

UPOV

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INTERNATIONAL UNION FOR THE PROTECTION OF NEW VARIETIES OF PLANTS

GENEVA

DRAFT

RUBBER

UPOV Code: HEVEA

Hevea Aubl.

GUIDELINES

FOR THE CONDUCT OF TESTS

FOR DISTINCTNESS, UNIFORMITY AND STABILITY

prepared by experts from Brazil and New Zealand

*to be considered by the
Technical Working Party for Ornamental Plants and Forest Trees
at its thirty-ninth session, to be held in Fortaleza, Ceará State, Brazil,
from August 28 to September 1, 2006*

Alternative Names: *

<i>Botanical name</i>	<i>English</i>	<i>French</i>	<i>German</i>	<i>Spanish</i>
<i>Hevea</i> Aubl.	Rubber

The purpose of these guidelines ("Test Guidelines") is to elaborate the principles contained in the General Introduction (document TG/1/3), and its associated TGP documents, into detailed practical guidance for the harmonized examination of distinctness, uniformity and stability (DUS) and, in particular, to identify appropriate characteristics for the examination of DUS and production of harmonized variety descriptions.

ASSOCIATED DOCUMENTS

These Test Guidelines should be read in conjunction with the General Introduction and its associated TGP documents.

* These names were correct at the time of the introduction of these Test Guidelines but may be revised or updated. [Readers are advised to consult the UPOV Code, which can be found on the UPOV Website (www.upov.int), for the latest information.]

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1. Subject of these Test Guidelines

These Test Guidelines apply to all propagative varieties of *Hevea* Aubl.

2. Material Required

2.1 The competent authorities decide on the quantity and quality of the plant material required for testing the variety and when and where it is to be delivered. Applicants submitting material from a State other than that in which the testing takes place must ensure that all customs formalities and phytosanitary requirements are complied with.

2.2 The material is to be supplied in the form of unshooted plants.

2.3 The minimum quantity of plant material, to be supplied by the applicant, should be:

10 plants.

2.4 The plant material supplied should be visibly healthy, not lacking in vigor, nor affected by any important pest or disease.

2.5 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If it has been treated, full details of the treatment must be given.

3. Method of Examination

3.1 *Number of Growing Cycles*

The minimum duration of tests should normally be six years.

3.2 *Testing Place*

Tests are normally conducted at one place. In the case of tests conducted at more than one place, guidance is provided in TGP/9 "Examining Distinctness".

3.3 *Conditions for Conducting the Examination*

3.3.1 The tests should be carried out under conditions ensuring satisfactory growth for the expression of the relevant characteristics of the variety and for the conduct of the examination.

3.3.2 The optimum stage of development for the assessment of each characteristic is indicated by a letter (a or b) in the second column of the Table of Characteristics. The stages of development denoted by each letter are described at the end of Chapter 8.

3.3.3 The recommended method of observing the characteristic is indicated by the following key in the second column of the Table of Characteristics:

MS: measurement of a number of individual plants or parts of plants

VG: visual assessment by a single observation of a group of plants or parts of plants

VS: visual assessment by observation of individual plants or parts of plants

3.3.4. The design consist of a row with spacement of 5 meters between plants.

3.3.5. Because daylight varies, color determinations made against a color chart should be made either in a suitable cabinet providing artificial daylight or in the middle of the day in a room without direct sunlight. The spectral distribution of the illuminant for artificial daylight should conform with the CIE Standard of Preferred Daylight D 6500 and should fall within the tolerances set out in the British Standard 950, Part I. These determinations should be made with the plant part placed against a white background.

3.4 *Test Design*

3.4.1. Each test should be designed to result in a total of at least 5 spaced plants.

3.4.2. The design of the tests should be such that plants or parts of plants may be removed for measurement or counting without prejudice to the observations which must be made up to the end of the growing cycle.

3.5 *Number of Plants / Parts of Plants to be Examined*

3.5.1. Unless otherwise indicated, all observations should be made on 5 plants or parts taken from each of 5 plants. In the case of parts of plants, the number to be taken from each of the plants should be 3.

3.6 *Additional Tests*

3.6.1. Additional tests, for examining relevant characteristics, may be established.

4. Assessment of Distinctness, Uniformity and Stability

4.1 *Distinctness*

4.1.1 General Recommendations

It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding distinctness. However, the following points are provided for elaboration or emphasis in these Test Guidelines.

4.1.2 Consistent Differences

The differences observed between varieties may be so clear that more than one growing cycle is not necessary. In addition, in some circumstances, the influence of the environment is not such that more than a single growing cycle is required to provide assurance that the differences observed between varieties are sufficiently consistent. One

means of ensuring that a difference in a characteristic, observed in a growing trial, is sufficiently consistent is to examine the characteristic in at least two independent growing cycles.

4.1.3 Clear Differences

Determining whether a difference between two varieties is clear depends on many factors, and should consider, in particular, the type of expression of the characteristic being examined, i.e. whether it is expressed in a qualitative, quantitative, or pseudo-qualitative manner. Therefore, it is important that users of these Test Guidelines are familiar with the recommendations contained in the General Introduction prior to making decisions regarding distinctness.

4.2 *Uniformity*

4.2.1. It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding uniformity. However, the following points are provided for elaboration or emphasis in these Test Guidelines:

4.2.2. For the assessment of uniformity of vegetatively propagated varieties, a population standard of 95% and an acceptance probability of at least 1% should be applied. In the case of a sample size of 5 plants, no off-types are allowed.

4.3 *Stability*

4.3.1 In practice, it is not usual to perform tests of stability that produce results as certain as those of the testing of distinctness and uniformity. However, experience has demonstrated that, for many types of variety, when a variety has been shown to be uniform, it can also be considered to be stable.

4.3.2 Where appropriate, or in cases of doubt, stability may be tested, either by growing a further generation, or by testing a new plant stock to ensure that it exhibits the same characteristics as those shown by the previous material supplied.

5. Grouping of Varieties and Organization of the Growing Trial

5.1 The selection of varieties of common knowledge to be grown in the trial with the candidate varieties and the way in which these varieties are divided into groups to facilitate the assessment of distinctness are aided by the use of grouping characteristics.

5.2 Grouping characteristics are those in which the documented states of expression, even where produced at different locations, can be used, either individually or in combination with other such characteristics: (a) to select varieties of common knowledge that can be excluded from the growing trial used for examination of distinctness; and (b) to organize the growing trial so that similar varieties are grouped together.

5.3 The following have been agreed as useful grouping characteristics:



5.4 Guidance for the use of grouping characteristics, in the process of examining distinctness, is provided through the General Introduction.

6. Introduction to the Table of Characteristics

6.1 *Categories of Characteristics*

6.1.1 Standard Test Guidelines Characteristics

Standard Test Guidelines characteristics are those which are approved by UPOV for examination of DUS and from which members of the Union can select those suitable for their particular circumstances.

6.1.2 Asterisked Characteristics

Asterisked characteristics (denoted by *) are those included in the Test Guidelines which are important for the international harmonization of variety descriptions and should always be examined for DUS and included in the variety description by all members of the Union, except when the state of expression of a preceding characteristic or regional environmental conditions render this inappropriate.

6.2 *States of Expression and Corresponding Notes*

States of expression are given for each characteristic to define the characteristic and to harmonize descriptions. Each state of expression is allocated a corresponding numerical note for ease of recording of data and for the production and exchange of the description.

6.3 *Types of Expression*

An explanation of the types of expression of characteristics (qualitative, quantitative and pseudo-qualitative) is provided in the General Introduction.

6.4 *Example Varieties*

Where appropriate, example varieties are provided to clarify the states of expression of each characteristic.

6.5 *Legend*

(*) Asterisked characteristic – see Chapter 6.1.2

QL: Qualitative characteristic – see Chapter 6.3

QN: Quantitative characteristic – see Chapter 6.3

PQ: Pseudo-qualitative characteristic – see Chapter 6.3

MS: measurement of a number of individual plants or parts of plants – see Chapter 3.3.1
VG: visual assessment by a single observation of a group of plants or parts of plants – Chapter 3.3.1
VS: visual assessment by observation of individual plants or parts of plants” –see Chapter 3.3.1

(a)-(c) See Explanations on the Table of Characteristics in Chapter 8.1

(+) See Explanations on the Table of Characteristics in Chapter 8.2

7. Table of Characteristics/Tableau des caractères/Merkmalstabelle/Tabla de caracteres

	English	français	deutsch	español	Example Varieties/ Exemples/ Beispielssorten/ Variedades ejemplo	Note/ Nota	
1.	VS	Leaf cluster : arrangement					
(*)							
(+)							
QL	(a)	type 1			RRIC 102, RRIM 600, PB 235	1	
		type 2			TP 749, IAN 717	2	
		type 3			RRIC 100	3	
		type 4			GT1	4	
2.	VS	Leaflet blade: attitude					
(+)							
QL	(a)	semi-erect				3	
	(c)	horizontal			RRIC 100	5	
		semi-drooping				7	
3.	VS	Leaflet blade: length					
QN	(a)	short				3	
	(c)	medium				5	
		long				7	
4.	VS	Leaflet blade: shape					
(*)							
(+)							
QL	(a)	lanceolate			RRIC 102	1	
	(c)	elliptic			BPM 1	2	
		obovate			GT1	3	
5.	VS	Leaflet blade: symmetry					
(*)							
QL	(a)	symmetric			GT1	1	
	(c)	asymmetric			PB 260	2	

