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INTERNATIONAL UNION FOR THE PROTECTION OF NEW VARIETIES OF PLANTS

Geneva

DRAFT

COCKSFOOT

UPOV Code(s): DCTLS_GLO

Dactylis glomerata L.

GUIDELINES

FOR THE CONDUCT OF TESTS

FOR DISTINCTNESS, UNIFORMITY AND STABILITY

prepared by experts from France
to be considered by the
Technical Working Party for Agricultural Crops
at its fifty-first session, to be held in Cambridge, United Kingdom,
from 2022-05-23 to 2022-05-27

Disclaimer: this document does not represent UPOV policies or guidance

Alternative names:*

Botanical name	*English*	*French*	*German*	*Spanish*
Dactylis glomerata L.	Cocksfoot; Orchard Grass	Dactyle	Knaulgras	Dactilo; Pasto ovillo

The purpose of these guidelines ("Test Guidelines") is to elaborate the principles contained in the General Introduction (document TG/1/3), and its associated TGP documents, into detailed practical guidance for the harmonized examination of distinctness, uniformity and stability (DUS) and, in particular, to identify appropriate characteristics for the examination of DUS and production of harmonized variety descriptions.

ASSOCIATED DOCUMENTS

These Test Guidelines should be read in conjunction with the General Introduction and its associated TGP documents.

* These names were correct at the time of the introduction of these Test Guidelines but may be revised or updated. [Readers are advised to consult the UPOV Code, which can be found on the UPOV Website (www.upov.int), for the latest information.]

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1. Subject of these Test Guidelines

These Test Guidelines apply to all varieties of *Dactylis glomerata* L.

2. Material Required

2.1 The competent authorities decide on the quantity and quality of the plant material required for testing the variety and when and where it is to be delivered. Applicants submitting material from a State other than that in which the testing takes place must ensure that all customs formalities and phytosanitary requirements are complied with.

2.2 The material is to be supplied in the form of seeds.

2.3 The minimum quantity of plant material, to be supplied by the applicant, should be:

500 grams.

The seed should meet the minimum requirements for germination, species and analytical purity, health and moisture content, specified by the competent authority.

2.4 The plant material supplied should be visibly healthy, not lacking in vigor, nor affected by any important pest or disease.

2.5 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If it has been treated, full details of the treatment must be given.

3. Method of Examination

3.1 Number of Growing Cycles

3.1.1 The minimum duration of tests should normally be two independent growing cycles.

3.1.2 The two independent growing cycles should be in the form of two separate plantings.

3.1.3 The testing of a variety may be concluded when the competent authority can determine with certainty the outcome of the test.

3.2 Testing Place

Tests are normally conducted at one place. In the case of tests conducted at more than one place, guidance is provided in TGP/9 “Examining Distinctness”.

3.3 Conditions for Conducting the Examination

3.3.1 The tests should be carried out under conditions ensuring satisfactory growth for the expression of the relevant characteristics of the variety and for the conduct of the examination.

3.3.2 The optimum stage of development for the assessment of each characteristic is indicated by a number in the Table of Characteristics. The stages of development denoted by each number are described in Chapter 8.

3.3.3 The recommended type of plot in which to observe the characteristic is indicated by the following key in the Table of Characteristics:

A: spaced plants
B: row plots
C: special tests

3.4 Test Design

3.4.1 Spaced plants: Each test should be designed to result in at least 60 plants, which should be divided between at least 2 replicates.

3.4.2 The test may include 8 meters of row plot which should be divided between at least 2 replicates. The sowing density should be such that around 200 plants per meter can be expected.

3.4.3 The design of the tests should be such that plants or parts of plants may be removed for measurement or counting without prejudice to the observations which must be made up to the end of the growing cycle.

3.5 Additional Tests

Additional tests, for examining relevant characteristics, may be established.

4. Assessment of Distinctness, Uniformity and Stability

4.1 Distinctness

4.1.1 General Recommendations

It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding distinctness. However, the following points are provided for elaboration or emphasis in these Test Guidelines.

