

INTERNATIONAL UNION FOR THE PROTECTION OF NEW VARIETIES OF PLANTS

Geneva

DRAFT

FODDER BEET

UPOV Code(s): BETAA_VUL_GVA

Beta vulgaris L. Fodder Beet Group

GUIDELINES

FOR THE CONDUCT OF TESTS

FOR DISTINCTNESS, UNIFORMITY AND STABILITY

prepared by an expert from France

to be considered by the

*Technical Working Party for Agricultural Crops at its fifty-fifth session,
to be held in Seoul, Republic of Korea, from 2026-06-15 to 2026-06-18*

Disclaimer: this document does not represent UPOV policies or guidance

Alternative Names:*

Botanical name	English	French	German	Spanish
<i>Beta vulgaris</i> L. Fodder Beet Group, <i>Beta vulgaris</i> L. ssp. <i>vulgaris</i> var. <i>alba</i> DC., <i>Beta vulgaris</i> L. ssp. <i>vulgaris</i> var. <i>crassa</i> Alef., <i>Beta vulgaris</i> L. ssp. <i>vulgaris</i> var. <i>crassa</i> Mansf., <i>Beta vulgaris</i> L. ssp. <i>vulgaris</i> var. <i>rapacea</i> K. Koch	Fodder beet	Betterave fourragère	Runkelrübe	Remolacha forrajera

The purpose of these guidelines (“Test Guidelines”) is to elaborate the principles contained in the General Introduction (document TG/1/3), and its associated TGP documents, into detailed practical guidance for the harmonized examination of distinctness, uniformity and stability (DUS) and, in particular, to identify appropriate characteristics for the examination of DUS and production of harmonized variety descriptions.

ASSOCIATED DOCUMENTS

These Test Guidelines should be read in conjunction with the General Introduction and its associated TGP documents.

* These names were correct at the time of the introduction of these Test Guidelines but may be revised or updated. [Readers are advised to consult the UPOV Code, which can be found on the UPOV Website (www.upov.int), for the latest information.]

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1. Subject of these Test Guidelines

These Test Guidelines apply to all varieties of *Beta vulgaris* L. Fodder Beet Group.

2. Material Required

2.1 The competent authorities decide on the quantity and quality of the plant material required for testing the variety and when and where it is to be delivered. Applicants submitting material from a State other than that in which the testing takes place must ensure that all customs formalities and phytosanitary requirements are complied with.

2.2 The material is to be supplied in the form of naked seeds.

2.3 The minimum quantity of plant material, to be supplied by the applicant, should be:

350 g

The seed should meet the minimum requirements for germination, species and analytical purity, health and moisture content, specified by the competent authority. In cases where the seed is to be stored, the germination capacity should be as high as possible and should be, stated by the applicant.

2.4 The plant material supplied should be visibly healthy, not lacking in vigor, nor affected by any important pest or disease.

2.5 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If it has been treated, full details of the treatment must be given.

3. Method of Examination

3.1 *Number of Growing Cycles*

3.1.1 The minimum duration of tests should normally be two independent growing cycles.

3.1.2 The two independent growing cycles should be in the form of two separate plantings.

3.1.3 The testing of a variety may be concluded when the competent authority can determine with certainty the outcome of the test.

3.2 *Testing Place*

Tests are normally conducted at one place. In the case of tests conducted at more than one place, guidance is provided in TGP/9 "Examining Distinctness".

3.3 *Conditions for Conducting the Examination*

3.3.1 The tests should be carried out under conditions ensuring satisfactory growth for the expression of the relevant characteristics of the variety and for the conduct of the examination.

3.3.2 The optimum stage of development for the assessment of each characteristic is indicated by a reference in the Table of Characteristics. The stages of development denoted by each reference are described in Chapter 8.

3.4 *Test Design*

Each test should be designed to result in a total of at least 200 plants, which should be divided between at least 2 replicates.

3.5 *Additional Tests*

Additional tests, for examining relevant characteristics, may be established.

4. Assessment of Distinctness, Uniformity and Stability

4.1 *Distinctness*

4.1.1 General Recommendations

It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding distinctness. However, the following points are provided for elaboration or emphasis in these Test Guidelines.

