



TG/276/2(proj.1)

ORIGINAL: English

DATE: 2022-04-04

INTERNATIONAL UNION FOR THE PROTECTION OF NEW VARIETIES OF PLANTS

Geneva

DRAFT

HEMP

UPOV Code(s): CANNB_SAT

Cannabis sativa L.

GUIDELINES

FOR THE CONDUCT OF TESTS

FOR DISTINCTNESS, UNIFORMITY AND STABILITY

*prepared by experts from the Netherlands
to be considered by the
Technical Working Party for Agricultural Crops
at its fifty-first session, to be held in Cambridge, United Kingdom,
from 2022-05-23 to 2022-05-27*

Disclaimer: this document does not represent UPOV policies or guidance

Alternative names:*

<i>Botanical name</i>	<i>English</i>	<i>French</i>	<i>German</i>	<i>Spanish</i>
<i>Cannabis sativa</i> L.	Hemp	Chanvre	Hanf	Cáñamo

The purpose of these guidelines ("Test Guidelines") is to elaborate the principles contained in the General Introduction (document TG/1/3), and its associated TGP documents, into detailed practical guidance for the harmonized examination of distinctness, uniformity and stability (DUS) and, in particular, to identify appropriate characteristics for the examination of DUS and production of harmonized variety descriptions.

ASSOCIATED DOCUMENTS

These Test Guidelines should be read in conjunction with the General Introduction and its associated TGP documents.

* These names were correct at the time of the introduction of these Test Guidelines but may be revised or updated. [Readers are advised to consult the UPOV Code, which can be found on the UPOV Website (www.upov.int), for the latest information.]

TABLE OF CONTENTS	PAGE
1. SUBJECT OF THESE TEST GUIDELINES.....	3
2. MATERIAL REQUIRED.....	3
3. METHOD OF EXAMINATION.....	3
3.1 Number of Growing Cycles.....	3
3.2 Testing Place.....	3
3.3 Conditions for Conducting the Examination.....	4
3.4 Test Design.....	5
3.5 Additional Tests.....	5
4. ASSESSMENT OF DISTINCTNESS, UNIFORMITY AND STABILITY.....	5
4.1 Distinctness.....	5
4.2 Uniformity.....	6
4.3 Stability.....	7
5. GROUPING OF VARIETIES AND ORGANIZATION OF THE GROWING TRIAL.....	8
6. INTRODUCTION TO THE TABLE OF CHARACTERISTICS.....	9
6.1 Categories of Characteristics.....	9
6.2 States of Expression and Corresponding Notes.....	9
6.3 Types of Expression.....	9
6.4 Example Varieties.....	9
6.5 Legend.....	10
7. TABLE OF CHARACTERISTICS/TABLEAU DES CARACTÈRES/MERKMALSTABELLE/TABLA DE CARACTERES.....	11
8. EXPLANATIONS ON THE TABLE OF CHARACTERISTICS.....	24
8.1 Explanations covering several characteristics.....	24
8.2 Explanations for individual characteristics.....	25
9. LITERATURE.....	32
10 TECHNICAL QUESTIONNAIRE.....	33

1. Subject of these Test Guidelines

These Test Guidelines apply to all varieties of *Cannabis sativa* L.

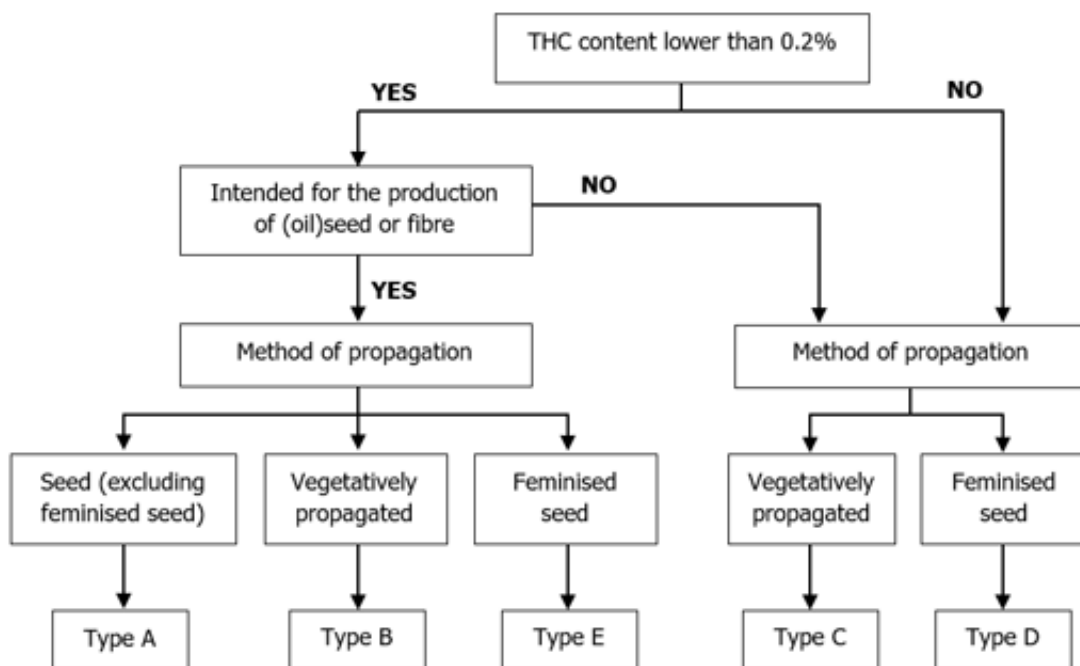
2. Material Required

2.1 The competent authorities decide on the quantity and quality of the plant material required for testing the variety and when and where it is to be delivered. Applicants submitting material from a State other than that in which the testing takes place must ensure that all customs formalities and phytosanitary requirements are complied with.

2.2 The material is to be supplied in the form of seed, feminized seed, or rooted cuttings.

2.3 The minimum quantity of plant material, to be supplied by the applicant, should be:

- Type A (fibre and seed production): 500 gr seeds
- Type B (fibre and seed production): 60 rooted cuttings
- Type E (fibre and seed production): 500 gr of feminized seeds
- Type C (uses other than fibre or seed production): 15 rooted cuttings
- Type D (uses other than fibre or seed production): 500 feminized seeds



In the case of seed, the seed should meet the minimum requirements for germination, species and analytical purity, health and moisture content, specified by the competent authority.

2.4 The plant material supplied should be visibly healthy, not lacking in vigor, nor affected by any important pest or disease.

2.5 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If it has been treated, full details of the treatment must be given.

3. Method of Examination

3.1 *Number of Growing Cycles*

3.1.1 The minimum duration of tests should normally be two independent growing cycles.

3.1.2 The two independent growing cycles should be in the form of two separate plantings.

3.1.3 For types C and D, the minimum duration of tests should normally be a single growing cycle when tests are performed indoors.