4.1.2 Consistent Differences

The differences observed between varieties may be so clear that more than one growing cycle is not necessary. In addition, in some circumstances, the influence of the environment is not such that more than a single growing cycle is required to provide assurance that the differences observed between varieties are sufficiently consistent. One means of ensuring that a difference in a characteristic, observed in a growing trial, is sufficiently consistent is to examine the characteristic in at least two independent growing cycles.

4.1.3 Clear Differences

Determining whether a difference between two varieties is clear depends on many factors, and should consider, in particular, the type of expression of the characteristic being examined, i.e. whether it is expressed in a qualitative, quantitative, or pseudo-qualitative manner. Therefore, it is important that users of these Test Guidelines are familiar with the recommendations contained in the General Introduction prior to making decisions regarding distinctness.

4.1.4 Number of Plants or Parts of Plants to be Examined

Unless otherwise indicated, for the purposes of distinctness, all observations on single plants should be made on 60 plants or parts of plants taken from each of 60 plants and any other observations made on all plants in the test, disregarding any off-type plants.

In the case of observations of parts taken from single plants, the number of parts to be taken from each of the plants should be 1.

4.1.5 Method of Observation

The recommended method of observing the characteristic for the purposes of distinctness is indicated by the following key in the Table of Characteristics (see document TGP/9 “Examining Distinctness”, Section 4 “Observation of characteristics”):

MG: single measurement of a group of plants or parts of plants
MS: measurement of a number of individual plants or parts of plants
VG: visual assessment by a single observation of a group of plants or parts of plants
VS: visual assessment by observation of individual plants or parts of plants

Type of observation: visual (V) or measurement (M)

"Visual" observation (V) is an observation made on the basis of the expert's judgment. For the purposes of this document, "visual" observation refers to the sensory observations of the experts and, therefore, also includes smell, taste and touch. Visual observation includes observations where the expert uses reference points (e.g. diagrams, example varieties, side-by-side comparison) or non-linear charts (e.g. color charts). Measurement (M) is an objective observation against a calibrated, linear scale e.g. using a ruler, weighing scales, colorimeter, dates, counts, etc.

Type of record: for a group of plants (G) or for single, individual plants (S)

For the purposes of distinctness, observations may be recorded as a single record for a group of plants or parts of plants (G), or may be recorded as records for a number of single, individual plants or parts of plants (S). In most cases, "G" provides a single record per variety and it is not possible or necessary to apply statistical methods in a plant-by-plant analysis for the assessment of distinctness.

In cases where more than one method of observing the characteristic is indicated in the Table of Characteristics (e.g. VG/MG), guidance on selecting an appropriate method is provided in document TGP/9, Section 4.2.

4.2 Uniformity

4.2.1 It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding uniformity. However, the following points are provided for elaboration or emphasis in these Test Guidelines:

4.2.2 These Test Guidelines have been developed for the examination of cross-pollinated varieties. For varieties with other types of propagation, the recommendations in the General Introduction and document TGP/13 "Guidance for new types and species" Section 4.5 "Testing Uniformity" should be followed.

4.2.3 The assessment of uniformity should be according to the recommendations for cross-pollinated varieties in the General Introduction.

4.3 Stability

4.3.1 In practice, it is not usual to perform tests of stability that produce results as certain as those of the testing of distinctness and uniformity. However, experience has demonstrated that, for many types of variety, when a variety has been shown to be uniform, it can also be considered to be stable.

4.3.2 Where appropriate, or in cases of doubt, stability may be further examined by testing a new seed stock to ensure that it exhibits the same characteristics as those shown by the initial material supplied.

5. Grouping of Varieties and Organization of the Growing Trial

5.1 The selection of varieties of common knowledge to be grown in the trial with the candidate varieties and the way in which these varieties are divided into groups to facilitate the assessment of distinctness are aided by the use of grouping characteristics.

5.2 Grouping characteristics are those in which the documented states of expression, even where produced at different locations, can be used, either individually or in combination with other such characteristics: (a) to select varieties of common knowledge that can be excluded from the growing trial used for examination of distinctness; and (b) to organize the growing trial so that similar varieties are grouped together.

5.3 The following have been agreed as useful grouping characteristics:

(a) Ploidy (characteristic 1)
(b) Plant: time of inflorescence emergence (characteristic 9)
(c) Stem: length (characteristic 14)

5.4 Guidance for the use of grouping characteristics, in the process of examining distinctness, is provided through the General Introduction and document TGP/9 “Examining Distinctness”.

