

**UPOV**

**TG/FESTL(proj.3)**

**ORIGINAL:** English

**DATE:** 2007-05-03

**INTERNATIONAL UNION FOR THE PROTECTION OF NEW VARIETIES OF PLANTS**

GENEVA

**DRAFT**

**FESTULOLIUM**

UPOV CODE: FESTL

(×*Festulolium* Aschers. et Graebn.)

**GUIDELINES**

**FOR THE CONDUCT OF TESTS**

**FOR DISTINCTNESS, UNIFORMITY AND STABILITY**

*prepared by an expert from the United Kingdom*

*to be considered by the*

*Technical Working Party for Agricultural Crops*

*at its thirty-sixth session, to be held in Budapest, Hungary, from May 28 to June 1, 2007*

Alternative Names:\*

<i>Latin</i>	<i>English</i>	<i>French</i>	<i>German</i>	<i>Spanish</i>
× <i>Festulolium</i> Aschers. et Graebn.	Festulolium	Festulolium	Festulolium, Schwingel	Festulolium, Festuca, Canuëla

The purpose of these guidelines (“Test Guidelines”) is to elaborate the principles contained in the General Introduction (document TG/1/3), and its associated TGP documents, into detailed practical guidance for the harmonized examination of distinctness, uniformity and stability (DUS) and, in particular, to identify appropriate characteristics for the examination of DUS and production of harmonized variety descriptions.

**ASSOCIATED DOCUMENTS**

These Test Guidelines should be read in conjunction with the General Introduction and its associated TGP documents.

\* These names were correct at the time of the introduction of these Test Guidelines but may be revised or updated. [Readers are advised to consult the UPOV Code, which can be found on the UPOV Website ([www.upov.int](http://www.upov.int)), for the latest information.]

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## 1. Subject of these Test Guidelines

These Test Guidelines apply to all varieties of hybrids resulting from the crossing of a species of the genus *Festuca* with species of the genus *Lolium* (*x Festulolium*) Aschers. et Graebn.

## 2. Material Required

2.1 The competent authorities decide on the quantity and quality of the plant material required for testing the variety and when and where it is to be delivered. Applicants submitting material from a State other than that in which the testing takes place must ensure that all customs formalities and phytosanitary requirements are complied with.

2.2 The material is to be supplied in the form of seed.

2.3 The minimum quantity of plant material, to be supplied by the applicant, should be:

1.5 kg

The seed should meet the minimum requirements for germination, species and analytical purity, health and moisture content, specified by the competent authority. In cases where the seed is to be stored, the germination capacity should be as high as possible and should be stated by the applicant.

2.4 The plant material supplied should be visibly healthy, not lacking in vigor, nor affected by any important pest or disease.

2.5 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If it has been treated, full details of the treatment must be given.

## 3. Method of Examination

### 3.1 *Number of Growing Cycles*

The minimum duration of tests should normally be two independent growing cycles.

### 3.2 *Testing Place*

Tests are normally conducted at one place. In the case of tests conducted at more than one place, guidance is provided in TGP/9 "Examining Distinctness".

### 3.3 *Conditions for Conducting the Examination*

3.3.1 The tests should be carried out under conditions ensuring satisfactory growth for the expression of the relevant characteristics of the variety and for the conduct of the examination.

3.3.2 The optimum stage of development for the assessment of each characteristic is indicated by a number in the second column of the Table of Characteristics. The stages of development denoted by each number are described at the end of Chapter 8.

3.3.3 The recommended method of observing the characteristic is indicated by the following key in the second column of the Table of Characteristics:

- MG: single measurement of a group of plants or parts of plants
- MS: measurement of a number of individual plants or parts of plants
- VG: visual assessment by a single observation of a group of plants or parts of plants
- VS: visual assessment by observation of individual plants or parts of plants

3.3.4 The recommended type of plot in which to observe the characteristic is indicated by the following key in the second column of the Table of Characteristics:

- A: spaced plants
- B: row plot
- C: special test

#### 3.4 *Test Design*

3.4.1 Each test should be designed to result in a total of at least 60 spaced plants which should be divided between at least 2 replicates. In addition, the test may include 8 meters of row plot which should be divided between at least 2 replicates. The density of the seed should be such that around 200 plants/meter can be expected.

3.4.2 The design of the tests should be such that plants or parts of plants may be removed for measurement or counting without prejudice to the observations which must be made up to the end of the growing cycle.

#### 3.5 *Number of Plants / Parts of Plants to be Examined*

3.5.1 Unless otherwise stated, all observations on single plants should be made on 60 plants or parts taken from each of 60 plants and any other observations made on all plants in the test. In the case of observations of parts taken from single plants, the number of parts to be taken from each of the plants should be 1.

#### 3.6 *Additional Tests*

Additional tests, for examining relevant characteristics, may be established.

### 4. Assessment of Distinctness, Uniformity and Stability

#### 4.1 *Distinctness*

##### 4.1.1 General Recommendations

It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding distinctness. However, the following points are provided for elaboration or emphasis in these Test Guidelines.