	English	français	deutsch	español	Example Varieties/ Exemples/ Beispielssorten/ Variedades ejemplo	Note/ Nota
6.	VS	Leaflet blade: axis in longitudinal section				
(*)						
(+)						
QL	(a)	straight			BPM1	1
	(c)	arched			GT1	2
		sigmoid				3
7.	VG	Leaflet blade: green colour of upper side				
(*)						
PQ	(a)	light			BPM 1, PB 235, RRIM 600	3
		medium			BPM 24	5
		dark			GT1	7
8.	VS	Leaflet blade: glossiness of upper side				
(*)						
QL	(a)	absent				1
		present				2
9.	VS	Leaflet blade: intensity of glossiness of upper side				
(*)						
QN	(a)	weak			BPM 24	3
		medium			GT1, RRIM 600	5
		strong			PA 31	7
10.	VS	Leaflet blade: surface of upper side				
(*)						
QL	(a)	smooth			GT1, PB235, RRIM 600	1
		intermediate				2
		rough			RRIC 101	3

	English	français	deutsch	español	Example Varieties/ Exemples/ Beispielssorten/ Variedades ejemplo	Note/ Nota
11.	VS	Leaflet blade: pubescence on veins on lower side				
QL	(a)	absent			PB 235, RRIM 600	1
	(c)	present			F 4542, RRIC 101	2
12.	VS	Leaflet blade: undulation of margin				
QL	(a)	absent			BPM 24, PB 235, RRIM600	1
		present			RRIC 100, PB 260, GT1	2
13.	VS	Leaflet blade: shape of apex				
QL	(a)	aristate			BPM 1	1
		acuminate			GT1, PB 235	2
		cuspidate				3
		apiculate				4
14.	VS	Petiolule: attitude				
(*)						
(+)						
QL	(a)	semi-erect			RRIM 600	3
		horizontal			GT1, PB 235	5
		semi-drooping				7
15.	VS	Petiole: attitude				
(+)						
(*)						
QL		semi-erect			RRIC 100, GT1, RRIM 600	3
		horizontal			PB 235	5
		semi-drooping				7
16.	VS	Trunk: axis				
(*)						
QL	(b)	straight			GT1, RRIM 600	1
		curved			TP 745	2

	English	français	deutsch	español	Example Varieties/ Exemples/ Beispielssorten/ Variedades ejemplo	Note/ Nota	
17.	MS	Trunk: girth (1 m from the ground)					
(*)	(b)	small				3	
QN		medium			RRIM 600	5	
		large				7	
18.	VS	Trunk: shape in cross section (lower third)					
QL	(b)	circular			PB 235	1	
		elliptic				2	
		fluted				3	
19.	VG	Trunk: predominant colour of bark					
QL	(b)	reddish				1	
		brown				2	
		grey			PB 235	3	
20.	VG	Trunk: texture of bark					
QL	(b)	smooth			PB 235	1	
		rough			GT1	2	
21.	VS	Crown: shape of canopy					
(*)	(b)	circular				1	
(+)		ovate				2	
		elliptic				3	
		obtriangular				4	
		irregular obtriangular				5	

	English	français	deutsch	español	Example Varieties/ Exemples/ Beispielssorten/ Variedades ejemplo	Note/ Nota
22.	VG					
(*)	Crown: density					
QN	(b)	open			PR 261, FDR 5788	3
		medium			PB 260	5
		dense			PB 217	7
23.	VS					
(*)	Coagulum: colour of surface (24 hours after tapping)					
QL	(b)	white			GT1, RRIM 600, PB 217	1
		light yellow			PB260	2
		yellow				3
		dark grey			IAN 3156, RRII 203	4
24.	VG					
	Tree: duration of wintering					
QN	(b)	short			GT1, PB 235, RRIM 600	3
		medium				5
		long			F 4512, FX 3899	7
25.	VG					
	Tree: begin of wintering					
PQ	(b)	early			BPM 1, PB 260	3
		medium			PB 235	5
		late			GT1, RRIM 600	7
26.	MS					
(*)	Seed: length					
(+)						
QN	(b)	short				3
		medium			RRIM 600, GT1	5
		long				7

	English	français	deutsch	español	Example Varieties/ Exemples/ Beispielssorten/ Variedades ejemplo	Note/ Nota
27.	MS					
	Seed: width					
(+)						
(*)						
QN	(b)	narrow			GT1	3
		medium			RRIM 600	5
		broad			RRIC 100	7
28.	MS					
	Seed: thickness					
(*)						
(+)						
QN	(b)	thin			RRIM 600	3
		medium			GT1	5
		thick				7
29.	VG					
	Seed: shape in dorsal view					
(*)						
(+)						
QL	(b)	circular			IRCA 339	1
		ovate			IRCA 621	2
		oblong			FDR 233	3
		square			FDR 18	4

8. Explanations on the Table of Characteristics

8.1 Explanations covering several characteristics

Characteristics containing the following key in the second column of the Table of Characteristics should be examined as indicated below:

- (a) 1 to 2 year-old plants
- (b) 4 to 6 year-old plants
- (c) observation should be made on the central leaflet

8.2 Explanations for individual characteristics

Ad. 1: Leaf cluster: arrangement (excluding terminal cluster)



