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## INTERNATIONAL UNION FOR THE PROTECTION OF NEW VARIETIES OF PLANTS

Geneva

DRAFT

### WHITE CLOVER

UPOV Code(s): TRFOL\_REP

*Trifolium repens* L.

### GUIDELINES

#### FOR THE CONDUCT OF TESTS

#### FOR DISTINCTNESS, UNIFORMITY AND STABILITY

*prepared by an expert from the United Kingdom*

*to be considered by the*

*Technical Working Party for Agricultural Crops at its fifty-fifth session,  
to be held in Seoul, Republic of Korea, from 2026-06-15 to 2026-06-18*

*Disclaimer: this document does not represent UPOV policies or guidance*

#### Alternative Names:\*

Botanical name	English	French	German	Spanish
<i>Trifolium repens</i> L.	White Clover	Trèfle blanc	Weißklee	Trébol blanco

The purpose of these guidelines ("Test Guidelines") is to elaborate the principles contained in the General Introduction (document TG/1/3), and its associated TGP documents, into detailed practical guidance for the harmonized examination of distinctness, uniformity and stability (DUS) and, in particular, to identify appropriate characteristics for the examination of DUS and production of harmonized variety descriptions.

#### ASSOCIATED DOCUMENTS

These Test Guidelines should be read in conjunction with the General Introduction and its associated TGP documents.

\* These names were correct at the time of the introduction of these Test Guidelines but may be revised or updated. [Readers are advised to consult the UPOV Code, which can be found on the UPOV Website ([www.upov.int](http://www.upov.int)), for the latest information.]

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## 1. Subject of these Test Guidelines

These Test Guidelines apply to all varieties of *Trifolium repens* L.

## 2. Material Required

2.1 The competent authorities decide on the quantity and quality of the plant material required for testing the variety and when and where it is to be delivered. Applicants submitting material from a State other than that in which the testing takes place must ensure that all customs formalities and phytosanitary requirements are complied with.

2.2 The material is to be supplied in the form of seed.

2.3 The minimum quantity of plant material, to be supplied by the applicant, should be:

750 grams

The seed should meet the minimum requirements for germination, species and analytical purity, health and moisture content, specified by the competent authority. In cases where the seed is to be stored, the germination capacity should be as high as possible and should be, stated by the applicant.

2.4 The plant material supplied should be visibly healthy, not lacking in vigor, nor affected by any important pest or disease.

2.5 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If it has been treated, full details of the treatment must be given.

## 3. Method of Examination

### 3.1 *Number of Growing Cycles*

3.1.1 The minimum duration of tests should normally be two independent growing cycles.

3.1.2 The two independent growing cycles should be in the form of two separate plantings.

3.1.3 The testing of a variety may be concluded when the competent authority can determine with certainty the outcome of the test.

### 3.2 *Testing Place*

Tests are normally conducted at one place. In the case of tests conducted at more than one place, guidance is provided in TGP/9 "Examining Distinctness".

### 3.3 *Conditions for Conducting the Examination*

3.3.1 The tests should be carried out under conditions ensuring satisfactory growth for the expression of the relevant characteristics of the variety and for the conduct of the examination.

3.3.2 The optimum stage of development for the assessment of each characteristic is indicated by a reference in the Table of Characteristics. The stages of development denoted by each reference are described in Chapter 8.

### 3.4 *Test Design*

Spaced plants: Each test should be designed to result in a total of at least 60 plants, which should be divided between at least 2 replicates.

Row plots: Each test should be designed to result in a total of at least 200 plants, which should be divided between at least 2 replicates.

Special test: Each test should be designed to result in a total of at least 60 plants, which should be divided between at least 2 replicates.

The design of the tests should be such that plants or parts of plants may be removed for measurement or counting without prejudice to the observations which must be made up to the end of the growing cycle.

Where observations can be made both in spaced plants and row plots, it is likely that the expression of the characteristics will be different. Therefore, these different observations may not be interchangeable in the DUS examination.

### 3.5 *Additional Tests*

Additional tests, for examining relevant characteristics, may be established.

## 4. Assessment of Distinctness, Uniformity and Stability

### 4.1 *Distinctness*

#### 4.1.1 General Recommendations

It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding distinctness. However, the following points are provided for elaboration or emphasis in these Test Guidelines.