4.1.2 Consistent Differences

The differences observed between varieties may be so clear that more than one growing cycle is not necessary. In addition, in some circumstances, the influence of the environment is not such that more than a single growing cycle is required to provide assurance that the differences observed between varieties are sufficiently consistent. One means of ensuring that a difference in a characteristic, observed in a growing trial, is sufficiently consistent is to examine the characteristic in at least two independent growing cycles.

4.1.3 Clear Differences

Determining whether a difference between two varieties is clear depends on many factors, and should consider, in particular, the type of expression of the characteristic being examined, i.e. whether it is expressed in a qualitative, quantitative, or pseudo-qualitative manner. Therefore, it is important that users of these Test Guidelines are familiar with the recommendations contained in the General Introduction prior to making decisions regarding distinctness.

4.1.4 Number of Plants or Parts of Plants to be Examined

Unless otherwise indicated, for the purposes of distinctness, all observations on single plants should be made on 45 plants or parts taken from each of 45 plants and any other observations made on all plants in the test, disregarding any off-type plants.

4.1.5 Method of Observation

The recommended method of observing the characteristic for the purposes of distinctness is indicated by the following key in the Table of Characteristics (see document TGP/9 "Examining Distinctness", Section 4 "Observation of characteristics"):

MG: single measurement of a group of plants or parts of plants

MS: measurement of a number of individual plants or parts of plants

VG: visual assessment by a single observation of a group of plants or parts of plants

VS: visual assessment by observation of individual plants or parts of plants

Type of observation: visual (V) or measurement (M)

"Visual" observation (V) is an observation made on the basis of the expert's judgment. For the purposes of this document, "visual" observation refers to the sensory observations of the experts and, therefore, also includes smell, taste and touch. Visual observation includes observations where the expert uses reference points (e.g. diagrams, example varieties, side-by-side comparison) or non-linear charts (e.g. color charts). Measurement (M) is an objective observation against a calibrated, linear scale e.g. using a ruler, weighing scales, colorimeter, dates, counts, etc.

Type of record: for a group of plants (G) or for single, individual plants (S)

For the purposes of distinctness, observations may be recorded as a single record for a group of plants or parts of plants (G), or may be recorded as records for a number of single, individual plants or parts of plants (S). In most cases, "G" provides a single record per variety and it is not possible or necessary to apply statistical methods in a plant-by-plant analysis for the assessment of distinctness.

In cases where more than one method of observing the characteristic is indicated in the Table of Characteristics (e.g. VG/MG), guidance on selecting an appropriate method is provided in document TGP/9, Section 4.2.

4.2 *Uniformity*

4.2.1 It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding uniformity. However, the following points are provided for elaboration or emphasis in these Test Guidelines:

4.2.2 These Test Guidelines have been developed for the examination of cross-pollinated and hybrid varieties. For varieties with other types of propagation, the recommendations in the General Introduction and document TGP/13 "Guidance for new types and species" Section 4.5 "Testing Uniformity" should be followed.

4.2.3 The recommended sample size for the assessment of uniformity is indicated by the following key in the table of characteristics:

A sample size of 100 plants/parts of plants

B sample size of 200 plants

4.2.4 If not otherwise indicated, for the assessment of uniformity, a population standard of 2% and an acceptance probability of at least 95% should be applied. In the case of a sample size of 200 plants, 7 off-types are allowed. For ploidy, a population standard of 10% and an acceptance probability of at least 95% should be applied. In the case of a sample size of 15 plants, 1 off-type is allowed.

4.2.5 For root characteristics, the assessment of uniformity can be done in 2 steps. In a first step, 45 roots are observed. If 3 or less off-types are observed, the variety is declared to be uniform. If more than 3 off-types are observed, an additional sample of 55 roots must be observed.

4.3 *Stability*

4.3.1 In practice, it is not usual to perform tests of stability that produce results as certain as those of the testing of distinctness and uniformity. However, experience has demonstrated that, for many types of variety, when a variety has been shown to be uniform, it can also be considered to be stable.

4.3.2 Where appropriate, or in cases of doubt, stability may be further examined by testing a new seed stock to ensure that it exhibits the same characteristics as those shown by the initial material supplied.

5. Grouping of Varieties and Organization of the Growing Trial

5.1 The selection of varieties of common knowledge to be grown in the trial with the candidate varieties and the way in which these varieties are divided into groups to facilitate the assessment of distinctness are aided by the use of grouping characteristics.