1 type 1



2 type 2

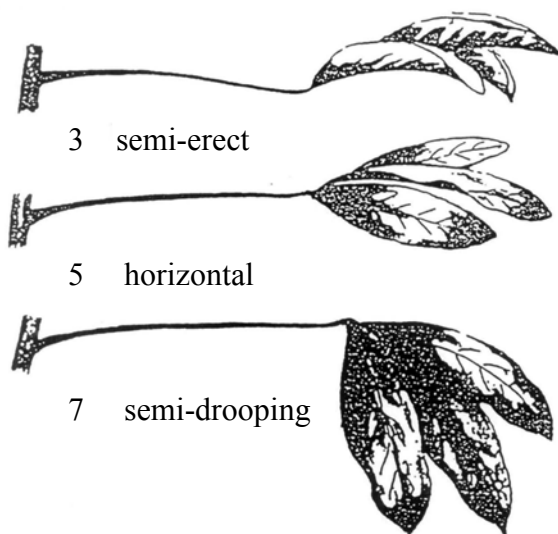


3 type 3

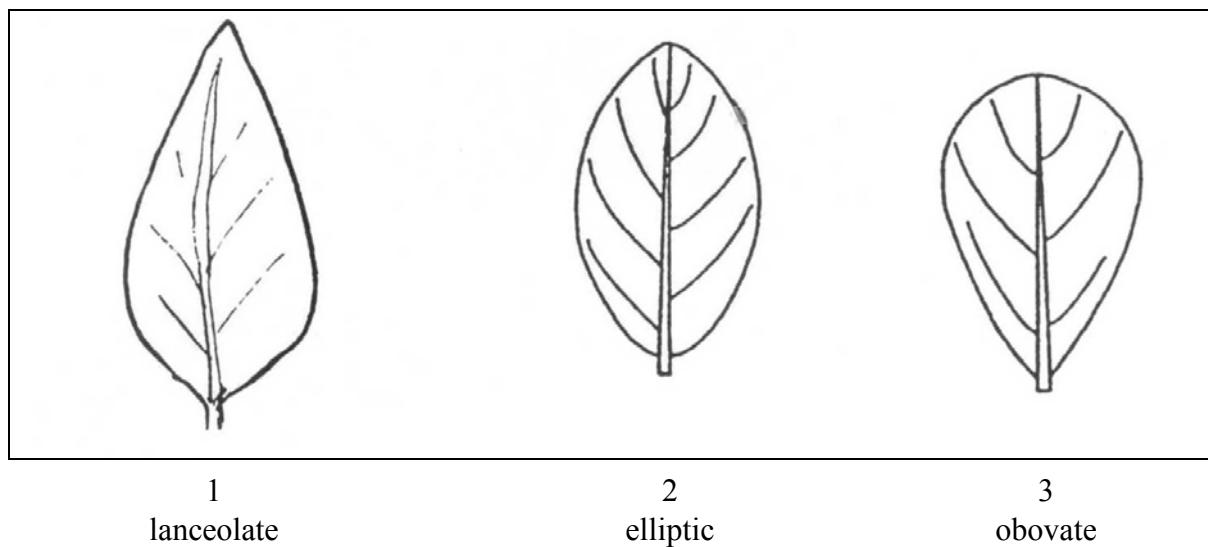


4 type 4

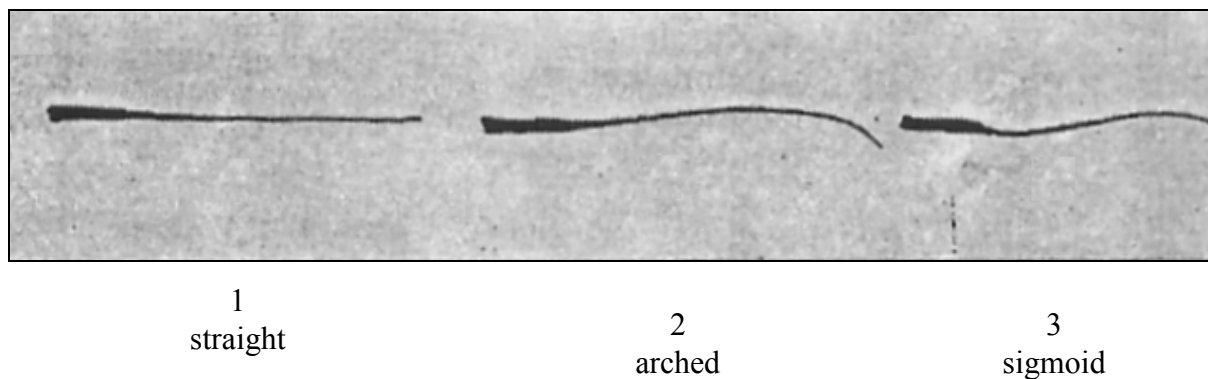
Ad. 2: Leaflet blade: attitude



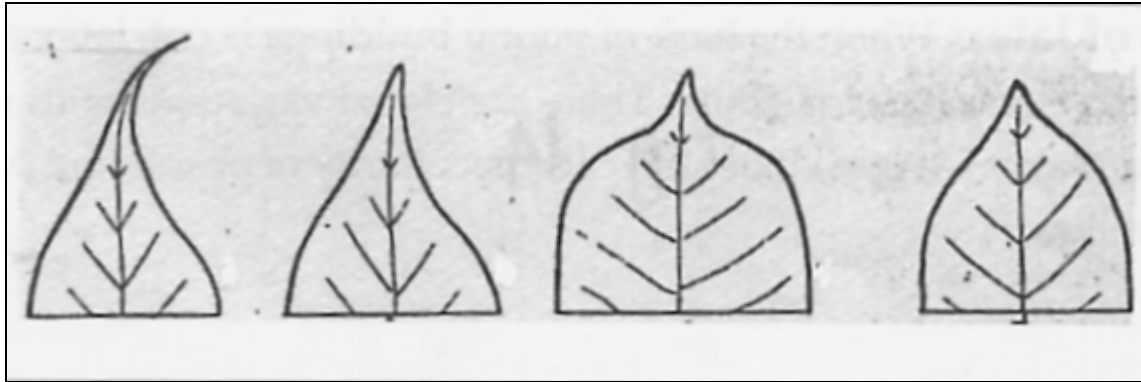
Ad. 4: Leaflet blade: shape



Ad. 6: Leaflet blade: axis in longitudinal section



Ad. 13. Leaflet blade : shape of apex.



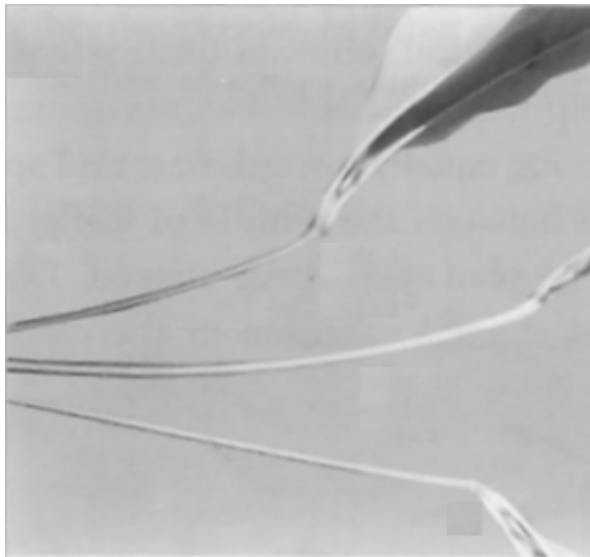
1
aristate

2
acuminate

3
cuspidate

4
apiculate

Ad.14: Petiolule: attitude

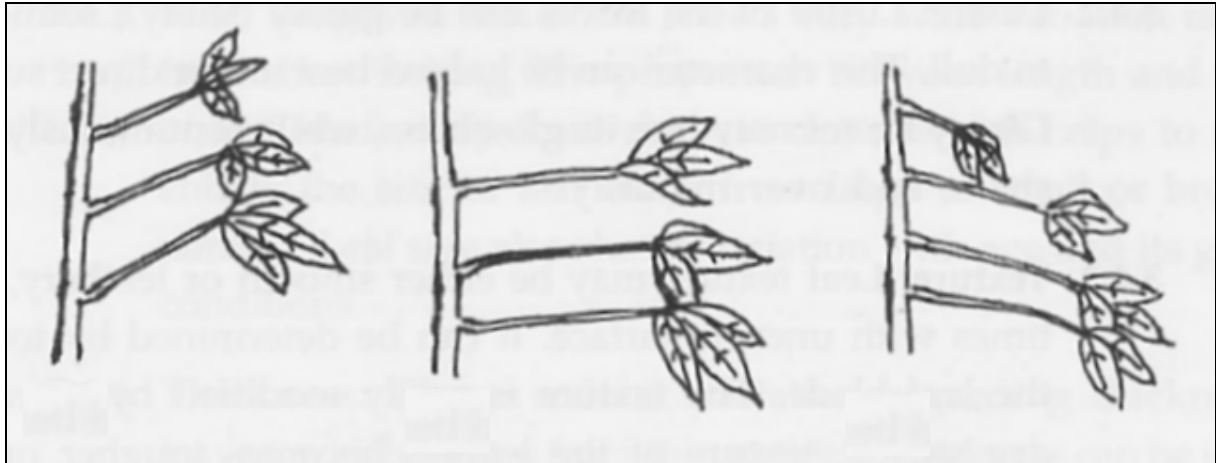


3
semi-erect

5
horizontal

7
semi-drooping

Ad. 15: Petiole : attitude

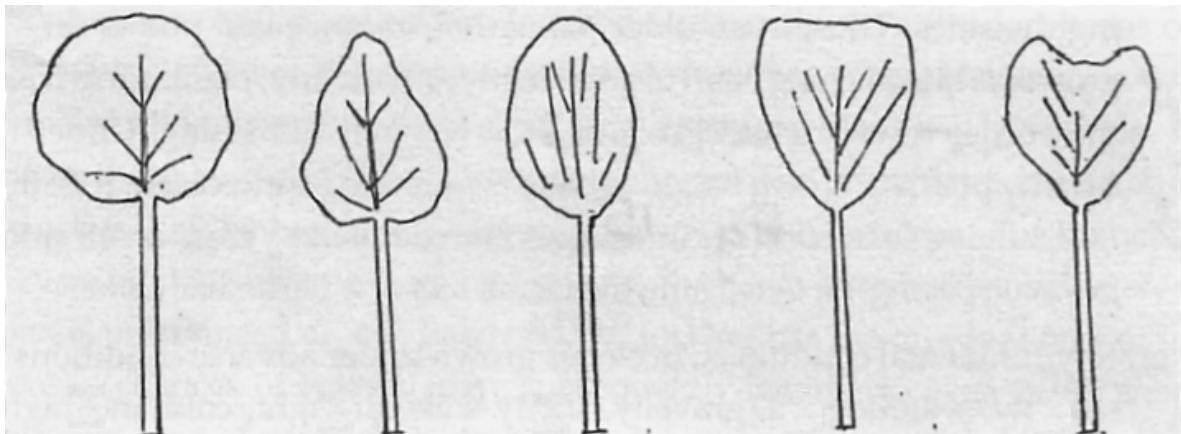


3
semi-erect

5
horizontal

7
semi-drooping

Ad. 21: Crown : shape of canopy



1
circular

2
ovate

3
elliptic

4
obtriangular

5
irregular obtriangular

Ad. 26: Seed: length

- 3 short: less than 20 mm
- 5 medium: between 20 and 30 mm
- 7 long: more than 30 mm

Ad. 27: Seed: width

- 3 narrow: less than 20 mm
- 5 medium: between 20 to 25 mm
- 7 broad: more than 25 mm

Ad. 28: Seed: thickness

- 3 thin: less than 16 mm
- 5 medium: between 16 to 20 mm
- 7 thick: more than 20 mm

Ad. 29: Seed: shape in dorsal view



4
circular



3
ovate



2
oblong



1
square

9. Literature

Besse P., Seguin M., Lebrun P., Chevallier M.H., Nicolas D., Lanaud C. (1994). Genetic diversity among wild and cultivated populations of *Hevea brasiliensis* assessed by nuclear RFLP analysis. *Theoretical and Applied Genetics*, 88 : 199-207.

Bobilioff W (1931) Colour reactions of latex as a mark of identification of *Hevea* clones. *Archives of Rubber Cultivation*, 15 : 289-309.