#### 4.1.2 Consistent Differences

The differences observed between varieties may be so clear that more than one growing cycle is not necessary. In addition, in some circumstances, the influence of the environment is not such that more than a single growing cycle is required to provide assurance that the differences observed between varieties are sufficiently consistent. One means of ensuring that a difference in a characteristic, observed in a growing trial, is sufficiently consistent is to examine the characteristic in at least two independent growing cycles.

#### 4.1.3 Clear Differences

Determining whether a difference between two varieties is clear depends on many factors, and should consider, in particular, the type of expression of the characteristic being examined, i.e. whether it is expressed in a qualitative, quantitative, or pseudo-qualitative manner. Therefore, it is important that users of these Test Guidelines are familiar with the recommendations contained in the General Introduction prior to making decisions regarding distinctness.

#### 4.1.4 Number of Plants or Parts of Plants to be Examined

Unless otherwise indicated, for the purposes of distinctness, all observations on single plants should be made on 10 plants or parts taken from each of 10 plants and any other observations made on all plants in the test, disregarding any off-type plants.

In the case of observations of parts taken from single plants, the number of parts to be taken from each of the plants should be 1

#### 4.1.5 Method of Observation

The recommended method of observing the characteristic for the purposes of distinctness is indicated by the following key in the Table of Characteristics (see document TGP/9 "Examining Distinctness", Section 4 "Observation of characteristics"):

MG: single measurement of a group of plants or parts of plants

MS: measurement of a number of individual plants or parts of plants

VG: visual assessment by a single observation of a group of plants or parts of plants

VS: visual assessment by observation of individual plants or parts of plants

Type of observation: visual (V) or measurement (M)

"Visual" observation (V) is an observation made on the basis of the expert's judgment. For the purposes of this document, "visual" observation refers to the sensory observations of the experts and, therefore, also includes smell, taste and touch. Visual observation includes observations where the expert uses reference points (e.g. diagrams, example varieties, side-by-side comparison) or non-linear charts (e.g. color charts). Measurement (M)

is an objective observation against a calibrated, linear scale e.g. using a ruler, weighing scales, colorimeter, dates, counts, etc.

Type of record: for a group of plants (G) or for single, individual plants (S)

For the purposes of distinctness, observations may be recorded as a single record for a group of plants or parts of plants (G), or may be recorded as records for a number of single, individual plants or parts of plants (S). In most cases, "G" provides a single record per variety and it is not possible or necessary to apply statistical methods in a plant-by-plant analysis for the assessment of distinctness.

In cases where more than one method of observing the characteristic is indicated in the Table of Characteristics (e.g. VG/MG), guidance on selecting an appropriate method is provided in document TGP/9, Section 4.2.

#### 4.2 *Uniformity*

4.2.1 It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding uniformity. However, the following points are provided for elaboration or emphasis in these Test Guidelines:

4.2.2 The assessment of uniformity should be according to the recommendations for cross-pollinated varieties in the General Introduction.

4.2.3 For the assessment of uniformity of a variety, the standard deviation of the mean value for each characteristic should be compared with the mean of the standard deviations of comparable varieties using a recognized statistical technique.

#### 4.3 *Stability*

4.3.1 In practice, it is not usual to perform tests of stability that produce results as certain as those of the testing of distinctness and uniformity. However, experience has demonstrated that, for many types of variety, when a variety has been shown to be uniform, it can also be considered to be stable.

4.3.2 Where appropriate, or in cases of doubt, stability may be further examined by testing a new seed stock to ensure that it exhibits the same characteristics as those shown by the initial material supplied.

### 5. Grouping of Varieties and Organization of the Growing Trial

5.1 The selection of varieties of common knowledge to be grown in the trial with the candidate varieties and the way in which these varieties are divided into groups to facilitate the assessment of distinctness are aided by the use of grouping characteristics.

5.2 Grouping characteristics are those in which the documented states of expression, even where produced at different locations, can be used, either individually or in combination with other such characteristics: (a) to select varieties of common knowledge that can be excluded from the growing trial used for examination of distinctness; and (b) to organize the growing trial so that similar varieties are grouped together.

5.3 The following have been agreed as useful grouping characteristics:

5.4 Guidance for the use of grouping characteristics, in the process of examining distinctness, is provided through the General Introduction and document TGP/9 "Examining Distinctness".

## 6. Introduction to the Table of Characteristics

### 6.1 *Categories of Characteristics*

#### 6.1.1 Standard Test Guidelines Characteristics

Standard Test Guidelines characteristics are those which are approved by UPOV for examination of DUS and from which members of the Union can select those suitable for their particular circumstances.