5.2 Grouping characteristics are those in which the documented states of expression, even where produced at different locations, can be used, either individually or in combination with other such characteristics: (a) to select varieties of common knowledge that can be excluded from the growing trial used for examination of distinctness; and (b) to organize the growing trial so that similar varieties are grouped together.

5.3 The following have been agreed as useful grouping characteristics:

- (a) Germity (characteristic 1)
- (b) Ploidy (characteristic 2)
- (c) Root: shape (characteristic 17)
- (d) Root: color below ground (characteristic 21)

5.4 Guidance for the use of grouping characteristics, in the process of examining distinctness, is provided through the General Introduction and document TGP/9 "Examining Distinctness".

6. Introduction to the Table of Characteristics

6.1 *Categories of Characteristics*

6.1.1 Standard Test Guidelines Characteristics

Standard Test Guidelines characteristics are those which are approved by UPOV for examination of DUS and from which members of the Union can select those suitable for their particular circumstances.

6.1.2 Asterisked Characteristics

Asterisked characteristics (denoted by *) are those included in the Test Guidelines which are important for the international harmonization of variety descriptions and should always be examined for DUS and included in the variety description by all members of the Union, except when the state of expression of a preceding characteristic or regional environmental conditions render this inappropriate.

6.2 *States of Expression and Corresponding Notes*

6.2.1 States of expression are given for each characteristic to define the characteristic and to harmonize descriptions. Each state of expression is allocated a corresponding numerical note for ease of recording of data and for the production and exchange of the description.

6.2.2 All relevant states of expression are presented in the characteristic.

6.2.3 Further explanation of the presentation of states of expression and notes is provided in document TGP/7 "Development of Test Guidelines".

6.3 *Types of Expression*

An explanation of the types of expression of characteristics (qualitative, quantitative and pseudo-qualitative) is provided in the General Introduction.

6.4 *Example Varieties*

Where appropriate, example varieties are provided to clarify the states of expression of each characteristic.

6.5 Legend

English				français		deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
1	2	3	4	5	6	7			
		Name of characteristics in English		Nom du caractère en français		Name des Merkmals auf Deutsch		Nombre del carácter en español	
		states of expression		types d'expression		Ausprägungsstufen		tipos de expresión	

1 Characteristic number

2 (*) sterisked characteristic – see Chapter 6.1.2

3 Type of expression
 QL Qualitative characteristic – see Chapter 6.3
 QN Quantitative characteristic – see Chapter 6.3
 PQ Pseudo-qualitative characteristic – see Chapter 6.3

4 Method of observation (and type of plot, if applicable)
 MG, MS, VG, VS – see Chapter 4.1.5

5 (+) See Explanations on the Table of Characteristics in Chapter 8.1

6 Not applicable

7 Growth stage key See Explanations on the Table of Characteristics in Chapter 8.2

A sample size of 100 plants/parts of plants – see Chapter 4.2.3

B sample size of 200 plants – see Chapter 4.2.3

7. Table of Characteristics/Tableau des caracteres/Merkmalstabelle/Tabla de caracteres

		English		français		deutsch		español		Example Varieties Exemples Beispielssorten Variedades ejemplo		Note/ Nota
1.	(*)	QN	VG	(+)		10-20						
		Germity										
		monogerm								Brick		1
		partly monogerm/ partly multigerm										2
		multigerm								Jaune d'Eckendorf		3
2.	(*)	PQ	MS	(+)		10-20						
		Ploidy										
		diploid								Brick		2
		triploid								Jamon		3
		tetraploid										4
		polyploid										5
3.		PQ	VG/VS	(+)		10-20						
		Hypocotyl: color										
		white										1
		green								Bergman, Perrine		2
		yellow								Bangor, Cerise		3
		orange								Splendide		4
		pink								Brick		5
		red								Sibemol		6
		red purple										7
4.	(*)	QN	VG B	(+)		33-39						
		Leaf: attitude										
		erect										1
		erect to semi-erect								Cerise		2
		semi-erect								Bolero		3
		semi-erect to intermediate										4
		intermediate								Jamon		5
		intermediate to semi-prostrate								Sibemol		6
		semi-prostrate										7
		semi-prostrate to prostrate								Rouge d'Eckendorf		8
		prostrate										9