Chevallier M.H. (1988) Genetic variability of *Hevea brasiliensis* germplasm, using isozyme markers. *Journal of Natural Rubber Research*, 3 : 42-53.

Dijkman M.J. (1939) Identificatiekenmerken van de voornaamste in de praktijk aageplante *Hevea* cloonen. *Buitenzorg Central Proefstations Vereeniging Archipel Drukkerij*. 263p.

Dijkman M.J. (1951) *Hevea*: Thirty years of research in the Far East. University of Miami Press, Florida, 155-177.

Leconte A, Lebrun P., Nicolas D. And Seguin M. (1994). Electrophoresis application to hevea clone identification. *Plantations, Recherche, Development*, 1(2): 34-36.

Mercykutty V.C., Marattukalam J.G., Saraswathyamma C.K. and Meenakumari T. (2002) Identification of *Hevea* clones. A manual. Rubber Research Institute of India. 103p.

Mercykutty V.C. (1998) Clone identification I. *Rubber*, 386 : 21-25.

Mercykutty V.C. (1998) Clone identification II. *Rubber*, 387 : 15-18.

Mercykutty V.C., Varghese Y., Licy J. and Panikkar A.O.N. (1991) Juvenile characters and seed morphology of certain modern *Hevea* clones. *Indian Journal of Natural Rubber Research*, 4(1) : 16-25.

Thomas V., Mercykutty V.C. and Saraswathyamma C.K. (1996) Seed morphology of para rubber tree (*Hevea brasiliensis*, Muell. Arg. Euphorbiaceae) : A review. *Phytomorphology*; 46(4) : 335-342.

10. Technical Questionnaire

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
		Application date: (not to be filled in by the applicant)
TECHNICAL QUESTIONNAIRE to be completed in connection with an application for plant breeders' rights		
1. Subject of the Technical Questionnaire		
1.1 Botanical name	<input type="text" value="{ Botanical name }"/>	
1.2 Common name	<input type="text" value="{ Common name }"/>	
1. Subject of the Technical Questionnaire (please complete):		
1.1 Botanical name:		
1.2 Common name:		
with the boxes left blank for completion by the applicant.		
2. Applicant		
Name	<input type="text"/>	
Address	<input type="text"/>	
Telephone No.	<input type="text"/>	
Fax No.	<input type="text"/>	
E-mail address	<input type="text"/>	
Breeder (if different from applicant)	<input type="text"/>	

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
<p>3. Proposed denomination and breeder's reference</p> <p>Proposed denomination (if available) <input data-bbox="625 405 1350 456" type="text"/></p> <p>Breeder's reference <input data-bbox="625 528 1350 580" type="text"/></p>		

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:						
<p>#4. Information on the breeding scheme and propagation of the variety</p> <p>4.1 Breeding scheme</p> <p>ASW 15</p>								
<p>(a) <i>Alternative 1</i></p>								
<p>“Variety resulting from:</p> <p>“4.1.1 Crossing</p> <table><tbody><tr><td>“(a) controlled cross (please state parent varieties)</td><td>[]</td></tr><tr><td>“(b) partially known cross (please state known parent variety(ies))</td><td>[]</td></tr><tr><td>“(c) unknown cross</td><td>[]</td></tr></tbody></table> <p>“4.1.2 Mutation (please state parent variety) []</p> <p>“4.1.3 Discovery and development (please state where and when discovered and how developed) []</p> <p>“4.1.4 Other (please provide details)” []”</p> <div data-bbox="443 1391 1137 1487" style="border: 1px solid black; height: 43px; width: 435px;"></div>			“(a) controlled cross (please state parent varieties)	[]	“(b) partially known cross (please state known parent variety(ies))	[]	“(c) unknown cross	[]
“(a) controlled cross (please state parent varieties)	[]							
“(b) partially known cross (please state known parent variety(ies))	[]							
“(c) unknown cross	[]							

Authorities may allow certain of this information to be provided in a confidential section of the Technical Questionnaire.

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:						
<p>(b) <i>Alternative 2</i></p> <p>“Variety resulting from:</p> <p>“4.1.1 Crossing</p> <table style="width: 100%; border: none;"> <tr> <td style="padding-left: 20px;">“(a) controlled cross (please state parent varieties)</td> <td style="text-align: right; vertical-align: bottom;">[]</td> </tr> <tr> <td style="padding-left: 20px;">“(b) partially known cross (please state known parent variety(ies))</td> <td style="text-align: right; vertical-align: bottom;">[]</td> </tr> <tr> <td style="padding-left: 20px;">“(c) unknown cross</td> <td style="text-align: right; vertical-align: bottom;">[]</td> </tr> </table> <p>“4.1.2 Discovery and development [] (please state where and when discovered and how developed)</p> <p>“4.1.3 Other []” (please provide details)”</p> <div style="border: 1px solid black; height: 30px; width: 45%; margin: 10px auto;"></div>			“(a) controlled cross (please state parent varieties)	[]	“(b) partially known cross (please state known parent variety(ies))	[]	“(c) unknown cross	[]
“(a) controlled cross (please state parent varieties)	[]							
“(b) partially known cross (please state known parent variety(ies))	[]							
“(c) unknown cross	[]							

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
4.2 Method of propagating the variety		
GN 31		
The examples below indicate how this section can be formatted and some appropriate terms which can be used:		
<i>Example 1</i>		
“4.2.1 Seed-propagated varieties		
“(a)	Self-pollination	[]
“(b)	Cross-pollination	
	(i) population	[]
	(ii) synthetic variety	[]
“(c)	Hybrid	[]
	{...see GN 32 for example...}	
“(d)	Other	[]
	(please provide details)”	
<div style="border: 1px solid black; height: 40px; width: 100%;"></div>		
“4.2.2 Vegetatively propagated varieties		
	{...see Example 2...}	[... ..]
“4.2.3 Other		[]”
	(please provide details)”	

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
<p><i>Example 2</i></p> <p>“4.2.1 Vegetative propagation</p> <p> “(a) cuttings []</p> <p> “(b) <i>in vitro</i> propagation []</p> <p> “(c) other (state method) []</p> <p>“4.2.2 Seed []</p> <p>“4.2.3 Other []” (please provide details)”</p> <div data-bbox="422 875 1177 954" style="border: 1px solid black; height: 35px; width: 473px; margin: 10px auto;"></div>		
<p>GN 32</p> <p>“In the case of hybrid varieties the production scheme for the hybrid should be provided on a separate sheet. This should provide details of all the parent lines required for propagating the hybrid e.g.</p> <p> “<i>Single Hybrid</i></p> <p> “(… female parent …) x (… male parent …)</p> <p> “<i>Three-Way Hybrid</i></p> <p> “(… female line …) x (… male line …)</p> <p> “=> single hybrid used as female parent x (… male parent …)</p> <p>“and should identify in particular:</p> <p> “(a) any male sterile lines</p> <p> “(b) maintenance system of male sterile lines.”</p>		

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:	
<p>5. Characteristics of the variety to be indicated (the number in brackets refers to the corresponding characteristic in Test Guidelines; please mark the note which best corresponds).</p>			
Characteristics	Example Varieties	Note	
<p>GN 13.3 Technical Questionnaire (TQ) characteristics</p> <p>GN 13.4 Relationship between Asterisked, Grouping and TQ characteristics</p>			

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:	
<p>6. Similar varieties and differences from these varieties</p> <p><i>Please use the following table and box for comments to provide information on how your candidate variety differs from the variety (or varieties) which, to the best of your knowledge, is (or are) most similar. This information may help the examination authority to conduct its examination of distinctness in a more efficient way.</i></p>			
Denomination(s) of variety(ies) similar to your candidate variety	Characteristic(s) in which your candidate variety differs from the similar variety(ies)	Describe the expression of the characteristic(s) for the similar variety(ies)	Describe the expression of the characteristic(s) for your candidate variety
<i>Example</i>	<i>[e.g. Flower color]</i>	<i>[e.g. orange]</i>	<i>[e.g. orange red]</i>
<p>Comments:</p>			

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
<p>#7. Additional information which may help in the examination of the variety</p> <p>7.1 In addition to the information provided in sections 5 and 6, are there any additional characteristics which may help to distinguish the variety?</p> <p>Yes [] No []</p> <p>(If yes, please provide details)</p> <p>7.2 Are there any special conditions for growing the variety or conducting the examination?</p> <p>Yes [] No []</p> <p>(If yes, please provide details)</p> <p>7.3 Other information</p>		
<p>8. Authorization for release</p> <p>(a) Does the variety require prior authorization for release under legislation concerning the protection of the environment, human and animal health?</p> <p>Yes [] No []</p> <p>(b) Has such authorization been obtained?</p> <p>Yes [] No []</p> <p>If the answer to (b) is yes, please attach a copy of the authorization.</p>		

Authorities may allow certain of this information to be provided in a confidential section of the Technical Questionnaire.