3.1.4 The testing of a variety may be concluded when the competent authority can determine with certainty the outcome of the test.

3.2 *Testing Place*

Tests are normally conducted at one place. In the case of tests conducted at more than one place, guidance is provided in TGP/9 "Examining Distinctness".

3.3 *Conditions for Conducting the Examination*

3.3.1 The tests should be carried out under conditions ensuring satisfactory growth for the expression of the relevant characteristics of the variety and for the conduct of the examination.

3.3.2 The optimum stage of development for the assessment of each characteristic is indicated by a number in the Table of Characteristics. The stages of development denoted by each number are described in Chapter 8.

3.4 *Test Design*

3.4.1 In the case of seed and feminized seed propagated varieties (types A and E), each test should be designed to result in a total of at least 200 plants which should be divided between at least 2 replicates.

3.4.2 In the case of vegetatively propagated varieties (type B), each test should be designed to result in a total of at least 60 plants which should be divided between at least 2 replicates.

3.4.3 In the case of vegetatively propagated varieties (type C), each test should be designed to result in a total of at least 10 plants.

3.4.4 In the case of feminized seed propagated varieties (type D), each test should be designed to result in a total of at least 20 plants which should be divided between at least 2 replicates.

3.4.5 The design of the tests should be such that plants or parts of plants may be removed for measurement or counting without prejudice to the observations which must be made up to the end of the growing cycle.

3.5 *Additional Tests*

Additional tests, for examining relevant characteristics, may be established.

4. Assessment of Distinctness, Uniformity and Stability

4.1 *Distinctness*

4.1.1 General Recommendations

It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding distinctness. However, the following points are provided for elaboration or emphasis in these Test Guidelines.

4.1.2 Consistent Differences

The differences observed between varieties may be so clear that more than one growing cycle is not necessary. In addition, in some circumstances, the influence of the environment is not such that more than a single growing cycle is required to provide assurance that the differences observed between varieties are sufficiently consistent. One means of ensuring that a difference in a characteristic, observed in a growing trial, is sufficiently consistent is to examine the characteristic in at least two independent growing cycles.

4.1.3 Clear Differences

Determining whether a difference between two varieties is clear depends on many factors, and should consider, in particular, the type of expression of the characteristic being examined, i.e. whether it is expressed in a qualitative, quantitative, or pseudo-qualitative manner. Therefore, it is important that users of these Test Guidelines are familiar with the recommendations contained in the General Introduction prior to making decisions regarding distinctness.

4.1.4 Number of Plants or Parts of Plants to be Examined

In the case of seed-propagated varieties (types A and E), unless otherwise indicated, for the purposes of distinctness, all observations on single plants should be made on 20 plants or parts taken from each of 20 plants and any other observation made on all plants in the test, disregarding any off-type plants.

In the case of vegetatively propagated varieties (type B), unless otherwise indicated, for the purposes of distinctness, all observations on single plants should be made on 20 plants or parts taken from each of 20 plants and any other observation made on all plants in the test, disregarding any off-type plants.

In the case of vegetatively propagated varieties (type C), unless otherwise indicated, for the purposes of distinctness, all observations on single plants should be made on 5 plants or parts taken from each of 5 plants and any other observation made on all plants in the test, disregarding any off-type plants.

In the case of seed-propagated varieties (feminised seed type D), unless otherwise indicated, for the purposes of distinctness, all observations on single plants should be made on 10 plants or parts taken from each of 10 plants and any other observation made on all plants in the test, disregarding any off-type plants.

4.1.5 Method of Observation

The recommended method of observing the characteristic for the purposes of distinctness is indicated by the following key in the Table of Characteristics (see document TGP/9 “Examining Distinctness”, Section 4 “Observation of characteristics”):

MG: single measurement of a group of plants or parts of plants

MS: measurement of a number of individual plants or parts of plants

VG: visual assessment by a single observation of a group of plants or parts of plants

VS: visual assessment by observation of individual plants or parts of plants

Type of observation: visual (V) or measurement (M)

“Visual” observation (V) is an observation made on the basis of the expert’s judgment. For the purposes of this document, “visual” observation refers to the sensory observations of the experts and, therefore, also includes smell, taste and touch. Visual observation includes observations where the expert uses reference points (e.g. diagrams, example varieties, side-by-side comparison) or non-linear charts (e.g. color charts). Measurement (M) is an objective observation against a calibrated, linear scale e.g. using a ruler, weighing scales, colorimeter, dates, counts, etc.

Type of record: for a group of plants (G) or for single, individual plants (S)

For the purposes of distinctness, observations may be recorded as a single record for a group of plants or parts of plants (G), or may be recorded as records for a number of single, individual plants or parts of plants (S). In most cases, “G” provides a single record per variety and it is not possible or necessary to apply statistical methods in a plant-by-plant analysis for the assessment of distinctness.

In cases where more than one method of observing the characteristic is indicated in the Table of Characteristics (e.g. VG/MG), guidance on selecting an appropriate method is provided in document TGP/9, Section 4.2.

4.2 Uniformity

4.2.1 It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding uniformity. However, the following points are provided for elaboration or emphasis in these Test Guidelines:

4.2.2 These Test Guidelines have been developed for the examination of cross-pollinated (type A), vegetatively propagated (types B and C), and feminized seed propagated (types D and E) varieties. For varieties with other types of propagation, the recommendations in the General Introduction and document TGP/13 "Guidance for new types and species" Section 4.5 "Testing Uniformity" should be followed.

4.2.3 **Type A:**

If uniformity is assessed by the combined over years uniformity method (COYU) the candidate variety is sufficiently uniform in the respective characteristic if the relative tolerance limit in relation to comparable varieties does not exceed the 1% significance level or less ($p < 0.01$) in a test over two consecutive cycles.

If uniformity is assessed by the relative variance method the candidate variety is sufficiently uniform in the respective characteristic if the relative tolerance limit in relation to comparable varieties does not exceed the threshold limit for relative variance, taking into account the sample size, in a test over either two or three consecutive cycles (e.g. for a sample size of 20 measurements the threshold limit ($P=0.01$) for relative variance is 1.88).

If uniformity is assessed by off-types, a population standard of 3% and an acceptance probability of at least 95% should be applied. In the case of a sample size of 200 plants, 10 off-types are allowed.

Type B

For the assessment of uniformity of vegetatively propagated varieties (type B), a population standard of 1% and an acceptance probability of at least 95% should be applied. In the case of a sample size of 60 plants, 2 off-types are allowed.

Type C

For the assessment of vegetatively propagated varieties (type C), a population standard of 1% and an acceptance probability of at least 95% should be applied. In the case of a sample size of 10 plants, 1 off-type is allowed.