#### 6.1.2 Asterisked Characteristics

Asterisked characteristics (denoted by \*) are those included in the Test Guidelines which are important for the international harmonization of variety descriptions and should always be examined for DUS and included in the variety description by all members of the Union, except when the state of expression of a preceding characteristic or regional environmental conditions render this inappropriate.

### 6.2 *States of Expression and Corresponding Notes*

6.2.1 States of expression are given for each characteristic to define the characteristic and to harmonize descriptions. Each state of expression is allocated a corresponding numerical note for ease of recording of data and for the production and exchange of the description.

6.2.2 All relevant states of expression are presented in the characteristic.

6.2.3 Further explanation of the presentation of states of expression and notes is provided in document TGP/7 "Development of Test Guidelines".

### 6.3 *Types of Expression*

An explanation of the types of expression of characteristics (qualitative, quantitative and pseudo-qualitative) is provided in the General Introduction.

### 6.4 *Example Varieties*

Where appropriate, example varieties are provided to clarify the states of expression of each characteristic.

## 6.5 Legend

English				français		deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
1	2	3	4	5	6	7			
		Name of characteristics in English		Nom du caractère en français		Name des Merkmals auf Deutsch		Nombre del carácter en español	
		states of expression		types d'expression		Ausprägungsstufen		tipos de expresión	

- 1 Characteristic number
- 2 (\*) sterisked characteristic – see Chapter 6.1.2
- 3 Type of expression  
 QL Qualitative characteristic – see Chapter 6.3  
 QN Quantitative characteristic – see Chapter 6.3  
 PQ Pseudo-qualitative characteristic – see Chapter 6.3
- 4 Method of observation (and type of plot, if applicable)  
 MG, MS, VG, VS – see Chapter 4.1.5
- 5 (+) See Explanations on the Table of Characteristics in Chapter 8.2
- 6 (a)-(d) See Explanations on the Table of Characteristics in Chapter 8.1
- 7 Not applicable
- A - spaced plants  
 B - row plots

7. Table of Characteristics/Tableau des caracteres/Merkmalestabelle/Tabla de caracteres

		English		français		deutsch		español		Example Varieties Exemples Beispielssorten Variedades ejemplo		Note/ Nota	
1.		QN	VS A	(+)									
		<b>Plant: tendency to form inflorescences before vernalization</b>											
		absent or very weak										1	
		very weak to weak										2	
		weak										3	
		weak to medium										4	
		medium										5	
		medium to strong										6	
		strong										7	
		strong to very strong										8	
		very strong										9	
2.	(*)	QN	VG B VS A	(+)									
		<b>Plant: intensity of green color</b>											
		very light										1	
		very light to light										2	
		light										3	
		light to medium										4	
		medium										5	
		medium to dark										6	
		dark										7	
		dark to very dark										8	
		very dark										9	
3.		QN	VG B VS A	(+)									
		<b>Plant: density of foliage</b>											
		very low										1	
		very low to low										2	
		low										3	
		low to medium										4	
		medium										5	
		medium to high										6	
		high										7	
		high to very high										8	
		very high										9	

		English		français		deutsch		español		Example Varieties Exemples Beispielsorten Variedades ejemplo		Note/ Nota
4.		QN	MS	(+)								
		<b>Plant: proportion of plants with cyanid glucoside</b>										
		absent or very low										1
		very low to low										2
		low										3
		low to medium										4
		medium										5
		medium to high										6
		high										7
		high to very high										8
		very high										9
5.	(*)	QN	MG B MS A	(+)								
		<b>Plant: time of flowering</b>										
		very early										1
		very early to early										2
		early										3
		early to medium										4
		medium										5
		medium to late										6
		late										7
		late to very late										8
		very late										9
6.		QN	MG B MS A		(a)							
		<b>Plant: height</b>										
		very short										1
		very short to short										2
		short										3
		short to medium										4
		medium										5
		medium to tall										6
		tall										7
		tall to very tall										8
		very tall										9

