL	VG	(+)		260-280			
		curvature of al axis						
	abser	nt					Taronda	1
	prese	nt					De Croissy	9

			English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
26. ((*)	QN	VG			260-280			
		<u>Swolle</u> only : wides	en root varieties Root: position of t point						
	·	above	middle					Marteau	1
		at mid	dle					Jaune boule d'or	2
		below	middle					Blanc dur d'hiver	3
27. ((*)	QN	MS/VG	(+)		260-280			
		<u>Swoll</u> only :	<u>en root varieties</u> Root: diameter						
	·	small						Hakutaka	3
	·	mediu	m					Rondo	5
		large						Massif	7
28. ((*)	QN	VG	(+)		260-280	•		-
		<u>Swoll</u> only : collar	<u>en root varieties</u> Root: shape of						
	·	strong	ly depressed						1
	·	depres	ssed					Milan White Forcing	3
		flat						Milan White	5
	ľ	raised						Taronda	7
		strong	ly raised					Agressa	9

		English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
29. (*) PQ	VG			260-280			
	<u>Swo</u> only apex	Ilen root varieties :Root: shape of						
	depr	essed					Milan White Forcing	1
	trunc						Milan White	3
	roun	ded					Frisia	5
		d acute					Kranjska Podolgovata	7
	narro	ow acute					Noir long, Hinona	9
30.	QN	MG/VG			260			
	only	Ilen root varieties :Root: time of est maturity						
	early						Oasis	3
	medi	um						5
	late						Aberdeen Green Top Yellow	7
31.	QN	VG	(+)		310	·	·	
		t : number of uts on the top of oot						
	one	or very few					Taronda	1
	few						Largo de Alsacia	3
	medi	um					Saô Cosme	5
	man	y					Globo blanco de Lugo	7
	verv	many	1				Grelos de Santiago	9

		English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
32.	QN	MG/VG		370			
	Plant flowe	: Time of ring					
	very e	early				Greleiro Temporâo	1
	early					Tyfon, Grelos de Santiago	3
	mediu	ım				Marteau, Globo blanco de Lugo	5
	late					Bola de nieve, Jaune boule d'or	7
	very la	ate				Platte Witte Mei, Golden Ball, Ordes	9
33.	QN	VG		370-400	1		
		er : intensity of w colour of petal					
	weak					Taronda	3
	mediu	ım					5
	strong)				Jaune boule d'or	7

8. Explanations on the Table of Characteristics

8.1 Explanations for individual characteristics

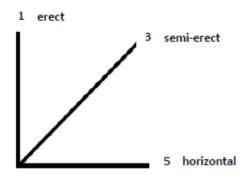
Ad. 1: Ploidy

The ploidy status of the plant can be checked by different methods as determination of the number :

- of chromosomes of the non-thickened root meristem (which is the most reliable method)
- and length of stomata on the lower side of the cotyledon (tetraploid varieties have more and longer stomata than diploid varieties)
- of chloroplasts of the guard cells on the lower side of the cotyledon (the guard cells of tetraploid varieties are bigger and contain more chloroplasts (> 20) than those of diploid varieties (> 10).

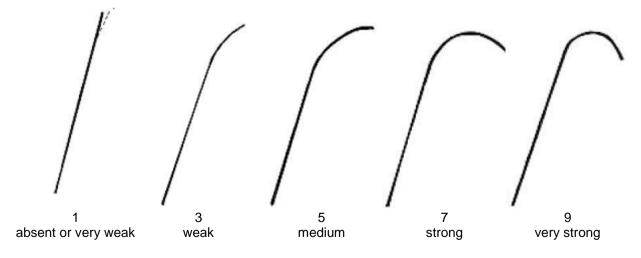
Another efficient method to determine the ploidy status is the flow cytometry (DNA quantification method).

Ad. 3: Leaf: attitude



Ad. 4: Leaf: degree of recurving of the top

The black line represents the profile of the whole leaf.



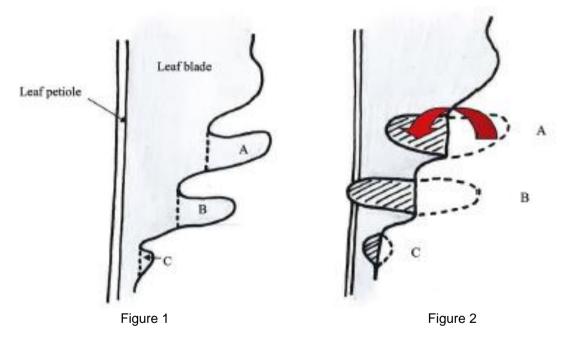
Ad. 6: Leaf: type



Ad. 7: Lobed-leaf varieties only: Leaf: number of lobes

Parts of the leaf blade are considered to be lobes if:

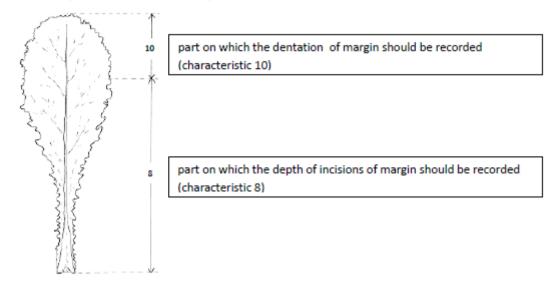
- 1. They have a minimum length of 1 cm and
- 2. When folded back to the midrib as shown in Figs 1 and 2, the folded tissue meets the midrib



- A is not a lobe as it does not meet the midrib when folded
- B is a lobe as it meets the midrib when folded
- C is too small to be a lobe as it is less than 1 cm in length and does not meet the midrib when folded

Ad. 8: Entire-leaf varieties only : Leaf : depth of incisions of margin

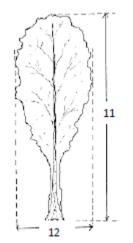
The broadest part of the leaf is the separation between characterics 8 and 10

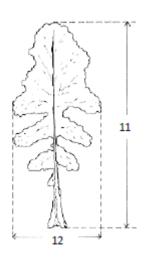


Ad. 10: Leaf: dentation of margin on the distal part

See Ad. 8

Ad. 11: Leaf: length

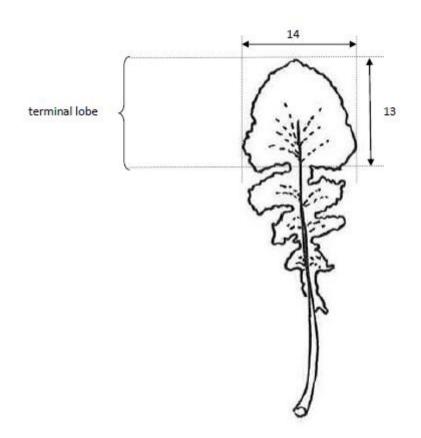




Ad. 12: Leaf: width

See Ad. 11

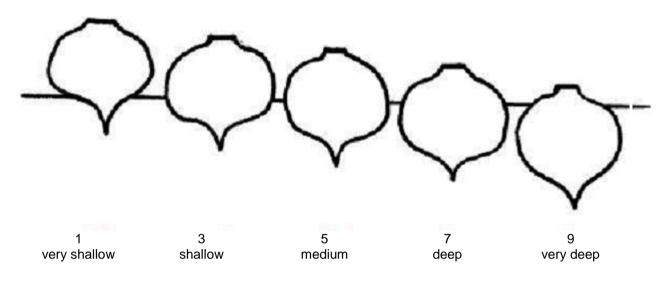
Ad. 13: Lobed-leaf varieties only : Leaf: length of terminal lobe



Ad. 14: Lobed-leaf varieties only : Leaf: width of terminal lobe

See Ad. 13

Ad. 17: Root: position in soil



Ad. 25: Root: curvature of vertical axis

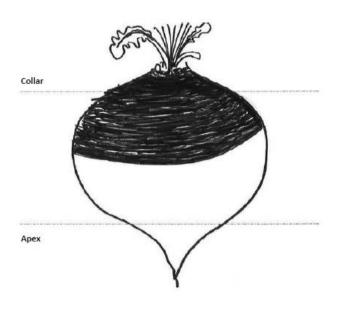
This characteristic refers to the curvature of the vertical axis for roots that are taller than they are wide.



Ad. 27: Swollen root varieties only :Root: diameter

The diameter of the root should be measured the widest point of the root

Ad. 28: Swollen root varieties only :Root: shape of collar



Ad. 31: Plant : number of sprouts on the top of the root



1 one or very few

9 very many

8.2 Key to Growth Stages

- 00 Dry seed
- 1-10 Germination and emergence through soil

Seedling growth

- 12 Elongation of emerging shoot
- 15 Elongation and opening of cotyledons
- 20 Cotyledons fully opened
- 30 Cotyledons fully opened and full development of first true leaf
- 40 Second leaf fully developed
- 50 Third leaf fully developed and initial senescence of cotyledons
- 60 Fourth leaf fully developed and partial senescence of cotyledons
- 70 Fifth leaf fully developed and advanced senescence/drop of cotyledons

Leaf development

- 80 Sixth leaf fully developed
- 90 Seventh leaf fully developed; initial senescence of first true leaf in early cultivars
- 100 Eighth leaf fully developed; 30 % senescence of first true leaf
- 110 Ninth leaf fully developed; 60% senescence of first true leaf
- 120 Tenth leaf fully developed; complete senescence and drop of first true leaf
- 130 Eleventh leaf fully developed.