		English		français		deutsch		español		Example Varieties Exemples Beispielssorten Variedades ejemplo		Note/ Nota
5.	(*)	QN	VG B			33-39						
		Leaf: blistering										
		weak								Brunium		1
		weak to medium								Jamon		2
		medium								Brick		3
		medium to strong								Eloquenta KWS		4
		strong										5
6.		QN	VG B			33-39						
		Leaf: glossiness										
		absent or weak								Perrine		1
		medium								Bergman		2
		strong								Laurena KWS		3
7.		QN	VG B			33-39						
		Leaf: undulation of margin										
		weak								Brunium, Cerise		1
		weak to medium										2
		medium								Perrine		3
		medium to strong								Eloquenta KWS		4
		strong										5
8.	(*)	QN	VG B			33-39						
		Leaf blade: intensity of green color										
		very light										1
		very light to light										2
		light								Lactimo		3
		light to medium										4
		medium								Jamon, Perrine		5
		medium to dark										6
		dark								Laurena KWS		7
		dark to very dark										8
		very dark										9

		English		français		deutsch		español		Example Varieties Exemples Beispielssorten Variedades ejemplo		Note/ Nota
9.	(*)	QN	MS/VG B	(+)		33-39						
		Leaf: length										
		very short										1
		very short to short										2
		short								Eloquenta KWS		3
		short to medium								Bergman, Brunium		4
		medium								Jamon, Laurena KWS		5
		medium to long								Derixia, Girida		6
		long								Bolero		7
		long to very long										8
		very long										9
10.		QN	MS/VG B			33-39						
		Leaf: width										
		very narrow										1
		very narrow to narrow										2
		narrow								Laurena KWS		3
		narrow to medium								Bergman, Girida		4
		medium								Bolero, Brunium		5
		medium to broad								Derixia, Jamon		6
		broad								Géante Rouge		7
		broad to very broad										8
		very broad										9
11.	(*)	QN	MS/VG B			33-39						
		Leaf blade: width in relation to length										
		very narrow										1
		very narrow to narrow										2
		narrow										3
		narrow to medium								Brunium, Jamon		4
		medium								Bergman		5
		medium to broad								Bolero		6
		broad								Girida, Laurena KWS		7
		broad to very broad										8
		very broad										9
12.		QN	VG B	(+)		33-39						
		Leaf blade: shape of apex										
		strongly acute								Laurena KWS, Splendide		1
		right angle								Bolero, Brick, Jamon		2
		obtuse								Cerise, Feldherr, Kokomo		3

		English		français		deutsch		español		Example Varieties Exemples Beispielssorten Variedades ejemplo		Note/ Nota
13.		QN	MS/VG B			33-39						
		Petiole: length										
		very short										1
		very short to short										2
		short								Bolero		3
		short to medium								Enermax		4
		medium								Brick, Perrine		5
		medium to long								Abramo		6
		long								Girida		7
		long to very long										8
		very long										9
14.	(*)	QN	MG/VG B			49						
		Plant: natural height										
		short										1
		short to medium								Brunium		2
		medium								Bolero, Brick		3
		medium to tall								Enermax, Perrine		4
		tall								Abramo		5
15.		QN	MS/VG A	(+)		49						
		Root: position in soil										
		very shallow										1
		very shallow to shallow										2
		shallow								Feldherr		3
		shallow to medium										4
		medium								Tarine		5
		medium to deep								Cerise		6
		deep								Brick, Eloquenta KWS		7
		deep to very deep										8
		very deep										9
16.		PQ	VG A			49						
		Root: color above ground										
		white										1
		green								Brick, Laurena KWS		2
		yellow										3
		orange										4
		red								Kokomo		5
		red purple										6