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:												
<p>9. Information on plant material to be examined or submitted for examination.</p> <p>9.1 The expression of a characteristic or several characteristics of a variety may be affected by factors, such as pests and disease, chemical treatment (e.g. growth retardants or pesticides), effects of tissue culture, different rootstocks, scions taken from different growth phases of a tree, etc.</p> <p>9.2 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If the plant material has undergone such treatment, full details of the treatment must be given. In this respect, please indicate below, to the best of your knowledge, if the plant material to be examined has been subjected to:</p> <table style="width: 100%; border: none;"> <tr> <td style="width: 70%;">(a) Microorganisms (e.g. virus, bacteria, phytoplasma)</td> <td style="width: 15%;">Yes []</td> <td style="width: 15%;">No []</td> </tr> <tr> <td>(b) Chemical treatment (e.g. growth retardant, pesticide)</td> <td>Yes []</td> <td>No []</td> </tr> <tr> <td>(c) Tissue culture</td> <td>Yes []</td> <td>No []</td> </tr> <tr> <td>(d) Other factors</td> <td>Yes []</td> <td>No []</td> </tr> </table> <p>Please provide details for where you have indicated “yes”.</p> <p>.....</p>			(a) Microorganisms (e.g. virus, bacteria, phytoplasma)	Yes []	No []	(b) Chemical treatment (e.g. growth retardant, pesticide)	Yes []	No []	(c) Tissue culture	Yes []	No []	(d) Other factors	Yes []	No []
(a) Microorganisms (e.g. virus, bacteria, phytoplasma)	Yes []	No []												
(b) Chemical treatment (e.g. growth retardant, pesticide)	Yes []	No []												
(c) Tissue culture	Yes []	No []												
(d) Other factors	Yes []	No []												
<p>10. I hereby declare that, to the best of my knowledge, the information provided in this form is correct:</p> <p>Applicant's name <input style="width: 550px; height: 25px;" type="text"/></p> <p>Signature <input style="width: 350px; height: 25px;" type="text"/> Date <input style="width: 180px; height: 25px;" type="text"/></p>														

[Annex follows]

ANNEX

Additional Useful Explanations

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Part I

Introduction

The following Annex contains a list of characteristics derived by using electrophoresis and a description of the method to be used. UPOV decided to place these characteristics in a Annex to establish the Test Guidelines, thereby creating a special category of characteristic, because the majority of the UPOV member States is of the view that it is not possible to establish distinctness solely on the basis of a difference found in a characteristic derived by using electrophoresis. Such characteristics should therefore be used as a complement to other differences in morphological or physiological characteristics. UPOV reconfirms that these characteristics are considered useful but that they might not be sufficient on their own to establish distinctness. They should not be used as a routine characteristic but at the request or with the agreement of the applicant of the candidate variety.

Procedure described hereafter can be particularly carried out according to the specifications made by CIRAD for *Hevea* genus (Leconte et al., 1994, *Electrophoresis application to Hevea clone identification*, Plantations, Recherche, Développement, 2:28-36 – Leconte 1997, *Starch gel electrophoresis for rubber clone identification. Practical handbook*). At its maximum, the analyses of the 13 enzymatic systems can be carried out simultaneously in the same day by using 2 starch gels respectively with pH6.6 and pH8. Analysis can also be focussed only on some of the 13 available enzymatic systems depending on the varieties being analysed.

For each enzymatic system, there are a number of different possible alleles at each locus and the analysis of each isoform is based on the recognition of the alleles from these proteins, which appear on gels as a series of well defined bands or patterns of bands. The alleles are described by band numbers according to the definition given to them by Cirad

Part II

Characteristics Derived by Using Electrophoresis

Characteristics	English	Français	Example Varieties	Note
Caractères			Exemples	
57. Malate dehydrogenase composition (+) Composition de la malate déshydrogenase	<i>band 1</i>	<i>bande 1</i>		1
	<i>band 1 + 2</i>	<i>bande 1 + 2</i>	CD 1078	2
	<i>band 1 + 3</i>	<i>bande 1 + 3</i>	IAN 3087	3
	<i>band 1 + 4</i>	<i>bande 1 + 4</i>	F 4542	4
	band 2	bande 2	GT1, PB 235	5
	band 2 + 3	bande 2 + 3	BPM 24, PB 260	6
	<i>band 2 + 4</i>	<i>bande 2 + 4</i>	FX 3899	7
	<i>band 3</i>	<i>bande 3</i>	PB 217, IRCA 111	8
	<i>band 3 + 4</i>	<i>bande 3 + 4</i>	SCH P 52	9
	<i>band 4</i>	<i>bande 4</i>	F 4506	10
58. Phospho glucose isomerase composition (+) Composition de la phospho glucose isomérase	band 1	bande 1	PB 260, PB 217, PB 260	1
	band 1 + 2	bande 1 + 2	RRIM 519	2
	band 1 + 3	bande 1 + 3	GT1	3
	<i>band 1 + 4</i>	<i>bande 1 + 4</i>	AC 58	4
	<i>band 1 + 5</i>	<i>bande 1 + 5</i>	IRCA 130	5
	<i>band 2</i>	<i>bande 2</i>	RO 51	6
	band 2 + 3	bande 2 + 3	PB 235	7
	<i>band 2 + 4</i>	<i>bande 2 + 4</i>	BPM 22	8
	<i>band 2 + 5</i>	<i>bande 2 + 5</i>	GU 174	9
	band 3	bande 3	PB 314	10
	<i>band 3 + 4</i>	<i>bande 3 + 4</i>	AC 53	11
	<i>band 3 + 5</i>	<i>bande 3 + 5</i>	RRIM 527	12
	<i>band 4</i>	<i>bande 4</i>	F 4542	13
59. Alanine amino peptidase composition (+) Composition de l'alanine amino peptidase	<i>band 1 + 2</i>	<i>bande 1 + 2</i>	RO 53	1
	band 2	bande 2	GT1, PB 217, PB 260, PB 235	2
	<i>band 2 + 3</i>	<i>bande 2 + 3</i>	RO 58	3
	<i>band 2 + 4</i>	<i>bande 2 + 4</i>	RRIC 130	4
	<i>band 3</i>	<i>bande 3</i>	RO 60	5
	<i>band 3 + 4</i>	<i>bande 3 + 4</i>	RO 61	6
	<i>band 4</i>	<i>bande 4</i>	F 4506	7
60. Leucine amino peptidase composition (+) Composition de la leucine amino peptidase	<i>band 1 + 2</i>	<i>bande 1 + 2</i>	FX 3899	1
	<i>band 1 + 6</i>	<i>bande 1 + 6</i>	RO 51	2
	band 2	bande 2	BPM 24, GT1, PB 217, PB 235	3
	<i>band 2 + 3</i>	<i>bande 2 + 3</i>	PB 5/63	4
	band 2 + 4	bande 2 + 4	PB 260	5
	<i>band 2 + 5</i>	<i>bande 2 + 5</i>	RRIC 100	6
	<i>band 2 + 6</i>	<i>bande 2 + 6</i>	RRIC 101	7
	<i>band 2 + 7</i>	<i>bande 2 + 7</i>	WAR 4	8
	<i>band 4</i>	<i>bande 4</i>	PB 312	9
	<i>band 4 + 6</i>	<i>bande 4 + 6</i>	RRIC 132	10
	<i>band 4 + 7</i>	<i>bande 4 + 7</i>	PA 31	11
	<i>band 5</i>	<i>bande 5</i>		12
	<i>band 6</i>	<i>bande 6</i>	MDF 180	13
	<i>band 6 + 7</i>	<i>bande 6 + 7</i>	MDF 372	14
	<i>band 6 + 8</i>	<i>bande 6 + 8</i>	AC 60	15
61. Esterase composition (+) Composition de l'estérase	band 1	bande 1	GT1	1
	<i>band 1 + 2</i>	<i>bande 1 + 2</i>	RO 58	2
	<i>band 1 + 3</i>	<i>bande 1 + 3</i>	AC 57	3
	<i>band 1 + 4</i>	<i>bande 1 + 4</i>	IRCA 621	4
	<i>band 1 + 5</i>	<i>bande 1 + 5</i>	FX 985	5
	band 1 + 6	bande 1 + 6	BPM 24, PB 217, PB 260	6
	<i>band 1 + 7</i>	<i>bande 1 + 7</i>	IAN 6590	7