Type D

For the assessment of feminized seed propagated varieties (type D), a population standard of 2% and an acceptance probability of at least 95% should be applied. In the case of a sample size of 20 plants, 2 off-types are allowed.

If uniformity is assessed by the relative variance method the candidate variety is sufficiently uniform in the respective characteristic if the relative tolerance limit in relation to comparable varieties does not exceed the threshold limit for relative variance, taking into account the sample size, in a test over either two or three consecutive cycles (e.g. for a sample size of 10 measurements the threshold limit ($P=0.01$) for relative variance is 2.32).

Type E

For the assessment of feminized seed propagated varieties (type E), a population standard of 2% and an acceptance probability of at least 95% should be applied. In the case of a sample size of 200 plants, 7 off-types are allowed.

If uniformity is assessed by the relative variance method the candidate variety is sufficiently uniform in the respective characteristic if the relative tolerance limit in relation to comparable varieties does not exceed the threshold limit for relative variance, taking into account the sample size, in a test over either two or three consecutive cycles (e.g. for a sample size of 20 measurements the threshold limit ($P=0.01$) for relative variance is 1.88).

4.3 *Stability*

4.3.1 In practice, it is not usual to perform tests of stability that produce results as certain as those of the testing of distinctness and uniformity. However, experience has demonstrated that, for many types of variety, when a variety has been shown to be uniform, it can also be considered to be stable.

4.3.2 Where appropriate, or in cases of doubt, stability may be further examined by testing a new seed or plant stock to ensure that it exhibits the same characteristics as those shown by the initial material supplied.

5. Grouping of Varieties and Organization of the Growing Trial
 - .1 The selection of varieties of common knowledge to be grown in the trial with the candidate varieties and the way in which these varieties are divided into groups to facilitate the assessment of distinctness are aided by the use of grouping characteristics.
 - 5.2 Grouping characteristics are those in which the documented states of expression, even where produced at different locations, can be used, either individually or in combination with other such characteristics: (a) to select varieties of common knowledge that can be excluded from the growing trial used for examination of distinctness; and (b) to organize the growing trial so that similar varieties are grouped together.
 - 5.3 The following have been agreed as useful grouping characteristics:
 - (a) Leaf: number of leaflets (characteristic 5)
 - (b) Central leaflet: width (characteristic 7)
 - (c) Only varieties of type A: Time of male flowering (characteristic 8)
 - (d) Only varieties of types B, C, D and E: Time of female flowering (characteristic 9)
 - (e) Plant: proportion of hermaphrodite plants (characteristic 12)
 - (f) Plant: proportion of female plants (characteristic 13)
 - (g) Plant: proportion of male plants (characteristic 14)
 - (h) Only varieties of types A, B and E: Plant: natural height (characteristic 18)
 - (i) Only varieties of types C and D: Plant: height (characteristic 19)
 - (j) Main stem: color (characteristic 20)
 - (k) Only varieties of types A, B and E: Inflorescence: THC content (characteristic 26)
 - (l) Only varieties of types C and D: Inflorescence: THC content (characteristic 27)
 - (m) Inflorescence: CBD content (characteristic 28)
 - 5.4 Guidance for the use of grouping characteristics, in the process of examining distinctness, is provided through the General Introduction and document TGP/9 "Examining Distinctness".
6. Introduction to the Table of Characteristics
 - 6.1 *Categories of Characteristics*
 - 6.1.1 Standard Test Guidelines Characteristics

Standard Test Guidelines characteristics are those which are approved by UPOV for examination of DUS and from which members of the Union can select those suitable for their particular circumstances.
 - 6.1.2 Asterisked Characteristics

Asterisked characteristics (denoted by *) are those included in the Test Guidelines which are important for the international harmonization of variety descriptions and should always be examined for DUS and included in the variety description by all members of the Union, except when the state of expression of a preceding characteristic or regional environmental conditions render this inappropriate.
 - 6.2 *States of Expression and Corresponding Notes*
 - 6.2.1 States of expression are given for each characteristic to define the characteristic and to harmonize descriptions. Each state of expression is allocated a corresponding numerical note for ease of recording of data and for the production and exchange of the description.
 - 6.2.2 All relevant states of expression are presented in the characteristic.
 - 6.2.3 Further explanation of the presentation of states of expression and notes is provided in document TGP/7 "Development of Test Guidelines".

6.3 *Types of Expression*

An explanation of the types of expression of characteristics (qualitative, quantitative and pseudo-qualitative) is provided in the General Introduction.

6.4 *Example Varieties*

Where appropriate, example varieties are provided to clarify the states of expression of each characteristic.

6.5 *Legend*

English		français		deutsch		español		Example Varieties Exemples Beispielssorten Variedades ejemplo		Note/ Nota
1	2	3	4	5	6	7				
Name of characteristics in English		Nom du caractère en français		Name des Merkmals auf Deutsch		Nombre del carácter en español				
states of expression		types d'expression		Ausprägungsstufen		tipos de expresión				

- 1 Characteristic number
- 2 (*) Asterisked characteristic – see Chapter 6.1.2
- 3 Type of expression
 - QL Qualitative characteristic – see Chapter 6.3
 - QN Quantitative characteristic – see Chapter 6.3
 - PQ Pseudo-qualitative characteristic – see Chapter 6.3
- 4 Method of observation (and type of plot, if applicable)
 - MG, MS, VG, VS – see Chapter 4.1.5
- 5 (+) See Explanations on the Table of Characteristics in Chapter 8.2
- 6 (a)-(c) See Explanations on the Table of Characteristics in Chapter 8.1
- 7 Growth stage key See Explanations on the Table of Characteristics in Chapter 8.3

7. Table of Characteristics/Tableau des caractères/Merkmalstabelle/Tabla de caracteres

	English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
1.	QL VG	(a)				
	Leaf: variegation					
	absent				Aida (type C), Futura 75 (type A)	1
	present				Divina (type C)	9
2.	QN VG	(a)				
	Only varieties with leaf variegation: absent: Leaf: intensity of green color					
	light				Aida (type C), Fibror 79 (type A)	1
	medium				Fedora 17 (type A), Theresa (type C)	2
	dark				Finola (type A), Gill (type C)	3
3.	QN MS/VG	(a), (b)				
	Leaf: length of petiole					
	short				Fibrol (type A), MGC 1013 (type C)	1
	medium				Bedrolite (type C), Divina (type C), Fedora 17 (type A)	2
	long				Carmagnola (type A)	3
4. (*)	QN VG	(a), (b)				
	Leaf: anthocyanin coloration of petiole					
	absent or very weak				Fibrol (type A), Gill (type C)	1
	weak				Ruby (type A), Theresa (type C)	2
	medium				Dioica 88 (type A), Gayle (type C)	3
	strong				M-1337 (type C)	4
	very strong				EVLS 113 (type C), Finola (type A)	5