		English		français		deutsch		español		Example Varieties Exemples Beispielssorten Variedades ejemplo		Note/ Nota
17.	(*)	PQ	VG A	(+)		49						
		Root: shape										
		obloid										1
		ellipsoid										2
		broad obovoid								Aversa		3
		narrow obovoid										4
		narrow obconic								Girida, Perrine		5
		broad obconic										6
		compressed oblong								Bolero		7
		medium oblong										8
		elongated oblong								Jaune d'Eckendorf		9
18.		QN	MS/VG A			49						
		Root: length										
		very short										1
		very short to short										2
		short								Brick		3
		short to medium								Energarci		4
		medium								Cerise		5
		medium to long								Bangor, Ribambelle		6
		long								Géante Blanche		7
		long to very long										8
		very long										9
19.		QN	MS/VG A			49						
		Root: width										
		very narrow										1
		very narrow to narrow										2
		narrow								Géante Blanche		3
		narrow to medium								Bangor		4
		medium								Brick, Cerise		5
		medium to broad								Ribambelle		6
		broad										7
		broad to very broad										8
		very broad										9

		English		français		deutsch		español		Example Varieties Exemples Beispielssorten Variedades ejemplo		Note/ Nota
20.	(*)	QN	MS/VG A			49						
		Root: length in relation to width										
		very short										1
		very short to short										2
		short										3
		short to medium								Brick		4
		medium								Energarci, Ribambelle		5
		medium to long										6
		long								Bangor		7
		long to very long										8
		very long								Géante Blanche		9
21.	(*)	PQ	VG A			49						
		Root: color below ground										
		white								Brick, Laurena KWS		1
		yellowish white										2
		yellow								Cerise		3
		yellow orange								Feldherr		4
		orange								Valence		5
		orange red								Dynamo, Kokomo		6
		red								Caribou		7
		light pink								Tarine		8
		pink								Merveille		9
		red purple										10
22.	(*)	QN	MG/MS	(+)		49						
		Root: dry matter content										
		very low								Rouge d'Eckendorf		1
		very low to low										2
		low										3
		low to medium								Brigadier		4
		medium								Jamon		5
		medium to high								Valence		6
		high								Brunium		7
		high to very high								Enermax, Tarine		8
		very high								Brick, Perrine		9

8. Explanations on the Table of Characteristics

8.1 *Explanations for individual characteristics*

Ad. 1: Germity

Germity should be observed on 100 seeds.

The attribution of notes for state of expressions is as follows:

Note 1 = monogerm with equal or more than 95% of monogerm seeds

Note 2 = partly monogerm/partly multigerm with less than 95% and more than 15% monogerm seeds

Note 3 = multigerm with equal or less than 15% monogerm seeds

For partly monogerm/partly multigerm varieties this characteristic should not be used to establish distinctness.

Ad. 2: Ploidy

Observations should be made by standard cytological methods.

Observations should be made on at least 5 plants. If any off-type is observed in a sample of 5 plants, another 10 plants should be observed.

The state of expression 5 - Polyploid is a mixture of diploids, triploids and tetraploids. For polyploid varieties this characteristic should not be used to establish distinctness

Ad. 3: Hypocotyl: color

Observations should be made on at least 100 seedlings, grown in the greenhouse, when plants are about 5 cm high. The occurrence of more than one color should not be regarded as a lack of uniformity but for varieties with more than one color this characteristic should not be used to establish distinctness.

Ad. 4: Leaf: attitude

Observations should be made from the angle formed by the petiole and the vertical axis through the root.

Ad. 9: Leaf: length

Observation should be made on the largest, fully expanded leaf including the petiole.