	<i>band 2 + 4</i>	<i>bande 2 + 4</i>	RO 50	8
	<i>band 3</i>	<i>bande 3</i>	AC 53	9
	<i>band 3 + 4</i>	<i>bande 3 + 4</i>		10
	<i>band 4</i>	<i>bande 4</i>	F 4542	11
	<i>band 4 + 5</i>	<i>bande 4 + 5</i>	PA 31	12
	<i>band 4 + 6</i>	<i>bande 4 + 6</i>	FX 3899	13
	<i>band 5</i>	<i>bande 5</i>	F 4512	14
	<i>band 5 + 6</i>	<i>bande 5 + 6</i>	RRIC 121	15
	band 6	bande 6	PB 235	16
	<i>band 6 + 7</i>	<i>bande 6 + 7</i>	IAN 6587	17
62.	<i>band 1</i>	<i>bande 1</i>	F 4512	1
	<i>band 1 + 2</i>	<i>bande 1 + 2</i>	MDF 114	2
	<i>band 1 + 3</i>	<i>bande 1 + 3</i>	PFB5	3
(+)	band 2	bande 2	IAN717	4
	<i>band 2 + 3</i>	<i>bande 2 + 3</i>	PiLA 44	5
	<i>band 2 + 6</i>	<i>bande 2 + 6</i>	SCH C 133	6
	band 3	bande 3	GT 1, BPM 24, PB 217	7
	<i>band 3 + 4</i>	<i>bande 3 + 4</i>	AC80	8
	<i>band 3 + 5</i>	<i>bande 3 + 5</i>	MDF 362	9
	<i>band 3 + 6</i>	<i>bande 3 + 6</i>	SCH C 814	10
	<i>band 4</i>	<i>bande 4</i>	F 4542	11
	<i>band 5</i>	<i>bande 5</i>	IAN 6158	12
	<i>band 5 + 6</i>	<i>bande 5 + 6</i>	CNSAM 7706	13
63.	<i>band 0</i>	<i>bande 0</i>	P 122	1
	<i>band 0 + 1</i>	<i>bande 0 + 1</i>	PA 31	2
	band 1	bande 1	GT1,BPM24, PB 217, PB 235	3
(+)	band 1 + 2	bande 1 + 2	RRIM 600	4
	<i>band 2</i>	<i>bande 2</i>	PB 235	5
64.	<i>band 2</i>	<i>bande 2</i>	PB 310	1
	band 2 + 3	bande 2 + 3	RRIM 600, PB 235	2
	<i>band 2 + 4</i>	<i>bande 2 + 4</i>	RRIC 132	3
(+)	band 3	bande 3	GT1, PB 217, PB 260	4
	<i>band 3 + 4</i>	<i>bande 3 + 4</i>	FX 3899	5
	<i>band 4</i>	<i>bande 4</i>	F 4542	6
65.	<i>band 1 + 2</i>	<i>bande 1 + 2</i>	IAN717	1
	<i>band 1 + 3</i>	<i>bande 1 + 3</i>	FX 3899	2
	<i>band 2</i>	<i>bande 2</i>	RRIC 103	3
(+)	<i>band 2 + 3</i>	<i>bande 2 + 3</i>	AC 53	4
	<i>band 2 + 4</i>	<i>bande 2 + 4</i>	CNSAM 7701	5
	band 3	bande 3	GT1	6
	band 3 + 4	bande 3 + 4	RRIM 600, PB 260, PB 235	7
	band 4	bande 4	PB 217	8
	<i>band 4 + 5</i>	<i>bande 4 + 5</i>	TU 45/525	9
66.	<i>band 1</i>	<i>bande 1</i>	RRIC 22	1
	band 1 + 2	bande 1 + 2	FX 3899	2
	band 1 + 3	bande 1 + 3	GT1, PR 107	3
(+)	band 2	bande 2	AVROS 2037, PB 217	4
	band 2 + 3	bande 2 + 3	BPM 24, IAN 873	5
	band 3	bande 3	PFB 5	6
67.	<i>band 1 + 2</i>	<i>bande 1 + 2</i>	AC 80	1
	<i>band 2</i>	<i>bande 2</i>	AC 53	2
	<i>band 2 + 3</i>	<i>bande 2 + 3</i>	RO 60	3
(+)	<i>band 2 + 4</i>	<i>bande 2 + 4</i>	IRCA 652	4
	<i>band 3</i>	<i>bande 3</i>	AC 62	5
	<i>band 3 + 4</i>	<i>bande 3 + 4</i>	IAN 710, RRIM 725	6
	<i>band 3 + 5</i>	<i>bande 3 + 5</i>	RRIM 729	7
	band 4	bande 4	RRIM600,GT1,PB260,PB217	8
	<i>band 4 + 5</i>	<i>bande 4 + 5</i>	RRIC 110	9
	<i>band 4 + 6</i>	<i>bande 4 + 6</i>	PUA 8	10
	<i>band 5</i>	<i>bande 5</i>	AVROS 152	11

68.	Glutamate oxaloacetate transaminase composition	<i>band 0</i>	<i>bande 0</i>	SCH P 48	1
		band 1	bande 1	RRIM600,GT1,PB217,PB260	2
		band 1 + 2	bande 1 + 2	RRIC 110, PB 86, PB 235	3
		<i>band 1 + 3</i>	<i>bande 1 + 3</i>	RO 50	4
		<i>band 2</i>	<i>bande 2</i>	IRCA 707	5
(+)	Composition de la glutamate oxaloacétate transaminase				
69.	Shikimate dehydrogenase composition	<i>band 1</i>	<i>bande 1</i>	AC 54	1
		<i>band 1 + 3</i>	<i>bande 1 + 3</i>	IRCA 37	2
		<i>band 1 + 4</i>	<i>bande 1 + 4</i>		3
		<i>band 1 + 6</i>	<i>bande 1 + 6</i>	GU 969	4
		<i>band 2</i>	<i>bande 2</i>	RO 55	5
		<i>band 2 + 3</i>	<i>bande 2 + 3</i>	PA 31, AVROS 152	6
		<i>band 2 + 5</i>	<i>bande 2 + 5</i>	CNSAM 7621	7
		band 3	bande 3	RRIM600,GT1,PB217,PB260	8
		<i>band 3 + 4</i>	<i>bande 3 + 4</i>	IRCA 621, RRIC 132	9
		<i>band 3 + 5</i>	<i>bande 3 + 5</i>	RO 46	10
		<i>band 3 + 6</i>	<i>bande 3 + 6</i>	FX 25	11
		<i>band 4</i>	<i>bande 4</i>	F 4512	12
		<i>band 4 + 5</i>	<i>bande 4 + 5</i>	F 4542	13
		<i>band 5</i>	<i>bande 5</i>	AC 68	14
		<i>band 5 + 6</i>	<i>bande 5 + 6</i>		15

Isoforms most abundant are in bold; Isoforms most rare are in italic.

Part III

Description of the Method to be Used

Starch gel electrophoresis for rubber clone identification

1. Apparatus and equipment

Any suitable horizontal electrophoresis system can be used. Specific gel moulds have been elaborated by Cirad for allowing the analysis of 13 enzymatic systems in one same process during a one-day time period. These moulds have been adapted in a bridge shape with holes so that both edges of the gel can be dipped in the buffer of electrode vessels for circulation of electric current through the gel.

2. Chemicals

All chemicals should be of 'Analytical Reagent' grade or better. The list of the products is given in one table at the end of this document.

3. Solutions

3.1 Extraction solutions

The extraction buffer has to be prepared the day of the analysis or the day before (conservation in the fridge)

Tris (TRIZMA BASE)	424 mg
Cystein	60 mg
Distilled water up to	40 ml
Adjust pH with HCl 1N solution ($7.2 < \text{pH} < 7.5$)	
Complete to 50 ml.	

Keep it under cool conditions.

3.2 Electrophoresis (running buffer)

Tris-Citrate pH 6.6 :

Tris	18.2 g
Citric acid	10.5 g
Distilled water up to	1000 ml
Control pH (approximately 6.6)	

Tris-Citrate pH 8 :

Tris	18.6 g
Citric acid	8.4 g
Distilled water up to	1000 ml
Control pH (approximately 8)	

3.3 Gel preparation solutions

Tris-histidin pH 6 :

Histidin 5.25 g
Distilled water up to 400ml
Titrate to pH 6 with Tris 1M
Adjust level with distilled water up to 500 ml

Tris-histidin pH 8 :

Histidine 5.10 g
Distilled water up to 400ml
Titrate to pH 8 with Tris 1M
Adjust level with distilled water up to 500 ml

3.4 Staining buffers

Tris HCl 0.5M pH 8.5

Tris 60.6 g
Distilled water up to 800 ml
Titrate to pH 8.5 with HCl 1N
Adjust level with distilled water up to 1000ml

Phosphate 0.1M pH 6.5

Na₂HPO₄, 2H₂O 3.8 g
KH₂PO₄ 7.8 g
Distilled water up to 1000 ml

Acetate 0.5M pH 5

Sodium acetate 28.9 g
Acetic acid 8.5 ml
Distilled water up to 800 ml
Titrate to pH 5.4 with NaOH 10N
Adjust level with distilled water up to 1000ml

Tris maleate 0.1M pH 5.4

Tris 12.1 g
Maleic acid 11.6 g
Distilled water up to 800 ml
Titrate to pH 5.4 with NaOH 10N
Adjust level with distilled water up to 1000ml

3.5 Substrate solutions

Na malate 1M pH 7

DL malic acid 6.7 g
Distilled water 40 ml
Titrate to pH 7 with NaOH 10N
Adjust level with distilled water up to 50 ml

Leucine

Leucine β-Naphtyl amide HCl 250 mg

Dissolved in 100 ml distilled water (**Caution** : Leucine is very toxic, gloves are obligatory)

α -Naphthyl acetate / acetone

α -Naphthyl acetate	300 mg
Acétone	60 ml

3.6 Co-factor solutions

<u>NAD+</u>	10 mg / ml H ₂ O
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<u>NADP+</u>	10 mg / ml H ₂ O
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MgCl₂ 0.4M

MgCl ₂ , 6 H ₂ O	8.1 g
Distilled water up to	100 ml

3.7 Enzyme solution

<u>G6PDH</u> to use	Glucose 6 phosphate dehydrogenase : 1000 units / 250 μ l ready to use
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3.8 Staining solutions

<u>PMS</u>	Phenazine methosulfate	1 mg / ml
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<u>MTT</u>	Dimethylthiazol tetrazolium	5 mg / ml
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<u>DCPIP</u>	Dichlorophenol indophenol	1 mg / ml
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4. Procedure

4.1 Protein extraction

Enzymes can be rapidly destroyed by moderately hot temperatures. Preservation must be applied by ensuring cold conditions, or by freeze-drying of the leaf samples soon after collection.