	English		français		deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
5.	(*)	QN	MS/VG	(+)	(a), (b)			
		Leaf: number of leaflets						
		very few					Bedrolite (type C), MGC 1013 (type C)	1
		few					Aida (type C), Finola (type A)	2
		medium					GRX53 (type D), Usó 31 (type A)	3
		many					Fibror 79 (type A)	4
		very many						5
6.		QN	MS/VG		(a), (b)			
		Central leaflet: length						
		very short					Damato Red (type C)	1
		very short to short					MGC 1013 (type C)	2
		short					Divina (type C)	3
		short to medium						4
		medium					Aida (type C)	5
		medium to long						6
		long					Felina 32 (type A)	7
		long to very long						8
		very long					Carmagnola (type A)	9
7.		QN	MS/VG		(a), (b)			
		Central leaflet: width						
		very narrow						1
		very narrow to narrow					Celeste (type C)	2
		narrow					MGC 1013 (type C)	3
		narrow to medium						4
		medium					Fibrol (type A), Theresa (type C)	5
		medium to broad					Hulkberry (type C)	6
		broad					Gill (type C), Usó 31 (type A)	7
		broad to very broad						8
		very broad					Carmagnola (type A), Enectabis (type D)	9

	English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
8. (*)	QN	MG/VG	(+)			
	Only varieties of type A: Time of male flowering					
	very early				Uso 31 (type A)	1
	very early to early					2
	early					3
	early to medium					4
	medium				Fibrol (type A)	5
	medium to late					6
	late				Felina 32 (type A)	7
	late to very late					8
	very late				Dioica 88 (type A)	9
9.	QN	MG/VG	(+)			
	Only varieties of types B, C, D and E: Time of female flowering					
	very early				Celeste (type C)	1
	very early to early					2
	early				Theresa (type C)	3
	early to medium					4
	medium				M-1337 (type C)	5
	medium to late					6
	late				Goya (type C)	7
	late to very late					8
	very late				HURV2019PL (type D E)	9
10	QN	VG		2102 2304		
	Only varieties of type A: Inflorescence: anthocyanin coloration of male flowers					
	absent or very weak				Santhica 27 (type A)	1
	very weak to weak					2
	weak				Uso 31 (type A)	3
	weak to medium					4
	medium				Felina 32 (type A)	5
	medium to strong					6
	strong				Adzelveisi (type A)	7
	strong to very strong				Finola (type A)	8
	very strong					9

	English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
11	QN	VG	(+)	2202b 2302b			
	Only varieties of types B, C, D and E: Inflorescence: anthocyanin coloration of female flowers						
	absent or weak					Aida (type C)	1
	medium						2
	strong					HURV2019PL (type D E)	3
12 (*)	QN	MS/VG	(+)	2102 2202 2302 2304			
	Plant: proportion of hermaphrodite plants						
	low						1
	low to medium						2
	medium						3
	medium to high						4
	high						5
13 (*)	QN	MS/VG	(+)	2102 2202 2302 2304			
	Plant: proportion of female plants						
	low						1
	low to medium						2
	medium						3
	medium to high						4
	high						5
14 (*)	QN	MS/VG	(+)	2102 2202 2302 2304			
	Plant: proportion of male plants						
	low						1
	low to medium						2
	medium						3
	medium to high						4
	high						5

	English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
15	QN	VG		2202b 2302b		
	Only varieties of types C and D: Flower: length of stigmas					
	short				EVLS 113 (type C)	1
	medium				Divina (type C)	2
	long				Bedrobinol (type C), HURV2019PL (type D E)	3
16	QN	VG		2202b 2302b		
	Only varieties of types C and D: Flower: thickness of stigmas					
	thin				HURV2019CBG (type C)	1
	medium				Divina (type C)	2
	thick				HURV2019PL (type D E)	3
17	QN	VG	(+)	2202b 2302b		
	Only varieties of types C and D: Flower: contortion of stigmas					
	absent or weak				Aida (type C)	1
	medium				HURV2019PL (type D E), MGC 1008 (type C)	2
	strong				MGC 1009 (type C)	3
18 (*)	QN	MG/VG	(+)	2202 2202b 2302 2302b		
	Only varieties of types A, B and E: Plant: natural height					
	very short				Adzelveisi (type A), Finola (type A)	1
	very short to short					2
	short					3
	short to medium					4
	medium				Usó 31 (type A)	5
	medium to long				Fibrol (type A)	6
	long				Felina 32 (type A)	7
	long to very long				Fibror 79 (type A)	8
	very long				Dioica 88 (type A)	9

	English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
19	QN	MG/VG	(+)	2202b 2302b		
	Only varieties of types C and D: Plant: height					
	very short				MGC 1027 (type C)	1
	very short to short					2
	short				Chuy (type C)	3
	short to medium					4
	medium				Aida (type C)	5
	medium to long					6
	long				Bedrolite (type C), EVLS 113 (type C)	7
	long to very long				Obi (type D)	8
	very long					9
20 (*)	PQ	VG	(c)	2202 2202b 2302 2303b		
	Main stem: color	Tige principale : couleur	Haupttrieb: Farbe	Tallo principal: color		
	yellow	jaune	gelb	amarillo	Fibror 79 (type A)	1
	medium green	vert moyen	mittelgrün	verde medio	Bedrobinol (type C), Felina 32 (type A), Theresa (type C)	2
	dark green	vert foncé	dunkelgrün	verde oscuro	Aida (type C), Dioica 88 (type A)	3
	purple	pourpre	purpurn	púrpura	EVLS 113 (type C), Fibranova (type A)	4
21	QN	MS/VG	(c)	2202 2202b 2302 2303b		
	Only varieties of types A, B and E: Main stem: length of internode					
	very short				Finola (type A)	1
	very short to short					2
	short					3
	short to medium					4
	medium				Uso 31 (type A)	5
	medium to long					6
	long				Futura 75 (type A)	7
	long to very long					8
	very long					9

	English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
22	QN	MS/VG	(c)	2202b 2302b		
	Only varieties of types C and D: Main stem: length of internode					
	very short					1
	very short to short				MGC 1027 (type C)	2
	short				Beatriz (type C), Divina (type C)	3
	short to medium					4
	medium				Aida (type C), HURV2019PL (type D E)	5
	medium to long				EVLS 113 (type C)	6
	long					7
	long to very long					8
	very long				Enectitaca (type D), Obi (type D)	9
23	QN	MS/VG	(c)	2202 2202b 2302 2303b		
	Only varieties of types A, B and E: Main stem: thickness					
	thin				Finola (type A)	1
	medium				Futura 75 (type A)	2
	thick				Dioica 88 (type A)	3
24	QN	MS/VG	(c)	2202b 2303b		
	Only varieties of types C and D: Main stem: thickness					
	thin				Celeste (type C)	1
	medium				Aida (type C)	2
	thick				Obi (type D)	3
25	QN	VG	(c)	2202 2202b 2302 2303b		
	Main stem: depth of grooves					
	shallow				Divina (type C), Finola (type A), Gill (type C)	1
	medium				Bedrolite (type C), Fedora 17 (type A), Theresa (type C)	2
	deep				Dioica 88 (type A), HURV2019PL (type D E)	3