Ad. 12: Leaf blade: shape of apex



1
strongly acute



2
right angle

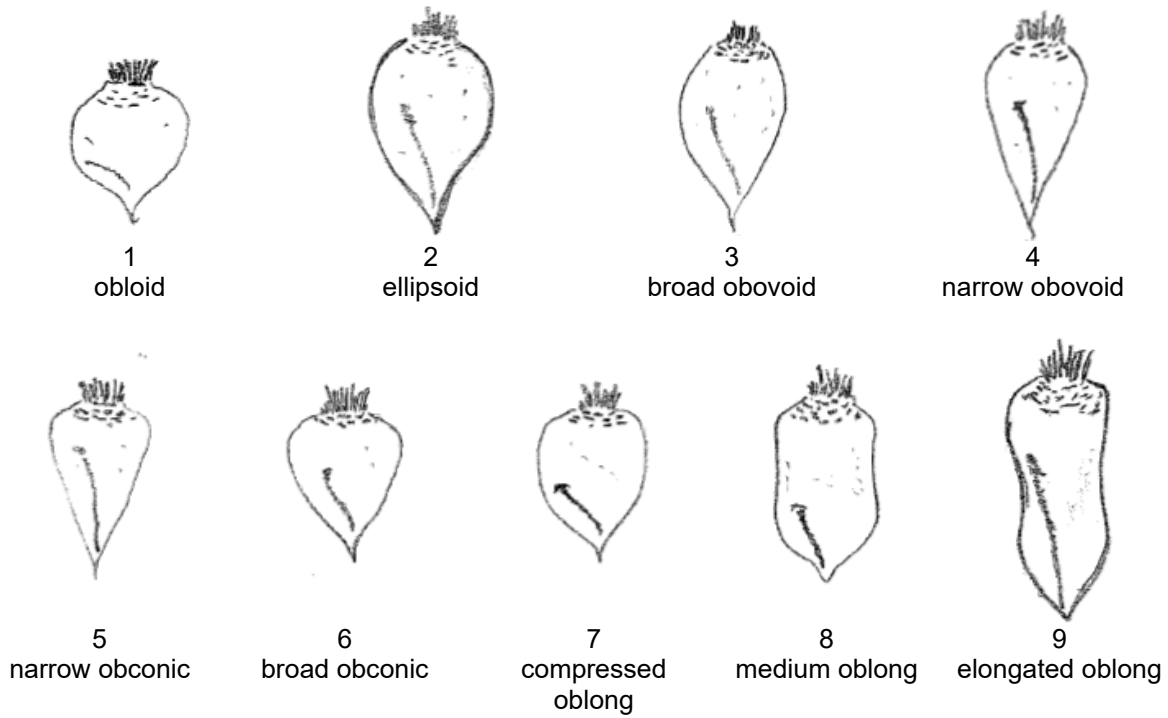


3
obtuse

Ad. 15: Root: position in soil

Observations should be made on the roots by assessing the height of the root above the soil, without harvesting.

Ad. 17: Root: shape



Ad. 22: Root: dry matter content

Observations should be made either on a bulk sample of 30 roots or on a sample of 30 individual roots.

8.2 Additional Explanations on the Table of Characteristic

Growth stage of *Beta vulgaris* L. adapted to the BBCH scale (Meier U., 1993)

Code Description

Principal growth stage 0: Germination

- 00 Dry seed
- 01 Imbibition – seed begins to take up water
- 03 End of seed imbibition – seed coat opened (pellet cracked)
- 05 Radicle emerged from seed
- 07 Shoot emerged from seed (pellet)
- 09 Emergence - shoot emerges at the soil surface

Principal growth stage 1: Leaf development (youth stage)

- 10 Cotyledons horizontally unfolded ; 1st leaf of pin-head-size
- 11 1st pair of leaves visible, of pea-size
- 12 2 leaves (first pair) unfolded
- 14 4 leaves (second pair) unfolded
- 15 5 leaves unfolded
- So on to...
- 19 9 and more leaves unfolded

Principal growth stage 3: Rosette growth (crop cover)

- 30 Beginning of crop cover formation - leaf contact of 10% of plants in adjacent rows
- 33 Contact of 30 % of plants in adjacent rows
- 39 Crop cover complete - contact of more than 90% of plants in adjacent rows

Principal growth stage 4: Development of harvestable vegetative plant parts- Beet-root

- 49 Beet-root has reached harvestable size

9. Literature

Meier, U.; L. Bachmann; H. Buhtz; H. Hack; R. Klose; B. Marlander; E. Weber (1993). "Phänologische Entwicklungsstadien der Beta-Rüben (*Beta vulgaris* L. ssp.). Codierung und Beschreibung nach der erweiterten BBCH-Skala (mit Abbildungen)". *Nachrichtenbl. Deut. Pflanzenschutzd.* 45: 37–41.

10. Technical Questionnaire

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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		Application date: (not to be filled in by the applicant)
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TECHNICAL QUESTIONNAIRE
to be completed in connection with an application for plant breeders' rights

1. Subject of the Technical Questionnaire

1.1 Botanical name

Beta vulgaris L. Fodder Beet Group

1.2 Common name

Fodder beet

2. Applicant

Name

Address

Telephone No.

Fax No.