6. Introduction to the Table of Characteristics

6.1 Categories of Characteristics

6.1.1 Standard Test Guidelines Characteristics

Standard Test Guidelines characteristics are those which are approved by UPOV for examination of DUS and from which members of the Union can select those suitable for their particular circumstances.

6.1.2 Asterisked Characteristics

Asterisked characteristics (denoted by *) are those included in the Test Guidelines which are important for the international harmonization of variety descriptions and should always be examined for DUS and included in the variety description by all members of the Union, except when the state of expression of a preceding characteristic or regional environmental conditions render this inappropriate.

6.2 States of Expression and Corresponding Notes

6.2.1 States of expression are given for each characteristic to define the characteristic and to harmonize descriptions. Each state of expression is allocated a corresponding numerical note for ease of recording of data and for the production and exchange of the description.

6.2.2 All relevant states of expression are presented in the characteristic.

6.2.3 Further explanation of the presentation of states of expression and notes is provided in document TGP/7 “Development of Test Guidelines”.

6.3 Types of Expression

An explanation of the types of expression of characteristics (qualitative, quantitative and pseudo-qualitative) is provided in the General Introduction.

6.4 Example Varieties

Where appropriate, example varieties are provided to clarify the states of expression of each characteristic.

6.5 Legend

	English	français	deutsch	español	Example Varieties Exemples Be ejemplo	Note
1 2	**3 4**	**5 6**	**7**			
	Name of characteristics in English	**Nom du caractère en français**	**Name des Merkmals auf Deutsch**	**Nombre del carácter en español**		
	states of expression	types d'expression	Ausprägungsstufen	tipos de expresión		

1 Characteristic number

2 (*) Asterisked characteristic – see Chapter 6.1.2

3 Type of expression
QL Qualitative characteristic – see Chapter 6.3
QN Quantitative characteristic – see Chapter 6.3
PQ Pseudo-qualitative characteristic – see Chapter 6.3

4 Method of observation (and type of plot, if applicable)
MG, MS, VG, VS – see Chapter 4.1.5

5 (+) See Explanations on the Table of Characteristics in Chapter 8.2

6 (a)-(b) See Explanations on the Table of Characteristics in Chapter 8.1

7 Growth stage key See Explanations on the Table of Characteristics in Chapter 8.3

7. Table of Characteristics/Tableau des caractères/Merkmalstabelle/Tabla de caracteres

		English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
1.	**(*)**	**QL MG|C**	**(+)**				
		Ploidy					
		diploid				Barmedal	2
		tetraploid				Beluga	4
2.		**QN VG|B**		**20-29**			
		Leaf: width					
		very narrow					1
		very narrow to narrow					2
		narrow				Barmedal	3
		narrow to medium					4
		medium				Galibier	5
		medium to broad					6
		broad				Oberweihst, Paykar	7
		broad to very broad					8
		very broad					9
3.		**QN VG|B/VS|A**	**(+)**				
		Plant: tendency to form inflorescences <u>without vernalization</u>					
		absent or very weak				RGT Beverly	1
		very weak to weak					2
		weak				Barmedal, Oberweihst	3
		weak to medium					4
		medium				Bartyle	5
		medium to strong					6
		strong				Bacchus, Inia le Oberon	7
		strong to very strong					8
		very strong					9

	English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
4.	**QN VG|B/VS|A**	**(a)**	**20-29**			
	Plant: growth habit <u>without vernalization</u>					
	erect					1
	erect to semi-erect					2
	semi-erect				Bacchus	3
	semi-erect to intermediate					4
	intermediate				Beluga	5
	intermediate to semi-prostrate					6
	semi-prostrate				Bargère, Priekulu 30	7
	semi-prostrate to prostrate					8
	prostrate				Laban	9
5.	**QN MS|A/VG|B**		**20-29**			
	Plant: natural height <u>without vernalization</u>					
	very short					1
	very short to short					2
	short				Oberweihst	3
	short to medium					4
	medium				Barmedal	5
	medium to tall					6
	tall				Bolide	7
	tall to very tall					8
	very tall					9
6.	**QN VG|B/VS|A**		**20-29**			
	Leaf: intensity of green color <u>without vernalization</u>					
	very light					1
	very light to light					2
	light				Bacchus, Mobite	3
	light to medium					4
	medium				Bargère	5
	medium to dark					6
	dark				Lupré	7
	dark to very dark					8
	very dark					9