	English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
26	(*)	QN	MG	(+)	2204 2204b 2305 2305b		
		Only varieties of types A, B and E: Inflorescence: THC content					
		absent or very low				Santhica 27 (type A)	1
		very low to low				Fedora 17 (type A)	2
		low				Futura 75 (type A)	3
27	(*)	QN	MG	(+)	2204b 2305b		
		Only varieties of types C and D: Inflorescence: THC content					
		absent or very low				Aida (type C)	1
		very low to low				Bedrolite (type C), Sara (type C)	2
		low					3
		low to medium				Beatriz (type C)	4
		medium				Bediol (type C)	5
		medium to high				Bedrobinol (type C), Raquel (type C)	6
		high				Bedrocan (type C), GRX53 (type D)	7
		high to very high				Original Blitz (type C)	8
		very high					9
28	(*)	QN	MG	(+)	2204 2204b 2305 2305b		
		Inflorescence: CBD content					
		absent or very low				Bedrobinol (type C), Bedrocan (type C), Enectacalm (type D), Raquel (type C), Santhica 27 (type A)	1
		very low to low				Fedora 17 (type A)	2
		low				Aida (type C), Futura 75 (type A), Theresa (type C)	3
		low to medium				Chuy (type C)	4
		medium				Bediol (type C), Divina (type C)	5
		medium to high				Sibari (type C)	6
		high				Goya (type C)	7
		high to very high				A1 Philadelphia (type C), Enectonica (type D)	8
		very high					9

	English		français		deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
29	QN	VG	(+)	(c)	2204 2202b 2306 2306b			
	Main stem: pith in cross-section							
	absent or thin						HURV2019PL (type D E), Santhica 27 (type A)	1
	medium						Divina (type C), Fedora 17 (type A)	2
	thick						Finola (type A), Gill (type C), MGC 1009 (type C)	3
30	QN	MG			2205 2307			
	Seed: 1,000 seed weight							
	very low						Finola (type A)	1
	low						Chamaeleon (type A), Enectitaca (type D)	2
	medium						Enectacalm (type D), Felina 32 (type A)	3
	high						Santhica 27 (type A)	4
	very high						Fibror 79 (type A)	5
31	PQ	VG			2205 2307			
	Seed: color of testa							
	light grey						Finola (type A)	1
	medium grey						Enectavio (type D), Usó 31 (type A)	2
	grey brown						Enectacalm (type D), Fedora 17 (type A)	3
	yellowish brown						Fibror 79 (type A)	4
	brown						Dioica 88 (type A), Enectitaca (type D)	5
32	QN	VG	(+)		2205 2307			
	Seed: marbling	Graine : marbrure	Samen: Marmorierung	Semilla: veteado				
	weak	faible	gering	débil			Enectacalm (type D), Finola (type A)	1
	medium	moyenne	mittel	medio			Enectavio (type D), Felina 32 (type A)	2
	strong	forte	stark	fuerte			Dioica 88 (type A)	3

8. Explanations on the Table of Characteristics

8.1 *Explanations covering several characteristics*

Characteristics containing the following key in the Table of Characteristics should be examined as indicated below:

- (a) Observations should be done in the period between the beginning of flowering (growth stage 2101, 2201 or 2301, whichever is earliest) and the beginning of seed maturity (Type A) or flower senescence (Types B, C, D and E).
- (b) For type A, observations should be done on the last opposite, fully expanded leaves. For Types B, C, D and E observations should be done on fully developed leaves from the centre of the plant.
- (c) For type A, observations should be done on the internode below the last opposite leaves of female and/or hermaphrodite plants only. In case of types B, C, D and E, observations should be done on the internode below a fully developed leaf from the centre of the plant.

8.2 *Explanations for individual characteristics*

Ad. 5: Leaf: number of leaflets

The predominant number of leaflets in the centre of the plant should be observed:

- 1 - very few = three leaflets or less
- 2 - few = five leaflets
- 3 - medium = seven leaflets
- 4 - many = nine leaflets
- 5 - very many = eleven leaflets or more

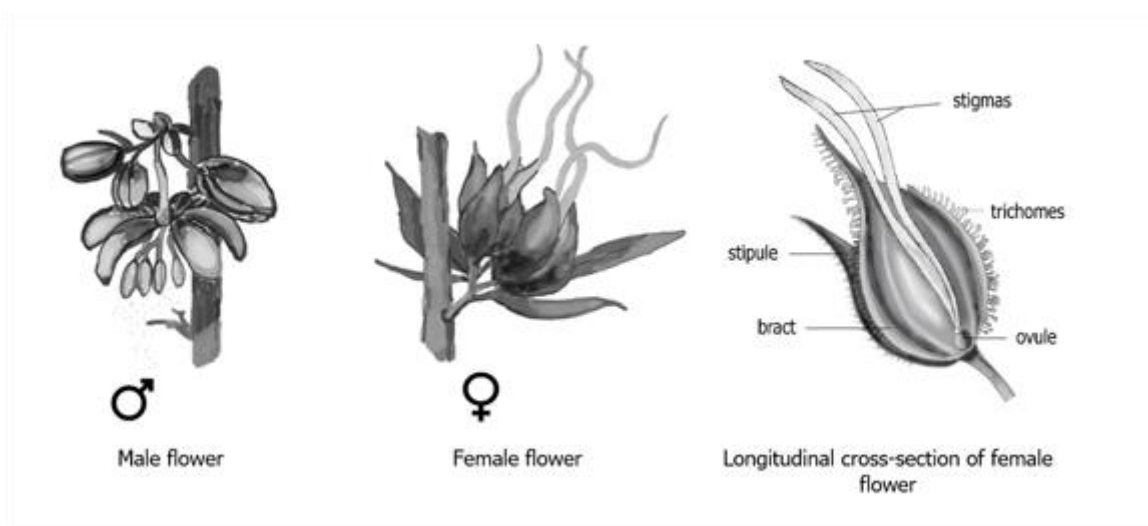
Ad. 8: Only varieties of type A: Time of male flowering

Monoecious varieties: 50 % of all plants with first male flower open

Other varieties: 50 % of all male plants with first male flower open

First male flowers mostly appear from the axils of the leaves on the main stem. Male flowers usually appear about 2 weeks before the stigmas of female flowers are visible.