		English		français		deutsch		español		Example Varieties Exemples Beispielssorten Variedades ejemplo		Note/ Nota
7.		QN	MS A		(a)							
		<b>Plant: width</b>										
		very narrow										1
		very narrow to narrow										2
		narrow										3
		narrow to medium										4
		medium										5
		medium to broad										6
		broad										7
		broad to very broad										8
		very broad										9
8.		QN	VG B VS A		(a)							
		<b>Plant: growth habit</b>										
		erect										1
		erect to semi-erect										2
		semi-erect										3
		semi-erect to intermediate										4
		intermediate										5
		intermediate to semi-prostrate										6
		semi-prostrate										7
		semi-prostrate to prostrate										8
		prostrate										9
9.		QN	MS A	(+)	(b)							
		<b>Stem: internode length of stolon</b>										
		very short										1
		very short to short										2
		short										3
		short to medium										4
		medium										5
		medium to long										6
		long										7
		long to very long										8
		very long										9

		English		français		deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
10.		QN	MS A	(+)	(b)				
		<b>Stem: thickness of stolon</b>							
		very thin							1
		very thin to thin							2
		thin							3
		thin to medium							4
		medium							5
		medium to thick							6
		thick							7
		thick to very thick							8
		very thick							9
11.	(*)	QN	VG B VS A		(b)				
		<b>Leaf: prominence of white markings</b>							
		absent or very weak							1
		very weak to weak							2
		weak							3
		weak to medium							4
		medium							5
		medium to strong							6
		strong							7
		strong to very strong							8
		very strong							9
12.		QN	MS A	(+)	(b), (c)				
		<b>Leaf: length of petiole</b>							
		very short							1
		very short to short							2
		short							3
		short to medium							4
		medium							5
		medium to long							6
		long							7
		long to very long							8
		very long							9

		English		français		deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
13.		QN	MS A	(+)	(b), (c)				
		<b>Leaf: thickness of petiole</b>							
		very thin							1
		very thin to thin							2
		thin							3
		thin to medium							4
		medium							5
		medium to thick							6
		thick							7
		thick to very thick							8
		very thick							9
14.	(*)	QN	MS A	(+)	(b), (c)				
		<b>Leaf: length of median leaflet</b>							
		very short							1
		very short to short							2
		short							3
		short to medium							4
		medium							5
		medium to long							6
		long							7
		long to very long							8
		very long							9
15.	(*)	QN	MS A	(+)	(b), (c)				
		<b>Leaf: width of median leaflet</b>							
		very narrow							1
		very narrow to narrow							2
		narrow							3
		narrow to medium							4
		medium							5
		medium to broad							6
		broad							7
		broad to very broad							8
		very broad							9

		English		français		deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
16.	(*)	QN	MS A	(+)	(b)				
		<b>Leaf: size of median leaflet</b>							
		very small							1
		very small to small							2
		small							3
		small to medium							4
		medium							5
		medium to large							6
		large							7
		large to very large							8
		very large							9
17.		QN	MS A	(+)	(b)				
		<b>Leaf: ratio of length to width of median leaflet</b>							
		very small							1
		very small to small							2
		small							3
		small to medium							4
		medium							5
		medium to large							6
		large							7
		large to very large							8
		very large							9
18.		QN	VS A		(d)				
		<b>Plant: number of inflorescences</b>							
		very few							1
		very few to few							2
		few							3
		few to medium							4
		medium							5
		medium to many							6
		many							7
		many to very many							8
		very many							9

		English		français		deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
19.		QN	MS A	(+)	(d)				
		<b>Inflorescence: diameter</b>							
		very small							1
		very small to small							2
		small							3
		small to medium							4
		medium							5
		medium to large							6
		large							7
		large to very large							8
20.		QN	MS A	(+)	(d)				
		<b>Inflorescence: length of peduncle</b>							
		very short							1
		very short to short							2
		short							3
		short to medium							4
		medium							5
		medium to long							6
		long							7
		long to very long							8
		very long							9
21.		QN	MS A	(+)	(d)				
		<b>Inflorescence: thickness of peduncle</b>							
		very thin							1
		very thin to thin							2
		thin							3
		thin to medium							4
		medium							5
		medium to thick							6
		thick							7
		thick to very thick							8
		very thick							9

## 8. Explanations on the Table of Characteristics

### 8.1 *Explanations covering several characteristics*

Characteristics containing the following key in the Table of Characteristics should be examined as indicated below:

- (a) Observations should be made at the time of flowering.
- (b) Observations should be made after all the plants of each variety in a replicate have flowered, and within 1-2 weeks after flowering. The longest healthily growing stolon should be selected from each plant for measurement.
- (c) Observations should be made on the third expanded leaf, counted from the growing tip of the stolon.
- (d) Observations should be made on all plants at maturity (normally 30 days after the mean date of flowering).