E-mail address

Breeder (if different from applicant)

3. Proposed denomination and breeder's reference

Proposed denomination (if available)

Breeder's reference

#4. Information on the breeding scheme and propagation of the variety

4.1 Breeding scheme

Variety resulting from:

4.1.1 Crossing

(a) controlled cross []

(please state parent varieties)

(.....) x (.....)

female parent male parent

(b) partially known cross []

(please state known parent variety(ies))

(.....) x (.....)

female parent male parent

(c) unknown cross []

4.1.2 Mutation
(please state parent varieties)

4.1.3 Discovery and development
(please state where and when discovered and how developed)

4.1.4 Other
(Please provide details)

Authorities may allow certain of this information to be provided in a confidential section of the Technical Questionnaire.

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4.2 Method of propagating the variety	[]
4.2.1 Seed-propagated varieties	[]
4.2.2 Other (Please provide details)	[]
<input type="text"/>	

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5. Characteristics of the variety to be indicated (the number in brackets refers to the corresponding characteristic in Test Guidelines; please mark the note which best corresponds).

	Characteristics	Example Varieties	Note
5.1 (1)	Germity		
	monogerm	Brick	1 []
	partly monogerm/ partly multigerm		2 []
	multigerm	Jaune d'Eckendorf	3 []
5.2 (2)	Ploidy		
	diploid	Brick	2 []
	triploid	Jamon	3 []
	tetraploid		4 []
	polyploid		5 []
5.3 (17)	Root: shape		
	obloid		1 []
	ellipsoid		2 []
	broad obovoid	Aversa	3 []
	narrow obovoid		4 []
	narrow obconic	Girida, Perrine	5 []
	broad obconic		6 []
	compressed oblong	Bolero	7 []
	medium oblong		8 []
	elongated oblong	Jaune d'Eckendorf	9 []
5.4 (21)	Root: color below ground		
	white	Brick, Laurena KWS	1 []
	yellowish white		2 []
	yellow	Cerise	3 []
	yellow orange	Feldherr	4 []
	orange	Valence	5 []
	orange red	Dynamo, Kokomo	6 []
	red	Caribou	7 []
	light pink	Tarine	8 []
	pink	Merveille	9 []
	red purple		10 []

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6. Similar varieties and differences from these varieties

Please use the following table and box for comments to provide information on how your candidate variety differs from the variety (or varieties) which, to the best of your knowledge, is (or are) most similar. This information may help the examination authority to conduct its examination of distinctness in a more efficient way

Denomination(s) of variety(ies) similar to your candidate variety	Characteristic(s) in which your candidate variety differs from the similar variety(ies)	Describe the expression of the characteristic(s) for the similar variety(ies)	Describe the expression of the characteristic(s) for your candidate variety
<i>Example</i>	<i>Root: shape</i>	<i>obloid</i>	<i>broad obconic</i>

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<p>Comments</p>

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#7. Additional information which may help in the examination of the variety

7.1 In addition to the information provided in sections 5 and 6, are there any additional characteristics which may help to distinguish the variety?

Yes No

(If yes, please provide details)

7.2 Are there any special conditions for growing the variety or conducting the examination?

Yes No

(If yes, please provide details)

7.3 Other information

Resistance to pests and diseases

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8. Authorization for release

(a) Does the variety require prior authorization for release under legislation concerning the protection of the environment, human and animal health?

Yes [] No []

(b) Has such authorization been obtained?

Yes [] No []

If the answer to (b) is yes, please attach a copy of the authorization.

9. Information on plant material to be examined or submitted for examination

9.1 The expression of a characteristic or several characteristics of a variety may be affected by factors, such as pests and disease, chemical treatment (e.g. growth retardants or pesticides), effects of tissue culture, different rootstocks, scions taken from different growth phases of a tree, etc.

9.2 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If the plant material has undergone such treatment, full details of the treatment must be given. In this respect, please indicate below, to the best of your knowledge, if the plant material to be examined has been subjected to:

- | | | |
|---|---------|--------|
| (a) Microorganisms (e.g. virus, bacteria, phytoplasma) | Yes [] | No [] |
| (b) Chemical treatment (e.g. growth retardant, pesticide) | Yes [] | No [] |
| (c) Tissue culture | Yes [] | No [] |
| (d) Other factors | Yes [] | No [] |

Please provide details for where you have indicated "yes".

10. I hereby declare that, to the best of my knowledge, the information provided in this form is correct:

Applicant's name

Signature

Date

[End of document]