***Cannabis* flowers:**



Ad. 9: Only varieties of types B, C, D and E: Time of female flowering

Vegetatively propagated and feminized seed varieties: 50% of plants with first stigmas visible

Ad. 11: Only varieties of types B, C, D and E: Inflorescence: anthocyanin coloration of female flowers

The colour of the bracts, stipules and sugarleaves³ should be observed.

³) Sugarleaves are the leaves between the clusters of female flowers.



Ad. 12: Plant: proportion of hermaphrodite plants

Type A:

Cannabis sativa L. is dioecious by nature, containing approximately equal proportions of male and female plants. Hermaphrodite plants (male and female flowers on one plant) occasionally occur naturally but are specially created by breeding activity (Bócsa, 1998). Several intersexual forms exist and sex expression can be modified by environmental factors.

Hermaphrodite plants: plants with both male and female flowers

Female plants: plants with female flowers only

Male plants: plants with male flowers only

Proportion	Note	Ranges (percentage)
low	1	<= 5 %
low to medium	2	6-35 %
medium	3	36-65 %
medium to high	4	66-95 %
high	5	>= 96 %

The proportion should be based on at least 200 plants.

Types B and C:

Vegetatively propagated varieties should show only one type of sex expression.

Types D and E:

Feminized seed varieties may be female, hermaphrodite or may show a mixture of female and hermaphrodite plants. Sex expression is affected by environmental conditions and stress.

Ad. 13: Plant: proportion of female plants

See Ad. 12

Ad. 14: Plant: proportion of male plants

See Ad. 12

Ad. 17: Only varieties of types C and D: Flower: contortion of stigmas



Ad. 18: Only varieties of types A, B and E: Plant: natural height

Plant height should be observed on female and/or hermaphrodite plants from soil level to top of the plant including inflorescence.

Ad. 19: Only varieties of types C and D: Plant: height

See Ad. 18

Ad. 26: Only varieties of types A, B and E: Inflorescence: THC content

The method to determine the THC and CBD content is based on a quantitative determination of Δ^9 -tetrahydrocannabinol (THC) and cannabidiol (CBD) by gas chromatography after extraction with a suitable solvent.

Sampling

The sample should be taken from the upper 30 cm of the main stem, containing well developed female inflorescences.

Types A, B and E: a mixture of 20 plants

Type C: a mixture of 5 plants

Type D: a mixture of 10 plants

(Sugar-)leaves should be removed as much as possible.

The sample should be dried as soon as possible (within 48 hours) at a temperature below 70° C. Samples should be dried to a constant weight and to a moisture content of 8 – 13 %. After drying samples can be stored (without crushing) at below 25° C in a dark place.

Determination of THC/CBD content (Adapted from: Commission Delegated Regulation (EU) No 639/2014 annex II (latest amended version)).

1. Preparation of the test sample

Remove stems and seeds over 2 mm in size from the dried samples.

Grind the dried samples to obtain a semi-fine powder (passing through a 1 mm mesh sieve).

The powder may be stored for 10 weeks at below 25° C in a dark dry place.

2. Reagents and extraction solution

Reagents:

- Δ^9 -tetrahydrocannabinol (THC), pure for chromatographic purposes.
- Cannabidiol (CBD), pure for chromatographic purposes
- squalane, pure for chromatographic purposes, as an internal standard.

Extraction solution:

- 35 mg of squalane per 100 ml hexane.

3. Extraction of cannabinoids

Weigh 100 mg of the powdered test sample, place in a centrifuge tube and add 5 ml of extraction solution containing the internal standard.

Place in an ultrasound bath and leave for 20 minutes. Centrifuge for 5 minutes at 3,000 r.p.m. and then remove the supernatant cannabinoid solution. Inject the solution into the chromatograph and carry out a quantitative analysis.

4. Gas chromatography

(a) Apparatus

- gas chromatograph with a flame ionization detector and a split/splitless injector

- column allowing good separation of cannabinoids, for example a glass capillary column 25 m long and 0.22 mm in diameter impregnated with a 5 % non-polar phenyl-methyl-siloxane phase.

(b) Calibration ranges

At least three points including points 0.04 and 0.50 mg/ml of each of the cannabinoids in extraction solution.

(c) Experimental conditions

The following conditions are given as an example for the column referred to in a).

oven temperature	260° C
injector temperature	300° C
detector temperature	300° C

(d) Injection volume: 1 μ l

Results

THC and CBD should be determined to two decimals in grams of Δ^9 -THC and CBD resp., per 100 grams of analytical sample dried to constant weight. A tolerance of 0.03 g per 100 grams applies.

Although varietal differences for THC and CBD content are consistent, absolute levels of THC and CBD content are sensitive to environmental variation.

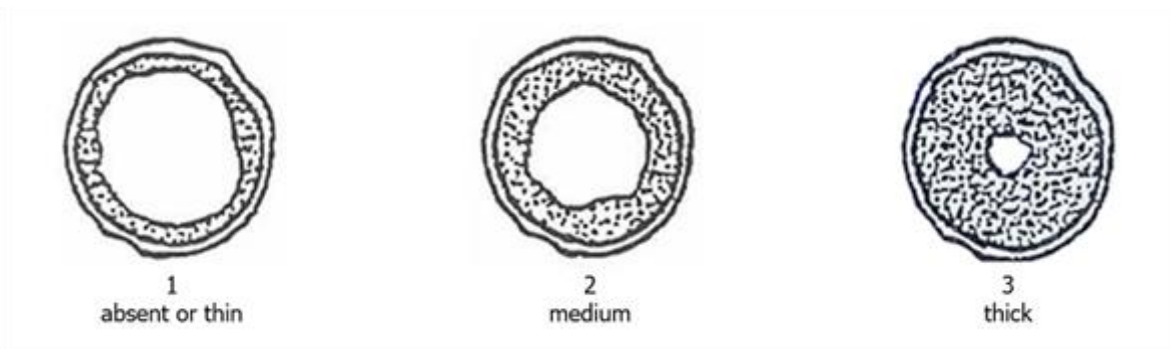
Ad. 27: Only varieties of types C and D: Inflorescence: THC content

See Ad. 26

Ad. 28: Inflorescence: CBD content

See Ad. 26

Ad. 29: Main stem: pith in cross-section



Ad. 32: Seed: marbling

Marbling of testa: black mosaic patterns



8.3 Growth stages

All characteristics should be recorded at the appropriate time of plant development concerned. Growth stages of hemp are recorded by a four-digit code describing the principal growth stages, depending on the sex of the plant followed by detailed developmental stages (Mediavilla, Vito *et al.*, 1998). This growth scale is slightly modified by adding definitions of certain stages (marked by *), to accommodate types B, C, D and E when no seed is formed. Seed formation affects the production of cannabinoids like THC and CBD and is therefore avoided for types C and D. Stages with the same number indicate the same growth stage (e.g. 1006=1006b).