		English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
7.		**QN VG|B/VS|A**	**(a)**	**30-39**			
		Plant: growth habit <u>after vernalization</u>					
		erect					1
		erect to semi erect					2
		semi erect				Lucharm	3
		semi erect to intermediate					4
		intermediate					5
		intermediate to semi prostrate					6
		semi prostrate				Ambassador	7
		semi prostrate to prostrate					8
		prostrate				Laban	9
8.		**QN VG|B/VS|A**		**30-39**			
		Leaf: intensity of green color <u>after vernalization</u>					
		very light					1
		very light to light					2
		light				Bacchus, Mobite	3
		light to medium					4
		medium				Bargère, Beluga	5
		medium to dark					6
		dark				Lupré	7
		dark to very dark					8
		very dark					9
9.	**(*)**	**QN MG|B/MS|A**	**(+)**				
		Plant: time of inflorescence emergence					
		very early					1
		very early to early					2
		early				Anksta	3
		early to medium					4
		medium				Coffee, Priekulu 30	5
		medium to late					6
		late				Beluga	7
		late to very late					8
		very late				Lumix	9

		English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
10		**QN MS|A**		**50-56**			
		Plant: natural height <u>at inflorescence emergence</u>					
		very short				Barmedal	1
		very short to short					2
		short				Musketier, Paykar	3
		short to medium					4
		medium				Safin	5
		medium to tall					6
		tall				Galibier	7
		tall to very tall					8
		very tall				Tardi	9
11		**QN VS|A**	**(a)**	**50-56**			
		Plant: growth habit <u>at inflorescence emergence</u>					
		erect					1
		erect to semi erect					2
		semi erect				Ambassador, Beluga	3
		semi erect to intermediate					4
		intermediate				Priekulu 30	5
		intermediate to semi prostrate					6
		semi prostrate					7
		semi prostrate to prostrate					8
		prostrate					9
12	**(*)**	**QN MS|A**	**(+) (b)**	**50-58**			
		Flag leaf: length					
		very short					1
		very short to short					2
		short				Musketier	3
		short to medium					4
		medium				Oberweihst	5
		medium to long					6
		long				Opina	7
		long to very long					8
		very long					9

		English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
13	**(*)**	**QN MS|A**	**(+) (b)**	**50-58**			
		Flag leaf: width					
		very narrow					1
		very narrow to narrow					2
		narrow				Barmedal	3
		narrow to medium					4
		medium				Beluga	5
		medium to broad					6
		broad				Opina	7
		broad to very broad					8
		very broad					9
14	**(*)**	**QN MS|A**	**(+) (b)**	**60-68**			
		Stem: length					
		very short				Barmedal	1
		very short to short					2
		short				Safin, Toscali	3
		short to medium					4
		medium				Dragoner	5
		medium to long					6
		long				Galibier	7
		long to very long					8
		very long					9