Root development

- 200 Slight swelling of the root at ground level
- 220 Development of a small swollen root above ground level
- 240 Swollen root increasing in size but not fully developed
- 260 Root fully developed with no cork on skin
- 270 Root fully developed with 40% cork development on skin
- 280 Root fully developed with 80 100% cork development
- 290 Root flesh becoming pithy and fibrous
- 300 Root flesh pithy and fibrous

Flowering and seed production on main stem

- 310 Initial formation and elongation of the flowering stem
- 330 Elongation of the flowering stem with clear space between leaves
- 350 First bud formation and further elongation of stem
- 360 Terminal inflorescence in bud

- 370 Terminal inflorescence with first open flower
- 380 Terminal inflorescence partially flowering
- 400 Terminal inflorescence fully flowering
- 420 Development of siliqua with elongation of flowering stem
- 430 Lowest fully developed siliqua green
- 450 Lowest fully developed siliqua senescing and going brown
- 475 Lowest fully developed siliqua dry with seed beginning to dry
- 500 Lowest fully developed siliqua dry with mature dry seed

9. <u>Literature</u>

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Baltjes, H. J., Klein Geltink, D. J. A., Nienhuis, K. H. and Luesink, B., 1985: Linking Distinctness and Description of Varieties, Journal National Institute Agricultural Botany. 17. p. 9-19.

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Scottish Crop Research Institute, Dundee. Kajanus, B., 1913: Über die Vererbungsweise gewisser Merkmale der Beta- und Brassica-Rüben. II Brassica. Zeitschrift für Pflanzenzüchtung, Band I (4): 419-466.

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McMaster Davey, V., 1931: Color Inheritance in Swedes and Turnips and its Bearing on the Identification of Commercial Stocks. Nat. Journ. Agric. XIV (3): 1-13.

Padilla, G., Cartea, M.E., Rodríguez, V., Ordás, A. 2005: Genetic diversity in a germplasm collection of Brassica rapa subsp. rapa L. from northwestern Spain. Euphytica 145 171-180

10. <u>Technical Questionnaire</u>

TECHN	NICAL Q	UESTIONNAIRE	Page {x} of {y}	Reference Number:
				Application date: (not to be filled in by the applicant)
			ECHNICAL QUESTION	NAIRE on for plant breeders' rights
1.	Subject	of the Technical Question	naire	
	1.1	Botanical name	Brassica rapa L. var. rap	a (L.) Thell.
	1.2	Common name	Turnip	
2.	Applica	nt		
	Name	Γ		
	Addres	s		
	Telepho	one No.		
	Fax No	. [
	E-mail a	address		
	Breede applica	r (if different from nt)		
3.	Propos	ed denomination and breed	er's reference	
	Propos (if avail	ed denomination		
	Breede	r's reference		

NICAL	QUESTIONNAIRE	Page {x} of {y}	Reference Number:	
Inforr	mation on the breeding scheme	and propagation of the va	riety	
4.1	Breeding scheme			
Varie	ety resulting from:			
4.1.1	Crossing			
(a)	controlled cross		[]	
(b)	partially known cross		[]	
(c)	unknown cross		[]	
4.1.2	Mutation		[]	
(plea	se state parent variety)			
4.1.3	Discovery and developmen	t	[]	
	Discovery and developmen se state where and when discovery			
	se state where and when discov			
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TECHNICAL Q	UESTIONNAIRE	Page {x} of {y}	Reference Number	r:
4.2	Method of propagating the	variety		
4.2.1	Seed-propagated varieties			
(a) (b) (c)	Cross-pollination Hybrid Other (please provide detail	ls)		[] [] []
4.2.2	Other (Please provide details)			[]

ТЕСН	NICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:	
5.	Characteristics of the variety to be indi characteristic in Test Guidelines; plea	icated (the number in t se mark the note whic	prackets refers to the corresponding hest corresponds).	
	Characteristics		Example Varieties	Note
5.1 (1)	Ploidy			
	diploid		Milan White	2[]
	tetraploid		Taronda	4[]
5.2 (2)	Petiole : anthocyanin coloration			
	absent		De Nancy à feuille entière	1[]
	present		Blanc globe à collet violet, Hinona, Onobeni, Scarlet Queen Red Stem	9[]
5.3 (5)	Leaf: intensity of green color			
	very light			1[]
	very light to light			2[]
	light		Leielander	3[]
	light to medium			4[]
	medium		Civasto R	5[]
	medium to dark			6[]
	dark		Frisia	7[]
	dark to very dark			8[]
	very dark		Aberdeen Green Top Yellow	9[]
5.4 (6)	Leaf: type			
	entire		Polybra	1[]
	lobed		Samson	2[]
5.5 (16)	Swollen root			
	weakly expressed			1[]
	strongly expressed		Blanc globe à collet violet, De Croissy, Jaune boule d'or, Noir long	9[]