Principal growth stages

Four principal stages describe the life cycle of a plant and are coded by the first digit of the four-digit code.

First-digit of code	Definition
0	Germination and emergence
1	Vegetative stage
2	Flowering and seed formation
3	Senescence

Secondary growth stages

The secondary growth stages are described by the second digit, which indicates the sex of the plant, and the third and fourth digit indicating the developmental stage of the plant.

Code	Definition	Remarks
Germination and emergence		
0000	Dry seed	
0003	Cotyledons unfolded	
Vegetative stage refers to main stem. Leaves are considered unfolded when leaflets are at least one cm long		
1002	1 st leaf pair	1 leaflet
1004	2 nd leaf pair	3 leaflets
1006	3 rd leaf pair	5 leaflets
1006b*	5 th leaf	In vegetatively propagated plants all leaves have the same phyllotaxis
10xx	Last opposite leaf pair	xx = 2 times n th leaf pair
Flowering and seed formation refers to the main stem including branches		
2000	GV point (i.e. induction of flowering)	Change of phyllotaxis on the main stem from opposite to alternate. Distance between petioles of alternate leaves at least 0.5 cm. In vegetatively propagated plants the GV point is absent, all leaves have the same phyllotaxis.
2001	Flower primordia	Sex nearly distinguishable
Male Plant		
2100	Flower formation	First closed staminate flowers
2101	Beginning of flowering	First opened staminate flowers
2102	Flowering	50 % opened staminate flowers
2103	End of flowering	95 % of staminate flowers opened or withered
Female Plant		
2200	Flower formation	First pistillate flowers Bract with no stigmas
2201	Beginning of flowering	Stigmas on first female flowers
2202	Flowering	50 % of bracts formed
2202b*	Flowering	50% of stigmas fully extended
2203	Beginning of seed maturity	First seeds hard
2203b*	Flowers senescence	10% of stigmas brown
2204	Seed maturity	50 % of seeds hard
2204b*	End of flowering	50% of stigmas brown
2205	End of seed maturity	95 % of seeds hard or shattered

	Hermaphrodite plant	
2300	Female flower formation	First pistillate flowers. Perigonal bracts with no stigmas
2301	Beginning of female flowering	First stigmas visible
2302	Female flowering	50 % of bracts formed
2302b*	Female flowering	50% of stigmas fully extended
2303	Male flower formation	First closed staminate flowers
2304	Male flowering	50 % opened staminate flowers
2305	Beginning of seed maturity	First seeds hard
2305b*	Female flower senescence	10% of stigmas brown
2306	Seed maturity	50 % of seeds hard
2306b*	End of female flowering	50% of stigmas brown
2307	End of seed maturity	95 % of seeds hard or shattered
	Senescence	
3001	Leaf desiccation	Leaves dry
3002	Stem desiccation	Leaves dropped
3003	Stem decomposition	Bast fibres free

9. Literature

- Bócsa, I., 1998: Genetic Improvement: Conventional Approaches. In: Advances in Hemp Research. Paolo Ranalli (Ed.). Haworth Food Products Press, New York. 272 pp.
- Bredemann, G., 1922 : Die Bestimmung des Fasergehaltes in Bastfaserpflanzen bei züchterischen Untersuchungen. Faserforschung 2. Leipzig: Hirzel Verlag. S. 239-258.
- Clarke, R.C., 1998: Botany of the Genus *Cannabis*. In: Advances in Hemp Research. Paolo Ranalli (Ed.). Haworth Food Products Press, New York. 272 pp.
- Cole, M.D., 2003: The analysis of controlled substances – a systematic approach. John Wiley and Sons Ltd., Chichester, UK. ISBN 0-471-49252-3.
- Mediavilla, V., Jonquera, M., Schmid-Slembrouck, I., Soldati, A., 1998. Decimal code for growth stages of hemp (*Cannabis sativa* L.). Journal of the International Hemp Association 5(2) 67-72.
- Meijer de, E.P.M., 1994: Diversity in Cannabis. Thesis Wageningen University, ISBN 90-5485-338-7: 131 pp
- Meijer de, E.P.M., 1995: Fibre hemp cultivars: A survey of origin, ancestry, availability and brief agronomic characteristics. Journal of the International Hemp Association 2(2): 66-73
- Meijer de, E.P.M., 1998: Cannabis Germplasm Resources. In: Advances in Hemp Research. Paolo Ranalli (Ed.). Haworth Food Products Press, New York. 272 pp.

10. Technical Questionnaire

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
		Application date: (not to be filled in by the applicant)
TECHNICAL QUESTIONNAIRE to be completed in connection with an application for plant breeders' rights		
1. Subject of the Technical Questionnaire		
1.1	Botanical name	<input type="text" value="Cannabis sativa L."/>
1.2	Common name	<input type="text" value="Hemp"/>
2. Applicant		
	Name	<input type="text"/>
	Address	<input type="text"/>
	Telephone No.	<input type="text"/>
	Fax No.	<input type="text"/>
	E-mail address	<input type="text"/>
	Breeder (if different from applicant)	<input type="text"/>
3. Proposed denomination and breeder's reference		
	Proposed denomination (if available)	<input type="text"/>
	Breeder's reference	<input type="text"/>

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
-------------------------	-----------------	-------------------

#4. Information on the breeding scheme and propagation of the variety

4.1 Breeding scheme

Variety resulting from:

4.1.1 Crossing

(a) controlled cross
(please state parent variety)

(.....) x (.....)

female parent male parent

(b) partially known cross
(please state known parent variety(ies))

(.....) x (.....)

female parent male parent

(c) unknown cross

4.1.2 Mutation
(please state parent variety)

4.1.3 Discovery and development
(please state where and when discovered and how developed)

4.1.4 Other
(Please provide details)

Authorities may allow certain of this information to be provided in a confidential section of the Technical Questionnaire.

4.2 Method of propagating the variety

4.2.1 Seed-propagated varieties

- (a) Cross-pollination []
- (b) Hybrid []
- (c) Other (please provide details) []

4.2.2 Vegetative propagation

- (a) Cuttings []
- (b) *In vitro* propagation []
- (c) Other (state method) []

4.2.3 Other []
(Please provide details)

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
-------------------------	-----------------	-------------------

5. Characteristics of the variety to be indicated (the number in brackets refers to the corresponding characteristic in Test Guidelines; please mark the note which best corresponds).