Three anthocyanic leaflets (brown-redish = B stage) are collected and immediately kept in fresh conditions in a cool box with ice. For best results, time between sampling and extraction should be shortened. At the laboratory, preparation of leaves and extraction may occur in cold conditions (air conditioned room and/or ice bed). Take out mortars and pestles kept cooled in a fridge since day before. In each mortar, add 20 mg PVPP (Polyvinyl Polypyrrolidone insoluble), 50 mg fresh leaflets (discard petiolus), 0.5-0.6 ml extraction buffer. Crush the whole with a pestle up to obtain an homogeneous mixture. On each mortar, place 1 slip blotting paper (one layer) and 2 pieces of Whatman paper n°3 (1.0 x 0.7 cm).

4.2 Preparation of the gels

The gels are prepared the day before. One single gel allows to visualise 6 or 7 enzymatic systems (alanine amino peptidase and leucine amino peptidase are jointly visualised). Prepare 2 gels : 1 gel at pH6 (Tris-histidin buffer, pH 6), and 1 gel at pH8 (Tris-histidin buffer, pH 8).

Obturate the holes of the two gel moulds with adhesive tape. Place each mould on a level table and adjust horizontality. In a 1-litre Büchner flask weigh 62.0 g of starch (12% gel). Add 50 ml gel buffer (pH6 or pH8), complete with 450 ml of distilled water. Add a 80-mm magnetic stirring rod. Obturate the flask with a mortar in the ami to avoid evaporation of water. Heat the suspension (300°C) under continuous stirring up to the boiling point (apparition of big bubbles). Cooking one gel needs around 15 to 20 min. The starch paste obtained is degassed in the Büchner flask for 2 – 3 minutes. This starch paste is then carefully poured into the first mould in avoiding the formation of air bubbles. Repeat these operations for the second gel. Allow starch polymerization to take place at room temperature (1h30) and then cover with a plastic film to avoid dessication. Let the plastic-covered gels at room temperature overnight, then put them at 4°C, 30 min before use.

Using a guide and a spatula (with thin edge), 26 slots are made in each gel, on a line located at a distance of 4 cm from one edge of the mould (26 different samples can be analysed for the 13 enzymatic systems over the two gels). This starting line of migration is marked with bromophenol blue solution. With thin tweezers, each Whatman piece of paper bearing the extract of one sample is inserted in its corresponding slot; for each sample, one Whatman piece of paper is inserted in the gel pH6, and a second one bearing the same extract is inserted in the gel pH8. After sample loading, gels are covered again with transparent plastic film, and adhesive tapes are removed from the moulds.

4.3 Electrophoresis

Migration is carried out under cool conditions (cool room of fridge). The two edges of each gel in its mould are placed in the two electrode vessels filled with appropriate buffer : Tris-Citrate pH 6.6 for gel pH6 and Tris-Citrate pH 8 for gel pH8. Migration is carried out at a constant amperage (50mA for one gel). Migration is over when bromophenol blue reaches the anode vessel, which takes 5 to 6 hours.

4.4 Slicing the gels

The cathodic and anode edges of the gel are removed first by cutting the cathode edge at 1mm from the slot line, and then by cutting the anode edge approximately 9 cm from this line on the anode side. In order to keep a mark for orienting the gel during the following works, one corner is removed at one specific angle chosen by the operator. Gel is then transferred on a slicing guide (with 1.0 to 1.2 mm flange). Using the gel-slicer, the starch slab of 1cm thick can be cut into 6 thin slices. Each slice obtained is placed in a staining tray, according to a previous chosen order.

The following order can be proposed (slice 1 = bottom of the mould; * obligatory pH)

Slice	Gel pH6	Gel pH8
1	Malate dehydrogenase *	Phosphoglucomutase *
2	Alanine amino peptidase and Leucine amino peptidase	Glutamate oxaloacetate transaminase *
3	Phospho glucose isomerase *	Esterase
4	Phosphatase acid *	Isocitrate dehydrogenase *
5	Phospho Gluconate Dehydrogenase	Alcohol dehydrogenase
6	Diaphorase *	Shikimate dehydrogenase

4.5 Staining

Esterase

The gel slice is pre-incubated during 15 min at room temperature in pH 6.5 phosphate buffer (50ml). The buffer is poured out and a-Naphtyl acetate /acetone (15ml) with phosphate buffer up to 50 ml is added. Incubate 15 min in the dark at 40°C. The second incubation solution is poured out and the Fast Blue RR (50 mg dissolved in 25 ml distilled water, in the dark) is added. Control appearance of coloured bands, rinse two times and stop staining with acetic acid 10%.

Alcohol dehydrogenase

Incubate the gel slice in the dark at 40°C with the mix 0.5M pH8.5 Tris HCl buffer (10ml), NAD (1ml), MTT (1ml), PMS (1ml), Ethanol 95° (2ml), distilled water up to 50 ml. Control appearance of coloured bands, rinse two times and stop staining with acetic acid 10%.

Glucose phosphate isomerase

Incubate the gel slice in the dark at 40°C with the mix 0.5M pH8.5 Tris HCl buffer (20ml), 0.4 M MgCl₂ (1ml), NADP (1ml), G6PDH (12.5µl), MTT (1ml), PMS (1ml), fructose 6 phosphate (50mg), distilled water up to 50 ml. Control appearance of coloured bands, rinse two times and stop staining with acetic acid 10%.

Phosphoglucomutase

Incubate the gel slice in the dark at 40°C with the mix 0.5M pH8.5 Tris HCl buffer (20ml), 0.4 M MgCl₂ (1ml), NADP (1ml), G6PDH (12.5µl), MTT (1ml), PMS (1ml), glucose 1 phosphate (50mg), distilled water up to 50 ml. Control appearance of coloured bands, rinse two times and stop staining with acetic acid 10%.

Alanine aminopeptidase and leucine aminopeptidase

The gel slice is pre-incubated during 15 min in pH4.5 Tris Maleate buffer (20ml) with distilled water up to 50 ml. Discard the incubate solution and stain in the dark at 40°C with the mix : Fast Black K (50 mg) dissolved in Tris Malate buffer (20ml), completed with leucine (10ml) and distilled water up to 50 ml. Control appearance of coloured bands, rinse two times and stop staining with acetic acid 10%.

Glutamate oxaloacetate transaminase

The gel slice is incubated in the dark at 40°C with the mix : pH8.5 Tris HCl buffer (20ml), aspartic acid (200mg), a-ketoglutaric acid (100mg), EDTA (50mg), PVP (200 mg, M.W. 10 000), distilled water up to 50 ml, completed before complete dissolution with Fast Blue BB (80 mg) and Pyridoxal 5 Phosphate (8mg). Control appearance of coloured bands, rinse two times and stop staining with acetic acid 10%.

Malate dehydrogenase

Incubate the gel slice in the dark at 40°C with the mix : pH8.5 Tris HCl buffer (10ml), Na malate (5ml), NAD (1ml), MTT (1ml), PMS (1ml) and water up to 50 ml. Control appearance of coloured bands, rinse two times and stop staining with acetic acid 10%.

Isocitrate dehydrogenase

Incubate the gel slice in the dark at 40°C with the mix : pH8.5 Tris HCl buffer (10ml), 0.4 M MgCl₂ (1ml), NADP (1ml), MTT (1ml), PMS (1ml), sodium isocitrate (100mg) and distilled water up to 50 ml. Control appearance of coloured bands, rinse two times and stop staining

with acetic acid 10%. Control appearance of coloured bands, rinse two times and stop staining with acetic acid 10%.

Acid phosphatase

Pre-incubate the gel at room temperature in pH 5.0 acetate buffer (15ml), 0.4M MgCl₂ (1ml) and distilled water up to 50 ml. Pour out the buffer and add a-Naphtyl acid phosphate (90mg), b-Naphtyl acid phosphate (10mg), Fast Garnet GBC (50mg) dissolved in pH5.0 acetate buffer (15ml), 0.4M MgCl₂ (1ml) and distilled water up to 50 ml. Incubate in the dark at 40°C.