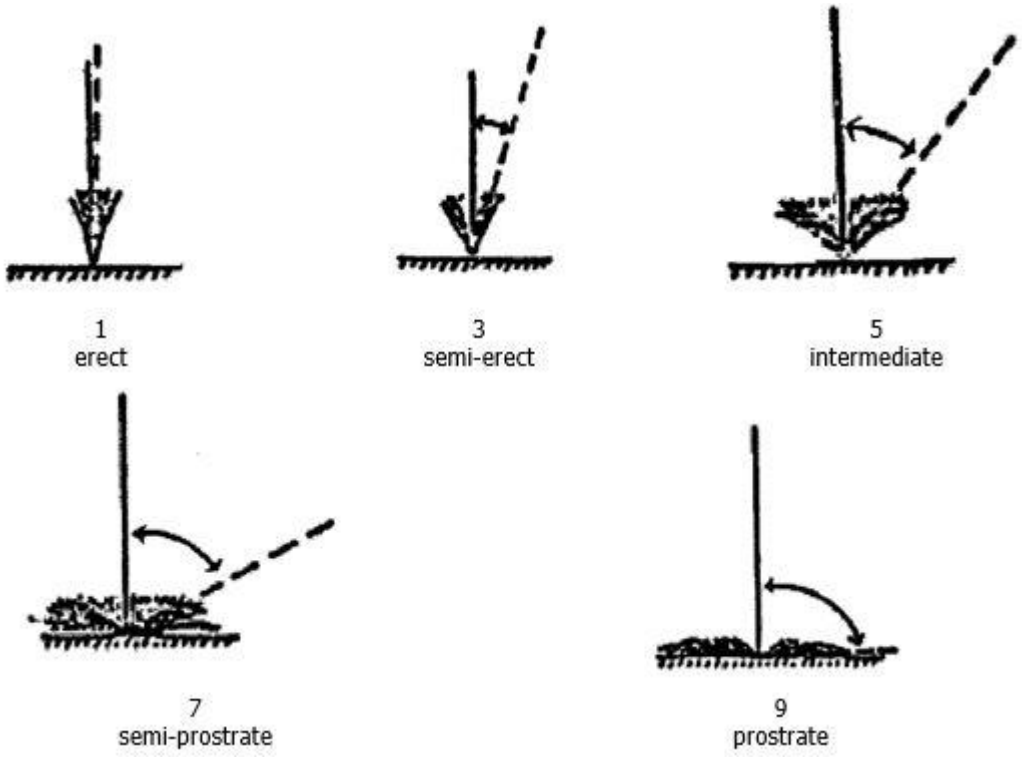
	English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
15	**QN MS|A**	**(+) (b)**	**60-68**			
	Stem: length of upper internode					
	very short					1
	very short to short					2
	short				Bacchus, Safin	3
	short to medium					4
	medium				Dragoner	5
	medium to long					6
	long				Paykar	7
	long to very long					8
	very long					9
16	**QN MS|A**	**(+) (b)**	**60-68**			
	Inflorescence: length					
	very short				Bacchus	1
	very short to short					2
	short				Dragoner, Safin	3
	short to medium					4
	medium				Oberweihst, RGT Beverly	5
	medium to long					6
	long					7
	long to very long					8
	very long					9

8. Explanations on the Table of Characteristics

8.1 Explanations covering several characteristics

Characteristics containing the following key in the Table of Characteristics should be examined as indicated below:

(a) Growth habit
Observations should be made visually from the attitude of the leaves of the plant as a whole. The angle formed by the imaginary line through the region of greatest leaf density and the vertical should be used.



(b) Observations should be made on the longest stem.

8.2 Explanations for individual characteristics

Ad. 1: Ploidy

Ploidy should be assessed by standard cytological methods.

Ad. 3: Plant: tendency to form inflorescences without vernalization

The number of plants showing at least three inflorescences should be recorded for each variety. To be assessed on one occasion on the whole trial when the varieties are judged to have reached their full expression of this characteristic.

Ad. 9: Plant: time of inflorescence emergence

Spaced plants or row plots should be observed at least twice per week.

A: Plots with spaced plants
A single plant is considered to have reached time of inflorescence emergence when the tip of three inflorescences can be seen protruding from the flag leaf sheath (Growth Stage DC 50).

B: Row plots
Time of inflorescence emergence is reached when the average plot stage is DC 54. This date should – if necessary– be obtained by interpolation. At each observation date, the average plot stage should be expressed in one of the following growth stages:

DC 50	First spikelet of inflorescence just visible
DC 52	25% of the inflorescence emerged (across all stems)
DC 54	50% of the inflorescence emerged (across all stems)
DC 56	75% of the inflorescence emerged (across all stems)

Ad. 12: Flag leaf: length

The flag leaf is the first leaf below the inflorescence. Measurements should be made on the same leaf.
Length should be measured from the tip of the leaf blade to the leaf sheath.
Width should be measured at the widest point of the leaf blade.