	Characteristics	Example Varieties	Note
5.6 (18)	Root: color of skin above soil		
	white	Tokyo Cross	1[]
	green	Leielander, Rondo	2[]
	yellow-orange	Jaune boule d'or	3[]
	red	Scarlet Queen Red Stem	4[]
	reddish purple	Falko, Hinona	5[]
	bluish purple	Barkant	6[]
	black	Noir long	7[]
5.7 (20)	Root: color of skin below soil		
	white	Taronda	1[]
	yellow	Goldana, Jaune boule d'or	2[]
	red	Scarlet Queen Red Stem	3[]
	purple		4[]
	black	Noir long	5[]
5.8 (21)	Root: color of flesh		
	white	Agressa	1[]
	light yellow	Goldana	2[]
	dark yellow	Jaune boule d'or	3[]
5.9 (23)	Root: shape in longitudinal section		
	oblate narrow elliptic	Platte Witte Mei	1[]
	oblate elliptic	Milan White	2[]
	circular	Rondo	3[]
	square	Champion Green Top, Yellow	4[]
	broad oblong	Barkant, Delilah	5[]
	narrow oblong	Long d'Alsace	6[]
	ovate		7[]
	narrow triangular	Grelos de Santiago	8[]
	obtriangular		9[]

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:			
6. Similar varieties and differences from these varieties Please use the following table and box for comments to provide information on how your candidate variety differs from the variety (or varieties) which, to the best of your knowledge, is (or are) most similar. This information may help the examination authority to conduct its examination of distinctness in a more efficient way.					
Denomination(s) of Characteristic variety(ies) similar to your candidate candidate variety from the simila	variety differs the character	ne expression of Describe the expression of ceristic(s) for the the characteristic(s) for you variety(ies) candidate variety			
Example					
Comments:					

TECH	NICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
#7.	Additional information which may I	help in the examination of th	e variety
7.1	In addition to the information provi help to distinguish the variety?	ded in sections 5 and 6, are	there any additional characteristics which may
	Yes []	No	[]
	(If yes, please provide details)		
7.2	Are there any special conditions f	or growing the variety or cor	ducting the examination?
	Yes []	No	[]
	(If yes, please provide details)		
7.3	Other information		

TECHNICAL QUESTIONNAIRE Page {x} of {y} Reference Number: 8. Authorization for release (a) Does the variety require prior authorization for release under legislation concerning the protection of the environment, human and animal health? Yes [] No [] (b) Has such authorization been obtained? Yes [] No [] If the answer to (b) is yes, please attach a copy of the authorization. 9. Information on plant material to be examined or submitted for examination 9.1 The expression of a characteristic or several characteristics of a variety may be affected by factors, such as pests and disease, chemical treatment (e.g. growth retardants or pesticides), effects of tissue culture, different rootstocks, scions taken from different growth phases of a tree, etc. 9.2 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety unless the competent authorities allow or request such treatment, full details of the treatment must be given. In this respect, please indicate below, to the best of your knowledge, if the plant material to be examined has been subjected to: (a) Microorganisms (e.g. virus, bacteria, phytoplasma) Yes [] No [] (b) Chemical treatment (e.g. growth retardant, pesticide) Yes [] No [] (b) Chemical treatment (e.g. growth retardant, pesticide) Yes [] No [] (c) Tissue culture Yes [] No [] No []											
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(c) Tissue culture Yes [] No [] (d) Other factors Yes [] No [] Please provide details for where you have indicated "yes".		(a)	Microorganisms	anisms (e.g. virus, bacteria, phytoplasma)				Yes []	No []	
(d) Other factors Yes [] No [] Please provide details for where you have indicated "yes". 10. I hereby declare that, to the best of my knowledge, the information provided in this form is correct: Applicant's name		(b)	Chemical treatme	Chemical treatment (e.g. growth retardant, pesticide)					No []	
Please provide details for where you have indicated "yes". 10. I hereby declare that, to the best of my knowledge, the information provided in this form is correct: Applicant's name		(c)	Tissue culture	Tissue culture					No []	
10. I hereby declare that, to the best of my knowledge, the information provided in this form is correct: Applicant's name		(d)	Other factors	Yes []	No []					
Applicant's name		Please provide details for where you have indicated "yes".									
Applicant's name											
Applicant's name											
	10.	10. I hereby declare that, to the best of my knowledge, the information provided in this form is correct:									
Signature		Арр	olicant's name								
Signature Date											
	Signature				Date						

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