Phosphogluconate dehydrogenase

Incubate the gel slice in the dark at 40°C with the mix pH8.5 Tris HCl buffer (10ml), 0.4 M MgCl₂ (1ml), NADP (1ml), MTT (1ml), PMS (1ml), 6-phosphogluconic acid (30 mg), and distilled water up to 50 ml. Control appearance of coloured bands, rinse two times and stop staining with acetic acid 10%.

Diaphorase

Incubate the gel slice in the dark at 40°C with the mix pH8.5 Tris HCl buffer (10ml), Dichlorophenol indophenol DCPIP (2 ml), MTT (2ml), NADH (20 mg), and distilled water up to 50 ml. Control appearance of coloured bands, rinse two times. DO NOT stop staining with acetic acid, but let the slice under water.

Shikimate dehydrogenase

Incubate the gel slice in the dark at 40°C with the mix pH8.5 Tris HCl buffer (10ml), 0.4 M MgCl₂ (1ml), NADP (1ml), MTT (1ml), PMS (1ml), shikimic acid (100mg) and distilled water up to 50 ml. Control appearance of coloured bands, rinse two times and stop staining with acetic acid 10%.

4.6 Recognition of alleles

This table is designed to illustrate the alleles described above and to assist in the recognition of the different bands. It depicts the position of all enzyme systems bands, as compared to those found in the Example Variety GT1.

Nomenclature of the individual bands and recognition of the corresponding alleles

Characteristic 57 : Malate dehydrogenase locus A

	Example variety		Note				
	GT1	2	5	6	7	8	10
Locus B	—	—	—	—	—	—	—
4				—	—	—	—
3	—	—	—	—	—	—	
2	—	—	—	—	—		
1		—					

Characteristic 58 : Phospho glucose isomerase locus

	Example variety		Note					
	GT1	1	2	3	4	5	7	10
5						—		
4					—	—		
3	—	—	—	—	—
2	—	—	—	—	—	—	—
1	—	—	—	—	—	—		

Characteristic 59 : Alanine amino peptidase locus

	Example variety		Note	
	GT1	2	4	7
4			—	—
2	—	—	—	

Characteristic 60 : Leucine amino peptidase locus

	Example variety		Note							
	GT1	1	3	5	6	9	10	11	13	
7							—		
6						—		—	
5				—					
4	—		—	—	—		
2	—	—	—	—	—					
1		—								

Characteristic 61 : Esterase locus

	Example variety		Note			
	GT1	1	4	6	15	16
6				—	—	—
5	—
4			—			
1	—	—	—	—		

Characteristic 62 : Diaphorase locus

	Example variety		Note	
	GT1	4	5	7
	—			—
3	—	—
	—	—	—	—
2	—	—
	—	—	—	—
		—	—	
		—	—	

Characteristic 63 : Phosphatase acid locus

	Example variety		Note	
	GT1	3	4	5
2			—	—
			—	
1	—	—	—	

Characteristic 64 : Alcohol dehydrogenase locus

	Example variety		Note			
	GT1	1	2	3	4	5
4				—		—
				—		—
3	—		—	—	—	—
			—			
2		—	—	—		

Characteristic 65 : Isocitrate dehydrogenase locus

	Example variety		Note	
	GT1	6	7	8
4	—	—
			—	—
3	—	—	—	

Characteristic 66 : Phosphogluconate dehydrogenase locus

	Example variety		Note			
	GT1	2	3	4	5	6
Not interpreted locus		—	—	—	—	—
3	—		—		—	—
2	—	—	—	—	—	
1	—	—	—			

Characteristic 67 : Phosphoglucomatase locus

	Example variety		Note	
	GT1	3	8	9
5				—
4	—		—	—
3	—	
2	—	

Characteristic 68 : Glutamate oxaloacetate transaminase locus

	Example variety		Note		
	GT1	2	3	4	5
Slow locus	—	—	—	—	—
3				—	
2			—		—
1	—	—	—	—	
Not interpreted locus	—	—	—	—	—

Characteristic 69 : Shikimate dehydrogenase locus

	Example variety		Note			
	GT1	8	9	12	13	15
6	—
5					—	—
4	—	—	—	
3	—	—	—			

**Rubber electrophoresis
Chemical supplies references**

CHEMICALS (Complete name)	CHEMICAL (Abbreviated form)	SUPPLIER	CATALOG NUMBER
Alpha-NAPHTYL ACID PHOSPHATE monosodium salt	a-NAPHTYL ACID PHOSPHATE	SIGMA	N 7000
Alpha-NAPHTYL ACETATE crystalline	a-NAPHTYL ACETATE	SIGMA	N 8505
DL-ASPARTIC ACID free acid	ASPARTIC ACID	SIGMA	A 9006
MALEIC ACID DISODIUM SALT repurified	MALEIC ACID	SIGMA	M 0375
DL-MALIC ACID free acid	MALIC ACID	SIGMA	M 0875
STARCH POTATO HYDROLYZED FOR ELECTROPHORESIS	STARCH	SIGMA	S 4501
beta-NAPHTYL ACID PHOSPHATE monosodium salt	b- NAPHTYL ACID PHOSPHATE	SIGMA	N 7375
BROMOPHENOL BLUE sodium salt	BROMOPHENOL BLUE	SIGMA	B 8026
L-CYSTEINE free base	CYSTEIN	SIGMA	C 7755
2,6-DICHLOROPHENOL INDOPHENOL sodium salt	DCPIP	SIGMA	D 1878
ETHYLENEDIAMINETETRAACETIC ACID	EDTA	SIGMA	E 5513
FAST BLACK K SALT practical grade	FAST BLACK K	SIGMA	F 7253
FAST BLUE BB SALT practical grade	FAST BLUE BB	SIGMA	F 0250
FAST BLUE RR SALT crystalline	FAST BLUE RR	SIGMA	F 0500
FAST GARNET GBC SALT practical grade	FAST GARNET	SIGMA	F 0875
D-FRUCTOSE-6-PHOSPHATE disodium salt	FRUCTOSE 6 PHOSPHATE	SIGMA	F 3627
GLUCOSE-6-PHOSPHATE DESHYDROGENASE type XXIII	G6PDH	SIGMA	G 5760
Alpha-D GLUCOSE 1-PHOSPHATE disodium salt, hydrate, crystalline	GLUCOSE 1 PHOSPHATE	SIGMA	G 1259
L-HISTIDINE monohydrochloride : monohydrate	HISTIDIN	SIGMA	H 8125
DL-ISOCITRIC ACID trisodium salt	SODIUM ISOCITRATE	SIGMA	I 1252
Alpha-KETOGLUTARIC ACID free acid, crystalline	KETOGLUTARIC ACID	SIGMA	K 1750
L-LEUCINE beta-NAPHTYLAMIDE hydrochloride, crystalline	LEUCINE	SIGMA	L 0376
MAGNESIUM CHLORIDE hexahydrate	MgCl ₂	SIGMA	M 0250
DIMETHYLTHIAZOL TETRAZOLIUM	MTT	SIGMA	M 2128
beta-NICOTINAMIDE ADENINE DINUCLEOTIDE	NAD	SIGMA	N 3014
beta-NICOTINAMIDE ADENINE DINUCLEOTIDE reduced form , disodium salt	NADH	SIGMA	N 6005
beta-NICOTINAMIDE ADENINE DINUCLEOTIDE PHOSPHATE sodium salt	NADP	SIGMA	N 0505
6-PHOSPHOGLUCONIC ACID trisodiumsalt , grade III, crystalline	PHOSPHOGLUCONIC ACID	SIGMA	P 6888
PHENAZINE METHOSULFATE	PMS	SIGMA	P 9625
POLYVINYL PYRROLIDONE (M.W. 10 000)	PVP P.M. 10 000	SIGMA	PVP 10
POLYVINYL POLYPYRROLIDONE	PVPP	SIGMA	P 6755

PYRIDOXAL 5-PHOSPHATE	PYRIDOXAL 5 PHOSPHATE	SIGMA	P 9255
SHIKIMIC ACID	SHIKIMIC ACID	SIGMA	S 5375
SODIUM ACETATE ANHYDROUS	SODIUM ACETATE	SIGMA	S 8750
TRIS (TRIZMA BASE)	TRIS	SIGMA	T 1503
TRITON X100 laboratory grade	TRITO X100	SIGMA	X 100
ACETONE	ACETONE	MERCK	14 1000
ACETIC ACID	ACETIC ACID	MERCK	62 1000
CHLORYDRIC ACID	CHLORHYDRIC ACID	MERCK	317 1000
CITRIC ACID MONOHYDRATE	CITRIC ACID	MERCK	214 1000
ETHANOL	ETHANOL	MERCK	983 1000
POTASSIUM DIHYDROGENOPHOSPHATE	KH_2PO_4	MERCK	48731000
SODIUM HYDROGENOPHOSPHATE	Na_2HPO_4	MERCK	6580 0500
SODIUM HYDROXYDE	NaOH	MERCK	6498 1000
GLYCEROL (TECHNICAL GRADE)	GLYCEROL	LABOSI	G 350

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