Characteristics	Example Varieties	Note
5.1 Leaf: number of leaflets (5)		
very few	Bedrolite (type C), MGC 1013 (type C)	1 []
few	Aida (type C), Finola (type A)	2 []
medium	GRX53 (type D), Uso 31 (type A)	3 []
many	Fibror 79 (type A)	4 []
very many		5 []
5.2 Central leaflet: width (7)		
very narrow		1 []
very narrow to narrow	Celeste (type C)	2 []
narrow	MGC 1013 (type C)	3 []
narrow to medium		4 []
medium	Fibrol (type A), Theresa (type C)	5 []
medium to broad	Hulkberry (type C)	6 []
broad	Gill (type C), Uso 31 (type A)	7 []
broad to very broad		8 []
very broad	Carmagnola (type A), Enectabis (type D)	9 []
5.3 Only varieties of type A: Time of male flowering (8)		
very early	Uso 31 (type A)	1 []
very early to early		2 []
early		3 []
early to medium		4 []
medium	Fibrol (type A)	5 []
medium to late		6 []
late	Felina 32 (type A)	7 []
late to very late		8 []
very late	Dioica 88 (type A)	9 []

Characteristics	Example Varieties	Note
5.4 Only varieties of types B, C, D and E: Time of female flowering (9)		
very early	Celeste (type C)	1 []
very early to early		2 []
early	Theresa (type C)	3 []
early to medium		4 []
medium	M-1337 (type C)	5 []
medium to late		6 []
late	Goya (type C)	7 []
late to very late		8 []
very late	HURV2019PL (type D E)	9 []
5.5 Plant: proportion of hermaphrodite plants (12)		
low		1 []
low to medium		2 []
medium		3 []
medium to high		4 []
high		5 []
5.6 Plant: proportion of female plants (13)		
low		1 []
low to medium		2 []
medium		3 []
medium to high		4 []
high		5 []
5.7 Plant: proportion of male plants (14)		
low		1 []
low to medium		2 []
medium		3 []
medium to high		4 []
high		5 []

Characteristics	Example Varieties	Note
5.8 Only varieties of types A, B and E: Plant: natural height (18)		
very short	Adzelveisi (type A), Finola (type A)	1 []
very short to short		2 []
short		3 []
short to medium		4 []
medium	Usa 31 (type A)	5 []
medium to long	Fibrol (type A)	6 []
long	Felina 32 (type A)	7 []
long to very long	Fibror 79 (type A)	8 []
very long	Dioica 88 (type A)	9 []
5.9 Only varieties of types C and D: Plant: height (19)		
very short	MGC 1027 (type C)	1 []
very short to short		2 []
short	Chuy (type C)	3 []
short to medium		4 []
medium	Aida (type C)	5 []
medium to long		6 []
long	Bedrolite (type C), EVLS 113 (type C)	7 []
long to very long	Obi (type D)	8 []
very long		9 []
5.10 Main stem: color (20)		
yellow	Fibror 79 (type A)	1 []
medium green	Bedrobinol (type C), Felina 32 (type A), Theresa (type C)	2 []
dark green	Aida (type C), Dioica 88 (type A)	3 []
purple	EVLS 113 (type C), Fibranova (type A)	4 []
5.11 Only varieties of types A, B and E: Inflorescence: THC content (26)		
absent or very low	Santhica 27 (type A)	1 []
very low to low	Fedora 17 (type A)	2 []
low	Futura 75 (type A)	3 []

Characteristics	Example Varieties	Note
5.12 Only varieties of types C and D: Inflorescence: THC content (27)		
absent or very low	Aida (type C)	1 []
very low to low	Bedrolite (type C), Sara (type C)	2 []
low		3 []
low to medium	Beatriz (type C)	4 []
medium	Bediol (type C)	5 []
medium to high	Bedrobinol (type C), Raquel (type C)	6 []
high	Bedrocan (type C), GRX53 (type D)	7 []
high to very high	Original Blitz (type C)	8 []
very high		9 []
5.13 Inflorescence: CBD content (28)		
absent or very low	Bedrobinol (type C), Bedrocan (type C), Enectacalm (type D), Raquel (type C), Santhica 27 (type A)	1 []
very low to low	Fedora 17 (type A)	2 []
low	Aida (type C), Futura 75 (type A), Theresa (type C)	3 []
low to medium	Chuy (type C)	4 []
medium	Bediol (type C), Divina (type C)	5 []
medium to high	Sibari (type C)	6 []
high	Goya (type C)	7 []
high to very high	A1 Philadelphia (type C), Enectonica (type D)	8 []
very high		9 []

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
-------------------------	-----------------	-------------------

6. Similar varieties and differences from these varieties

Please use the following table and box for comments to provide information on how your candidate variety differs from the variety (or varieties) which, to the best of your knowledge, is (or are) most similar. This information may help the examination authority to conduct its examination of distinctness in a more efficient way.

Denomination(s) of variety(ies) similar to your candidate variety	Characteristic(s) in which your candidate variety differs from the similar variety(ies)	Describe the expression of the characteristic(s) for the similar variety(ies)	Describe the expression of the characteristic(s) for your candidate variety
<i>Example</i>	<i>Plant: natural height</i>	<i>short</i>	<i>long</i>
Comments:			

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
-------------------------	-----------------	-------------------

#7. Additional information which may help in the examination of the variety

7.1 In addition to the information provided in sections 5 and 6, are there any additional characteristics which may help to distinguish the variety?

Yes No

(If yes, please provide details)

7.2 Are there any special conditions for growing the variety or conducting the examination?

Yes No

(If yes, please provide details)

7.3 Other information

Main use

(a) bast fibre and woody core

(b) (oil-) seed

(c) pharmaceuticals

(d) ornamental

(e) other

(please provide details)

Authorities may allow certain of this information to be provided in a confidential section of the Technical Questionnaire.

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
-------------------------	-----------------	-------------------

8. Authorization for release

(a) Does the variety require prior authorization for release under legislation concerning the protection of the environment, human and animal health?

Yes [] No []

(b) Has such authorization been obtained?

Yes [] No []

If the answer to (b) is yes, please attach a copy of the authorization.

9. Information on plant material to be examined or submitted for examination

9.1 The expression of a characteristic or several characteristics of a variety may be affected by factors, such as pests and disease, chemical treatment (e.g. growth retardants or pesticides), effects of tissue culture, different rootstocks, scions taken from different growth phases of a tree, etc.

9.2 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If the plant material has undergone such treatment, full details of the treatment must be given. In this respect, please indicate below, to the best of your knowledge, if the plant material to be examined has been subjected to:

(a) Microorganisms (e.g. virus, bacteria, phytoplasma)	Yes []	No []
(b) Chemical treatment (e.g. growth retardant, pesticide)	Yes []	No []
(c) Tissue culture	Yes []	No []
(d) Other factors	Yes []	No []

Please provide details for where you have indicated "yes".

.....

10. I hereby declare that, to the best of my knowledge, the information provided in this form is correct:

Applicant's name

Signature Date

[End of document]