Ad. 13: Flag leaf: width

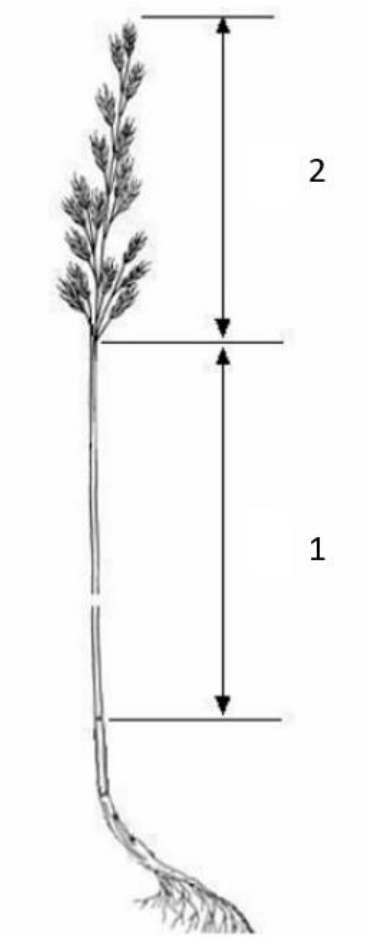
See Ad. 12

Ad. 14: Stem: length

Observations should be made on the longest stem, inflorescence included, in the field from ground level, when the inflorescence is fully expanded.

Ad. 15: Stem: length of upper internode

Char. 15: 1 = The part of the stem above the top node up to the beginning of the inflorescence is the upper internode.
Char. 16: 2 = Length of the inflorescence.



Ad. 16: Inflorescence: length

See Ad. 15

8.3 Explanations on growth stages

All characteristics should be recorded at the appropriate time for the plant concerned. Growth stages of grasses are indicated by decimal codes which are derived from the decimal code for the growth stages of cereals (Zadoks, et al., 1974). This decimal code is in close conformity with the BBCH-code (Meier, 1997).

Seedling growth (seedling: one shoot)

DC 10 First leaf through coleoptile
DC 15 Five leaves unfolded
DC 19 Nine or more leaves unfolded

Tillering

DC 20 Main shoot only (beginning of tillering)
DC 23 Main shoot and 3 tillers
DC 25 Main shoot and 5 tillers
DC 29 Main shoot and 9 or more tillers

Stem elongation

DC 30 Pseudo-stem erection (formed by sheaths of leaves)
DC 31 First node detectable (early stem extension across all stems)
DC 35 Fifth node detectable (50 % extension across all stems)
DC 39 Flag leaf ligula/collar just visible (pre-boot stage)

Booting

DC 41 Flag leaf sheath extending (little enlargement of the inflorescence, early boot-stage)
DC 45 Boots swollen (late-boot stage)
DC 47 First leaf sheath opening
DC 49 first awns visible (in awned forms only)

Inflorescence emergence (mostly non-synchronous)

DC 50 First spikelet of inflorescence just visible
DC 52 25 % of the inflorescence emerged (across all stems)
DC 54 50 % of the inflorescence emerged (across all stems)
DC 56 75 % of the inflorescence emerged (across all stems)
DC 58 Emergence of inflorescence completed

Anthesis (mostly non-synchronous)

DC 60 Beginning of anthesis
DC 64 Anthesis half-way
DC 68 Anthesis complete

9. Literature

Meier, U., 2001: Growth stages of mono- and dicotyledonous plants. BBCH-Monograph, German Federal Biological Research Centre for Agriculture and Forestry.

Zadoks, J. C., Chang, T. T. and Konzak, C. F., 1974: A decimal code for the growth stages of cereals. Weed Research, 14: pp. 415 to 421.

10. Technical Questionnaire

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:

	Application date: (not to be filled in by the applicant)

TECHNICAL QUESTIONNAIRE
to be completed in connection with an application for plant breeders' rights

1. Subject of the Technical Questionnaire

1.1	Botanical name	*Dactylis glomerata* L.
1.2	Common name	Cocksfoot; Orchard Grass

2. Applicant

Name	
Address	
Telephone No.	
Fax No.	
E-mail address	
Breeder (if different from applicant)	

3. Proposed denomination and breeder's reference

Proposed denomination (if available)	
Breeder's reference	

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:

#4. Information on the breeding scheme and propagation of the variety

4.1 Breeding scheme

Variety resulting from:

4.1.1 Crossing

4.1.2 Mutation [ ]
(please state parent variety)

4.1.3 Discovery and development [ ]
(please state where and when discovered and how developed)

4.1.4 Other [ ]
(Please provide details)

Authorities may allow certain of this information to be provided in a confidential section of the Technical Questionnaire.