### 8.2 *Explanations for individual characteristics*

#### Ad. 1: Plant: tendency to form inflorescences before vernalization

Observations should be made before the period of vernalization. The number of flower heads produced on each plant should be assessed and scored.

#### Ad. 2: Plant: intensity of green color

Observations should be made in the vegetative phase by examination and scoring of the overall green color of the plant.

#### Ad. 3: Plant: density of foliage

Observations should be made in the vegetative phase by examination and scoring of the overall ground cover of the foliage of the plant.

Ad. 4: Plant: proportion of plants with cyanid glucoside

**Preparation of picro-sodic paper (indicator paper):**

1. Weigh out 96.25g of picric acid solution.
2. 6.25g of sodium carbonate is dissolved in 125 ml of distilled water.
3. Add the sodium carbonate solution to the picric acid solution, mix and store in a labelled amber reagent bottle.
4. Strips of Whatmann No. 1 filter paper are dipped in this solution and can be stored dry in a desiccator.

**Test Procedure:**

1. Healthy leaves (preferably folded) are selected from each of the sixty plants and put into separate eppendorf tubes (one trifoliate leaf per tube).
2. The tubes are closed and placed in a freezer at  $-18^{\circ}\text{C}$  for a minimum of two hours.
3. After freezing, a strip of indicator paper is placed across the opening of the eppendorf tubes and the lid closed. This is sufficient to hold the paper in place.
4. The tubes are placed in darkness in a water bath at  $50^{\circ}\text{C}$  for two hours.
5. If there is cyanid glucoside (HCN) present the paper will change from yellow to red. The color reaction is recorded as presence/absence of red color for each of the sixty plants.

Ad. 5: Plant: time of flowering

Observations should be made on all plants.

(a) In single spaced plants, the time of flowering of the variety is the time when 50% of the plants have three inflorescences per plant showing color.

(b) In row plots the time of flowering for all plants in a variety is observed and the time of flowering of the variety is the time when 80% of the plants would have been flowering. - can examiners who use row plots please provide updated wording for this?

Ad. 9: Stem: internode length of stolon

Observations should be made between the third and fourth node counted from the growing tip.

Ad. 10: Stem: thickness of stolon

Observations should be made at the midway point between the third and the fourth node counted from the growing tip.

Ad. 12: Leaf: length of petiole

Observations should be made from the base of the median trifoliate leaflet to the stolon.

Ad. 13: Leaf: thickness of petiole

Observations should be taken at the widest point of the petiole.

Ad. 14: Leaf: length of median leaflet

Observations should be made on the median trifoliate leaflet.

Ad. 15: Leaf: width of median leaflet

See ad. 15.

Ad. 16: Leaf: size of median leaflet

Calculated from the measurements of Leaf: length (characteristic 14) x Leaf: width (characteristic 15)

Ad. 17: Leaf: ratio of length to width of median leaflet

Calculated from the ratio of Leaf: length (characteristic 14)/Leaf: width (characteristic 15).

Ad. 19: Inflorescence: diameter

Observations should be made on inflorescence taken from close to the center of the plant.

Ad. 20: Inflorescence: length of peduncle

See ad. 19.

The length of the peduncle should be measured from the base of the inflorescence to the stolon.

Ad. 21: Inflorescence: thickness of peduncle

See ad 19.

The thickness of the peduncle should be measured at a point midway between the base of the inflorescence and the stolon.

9. Literature

No specific literature.

10. Technical Questionnaire

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
-------------------------	-----------------	-------------------

		Application date: (not to be filled in by the applicant)
--	--	---

TECHNICAL QUESTIONNAIRE  
to be completed in connection with an application for plant breeders' rights

1. Subject of the Technical Questionnaire

1.1 Botanical name

*Trifolium repens* L.

1.2 Common name

White Clover

2. Applicant

Name

Address

Telephone No.

Fax No.

E-mail address

Breeder (if different from  
applicant)

3. Proposed denomination and breeder's reference

Proposed denomination (if  
available)

Breeder's reference

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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#4. Information on the breeding scheme and propagation of the variety

4.1 Breeding scheme

Variety resulting from:

4.1.1 Other  
(Please provide details)

# Authorities may allow certain of this information to be provided in a confidential section of the Technical Questionnaire.