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:

4.2 Method of propagating the variety

4.2.1 Seed-propagated varieties

(a) Cross-pollination [ ]
(i) Population [ ]
(ii) Synthetic variety [ ]
(b) Other (please provide details) [ ]

4.2.2 Other [ ]
(Please provide details)

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:

5. Characteristics of the variety to be indicated (the number in brackets refers to the corresponding characteristic in Test Guidelines; please mark the note which best corresponds).

	Characteristics	Example Varieties	Note
5.1 (1)	**Ploidy**		
	diploid	Barmedal	2 [ ]
	tetraploid	Beluga	4 [ ]
5.2 (9)	**Plant: time of inflorescence emergence**		
	very early		1 [ ]
	very early to early		2 [ ]
	early	Anksta	3 [ ]
	early to medium		4 [ ]
	medium	Coffee, Priekulu 30	5 [ ]
	medium to late		6 [ ]
	late	Beluga	7 [ ]
	late to very late		8 [ ]
	very late	Lumix	9 [ ]
5.3 (14)	**Stem: length**		
	very short	Barmedal	1 [ ]
	very short to short		2 [ ]
	short	Safin, Toscali	3 [ ]
	short to medium		4 [ ]
	medium	Dragoner	5 [ ]
	medium to long		6 [ ]
	long	Galibier	7 [ ]
	long to very long		8 [ ]
	very long		9 [ ]

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:

6. Similar varieties and differences from these varieties

Please use the following table and box for comments to provide information on how your candidate variety differs from the variety (or varieties) which, to the best of your knowledge, is (or are) most similar. This information may help the examination authority to conduct its examination of distinctness in a more efficient way.

Denomination(s) of variety(ies) similar to your candidate variety	Characteristic(s) in which your candidate variety differs from the similar variety(ies)	Describe the expression of the characteristic(s) for the **similar** variety(ies)	Describe the expression of the characteristic(s) for **your** candidate variety
Example	*Plant: time of inflorescence emergence*	*early*	*medium*

Comments:

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:

[#]7. Additional information which may help in the examination of the variety

7.1 In addition to the information provided in sections 5 and 6, are there any additional characteristics which may help to distinguish the variety?

Yes [ ] No [ ]

(If yes, please provide details)

7.2 Are there any special conditions for growing the variety or conducting the examination?

Yes [ ] No [ ]

(If yes, please provide details)

7.3 Other information

- Resistance to pests and diseases

Authorities may allow certain of this information to be provided in a confidential section of the Technical Questionnaire.

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:

8. Authorization for release

(a) Does the variety require prior authorization for release under legislation concerning the protection of the environment, human and animal health?

Yes [ ] No [ ]

(b) Has such authorization been obtained?

Yes [ ] No [ ]

If the answer to (b) is yes, please attach a copy of the authorization.

9. Information on plant material to be examined or submitted for examination

9.1 The expression of a characteristic or several characteristics of a variety may be affected by factors, such as pests and disease, chemical treatment (e.g. growth retardants or pesticides), effects of tissue culture, different rootstocks, scions taken from different growth phases of a tree, etc.

9.2 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If the plant material has undergone such treatment, full details of the treatment must be given. In this respect, please indicate below, to the best of your knowledge, if the plant material to be examined has been subjected to:

(a)	Microorganisms (e.g. virus, bacteria, phytoplasma)	Yes [ ]	No [ ]
(b)	Chemical treatment (e.g. growth retardant, pesticide)	Yes [ ]	No [ ]
(c)	Tissue culture	Yes [ ]	No [ ]
(d)	Other factors	Yes [ ]	No [ ]

Please provide details for where you have indicated “yes”.

.....

10. I hereby declare that, to the best of my knowledge, the information provided in this form is correct:

Applicant’s name

Signature Date

[End of document]