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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4.2 Method of propagating the variety

4.2.1 Seed-propagated varieties

(a) Cross-pollination

(b) Other (Please provide details)

4.2.2 Other (Please provide details)

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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5. Characteristics of the variety to be indicated (the number in brackets refers to the corresponding characteristic in Test Guidelines; please mark the note which best corresponds).

	Characteristics	Example Varieties	Note
<b>5.1 (5)</b>	<b>Plant: time of flowering</b>		
	very early		1 [ ]
	very early to early		2 [ ]
	early		3 [ ]
	early to medium		4 [ ]
	medium		5 [ ]
	medium to late		6 [ ]
	late		7 [ ]
	late to very late		8 [ ]
	very late		9 [ ]
<b>5.2 (11)</b>	<b>Leaf: prominence of white markings</b>		
	absent or very weak		1 [ ]
	very weak to weak		2 [ ]
	weak		3 [ ]
	weak to medium		4 [ ]
	medium		5 [ ]
	medium to strong		6 [ ]
	strong		7 [ ]
	strong to very strong		8 [ ]
	very strong		9 [ ]
<b>5.3 (14)</b>	<b>Leaf: length of median leaflet</b>		
	very short		1 [ ]
	very short to short		2 [ ]
	short		3 [ ]
	short to medium		4 [ ]
	medium		5 [ ]
	medium to long		6 [ ]
	long		7 [ ]
	long to very long		8 [ ]
	very long		9 [ ]

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Characteristics	Example Varieties	Note
<b>5.4 (15)</b>	<b>Leaf: width of median leaflet</b>	
	very narrow	1 [ ]
	very narrow to narrow	2 [ ]
	narrow	3 [ ]
	narrow to medium	4 [ ]
	medium	5 [ ]
	medium to broad	6 [ ]
	broad	7 [ ]
	broad to very broad	8 [ ]
	very broad	9 [ ]
<b>5.5 (16)</b>	<b>Leaf: size of median leaflet</b>	
	very small	1 [ ]
	very small to small	2 [ ]
	small	3 [ ]
	small to medium	4 [ ]
	medium	5 [ ]
	medium to large	6 [ ]
	large	7 [ ]
	large to very large	8 [ ]
	very large	9 [ ]

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6. Similar varieties and differences from these varieties

*Please use the following table and box for comments to provide information on how your candidate variety differs from the variety (or varieties) which, to the best of your knowledge, is (or are) most similar. This information may help the examination authority to conduct its examination of distinctness in a more efficient way*

Denomination(s) of variety(ies) similar to your candidate variety	Characteristic(s) in which your candidate variety differs from the similar variety(ies)	Describe the expression of the characteristic(s) for the <b>similar</b> variety(ies)	Describe the expression of the characteristic(s) for <b>your</b> candidate variety
<i>Example</i>	<i>time of flowering</i>	<i>early</i>	<i>medium</i>

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<p>Comments</p>
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#7. Additional information which may help in the examination of the variety

7.1 In addition to the information provided in sections 5 and 6, are there any additional characteristics which may help to distinguish the variety?

Yes  No

(If yes, please provide details)

7.2 Are there any special conditions for growing the variety or conducting the examination?

Yes  No

(If yes, please provide details)

7.3 Other information

8. Authorization for release

(a) Does the variety require prior authorization for release under legislation concerning the protection of the environment, human and animal health?

Yes [ ]                  No [ ]

(b) Has such authorization been obtained?

Yes [ ]                  No [ ]

If the answer to (b) is yes, please attach a copy of the authorization.

9. Information on plant material to be examined or submitted for examination

9.1 The expression of a characteristic or several characteristics of a variety may be affected by factors, such as pests and disease, chemical treatment (e.g. growth retardants or pesticides), effects of tissue culture, different rootstocks, scions taken from different growth phases of a tree, etc.

9.2 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If the plant material has undergone such treatment, full details of the treatment must be given. In this respect, please indicate below, to the best of your knowledge, if the plant material to be examined has been subjected to:

- |   |         |        |
|---|---------|--------|
| (a) Microorganisms (e.g. virus, bacteria, phytoplasma)    | Yes [ ] | No [ ] |
| (b) Chemical treatment (e.g. growth retardant, pesticide) | Yes [ ] | No [ ] |
| (c) Tissue culture  | Yes [ ] | No [ ] |
| (d) Other factors   | Yes [ ] | No [ ] |

Please provide details for where you have indicated "yes".

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10. I hereby declare that, to the best of my knowledge, the information provided in this form is correct:

Applicant's name

Signature

Date

[End of document]