#### 4.1.2 Consistent Differences

The differences observed between varieties may be so clear that more than one growing cycle is not necessary. In addition, in some circumstances, the influence of the environment is not such that more than a single growing cycle is required to provide assurance that the differences observed between varieties are sufficiently consistent. One means of ensuring that a difference in a characteristic, observed in a growing trial, is sufficiently consistent is to examine the characteristic in at least two independent growing cycles.

#### 4.1.3 Clear Differences

Determining whether a difference between two varieties is clear depends on many factors, and should consider, in particular, the type of expression of the characteristic being examined, i.e. whether it is expressed in a qualitative, quantitative, or pseudo-qualitative manner. Therefore, it is important that users of these Test Guidelines are familiar with the recommendations contained in the General Introduction prior to making decisions regarding distinctness.

### 4.2 *Uniformity*

4.2.1 It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding uniformity. However, the following points are provided for elaboration or emphasis in these Test Guidelines:

4.2.2 The assessment of uniformity should be according to the recommendations for cross-pollinated varieties in the General Introduction.

### 4.3 *Stability*

4.3.1 In practice, it is not usual to perform tests of stability that produce results as certain as those of the testing of distinctness and uniformity. However, experience has demonstrated that, for many types of variety, when a variety has been shown to be uniform, it can also be considered to be stable.

4.3.2 Where appropriate, or in cases of doubt, stability may be tested, either by growing a further generation, or by testing a new seed stock to ensure that it exhibits the same characteristics as those shown by the previous material supplied.

## 5. Grouping of Varieties and Organization of the Growing Trial

5.1 The selection of varieties of common knowledge to be grown in the trial with the candidate varieties and the way in which these varieties are divided into groups to facilitate the assessment of distinctness are aided by the use of grouping characteristics.

5.2 Grouping characteristics are those in which the documented states of expression, even where produced at different locations, can be used, either individually or in combination with other such characteristics: (a) to select varieties of common knowledge that can be excluded from the growing trial used for examination of distinctness; and (b) to organize the growing trial so that similar varieties are grouped together.

5.3 The following have been agreed as useful grouping characteristics:

- (a) Plant: ploidy (characteristic 1)
- (b) Plant: time of inflorescence emergence after vernalization (characteristic 10)
- (c) Plant: length of longest stem, inflorescence included (when fully expanded) (characteristic 16)

5.4 Guidance for the use of grouping characteristics, in the process of examining distinctness, is provided through the General Introduction.

## 6. Introduction to the Table of Characteristics

### 6.1 *Categories of Characteristics*

#### 6.1.1 Standard Test Guidelines Characteristics

Standard Test Guidelines characteristics are those which are approved by UPOV for examination of DUS and from which members of the Union can select those suitable for their particular circumstances.

#### 6.1.2 Asterisked Characteristics

Asterisked characteristics (denoted by \*) are those included in the Test Guidelines which are important for the international harmonization of variety descriptions and should always be examined for DUS and included in the variety description by all members of the Union, except when the state of expression of a preceding characteristic or regional environmental conditions render this inappropriate.

### 6.2 *States of Expression and Corresponding Notes*

States of expression are given for each characteristic to define the characteristic and to harmonize descriptions. Each state of expression is allocated a corresponding numerical note for ease of recording of data and for the production and exchange of the description.

### 6.3 *Types of Expression*

An explanation of the types of expression of characteristics (qualitative, quantitative and pseudo-qualitative) is provided in the General Introduction.

### 6.4 *Example Varieties*

Where appropriate, example varieties are provided to clarify the states of expression of each characteristic.

## 6.5 *Legend*

(\*) Asterisked characteristic – see Chapter 6.1.2

QL: Qualitative characteristic – see Chapter 6.3

QN: Quantitative characteristic – see Chapter 6.3

PQ: Pseudo-qualitative characteristic – see Chapter 6.3

MG, MS, VG, VS: See Chapter 3.3.3

A, B, C: See Chapter 3.3.4

(a)-(f) See Explanations on the Table of Characteristics in Chapter 8.1

(+) See Explanations on the Table of Characteristics in Chapter 8.2

(10)-(68) See Explanations on the Table of Characteristics in Chapter 8.3.

7. Table of Characteristics/Tableau des caractères/Merkmalstabelle/Tabla de caracteres

Char No.	Method of Examination	English	français	deutsch	español	Example Varieties/ Exemples/ Beispielssorten/ Variedades ejemplo	Note/ Nota	
<b>1.</b> <b>(*)</b> <b>(+)</b>	<b>C</b>	<b>Plant: ploidy</b>						
<b>QL</b>		diploid				<b>Matrix</b>	2	
		tetraploid				Perun, Paulita, Prior	4	
		<b>hexaploid</b>	<b>■</b>	<b>■</b>	<b>■</b>	<b>Felina, Hykor</b>	<b>6</b>	
<b>2</b>	<b>20-29</b>	<b>Plant: vegetative growth habit</b>						
		<b>VS A (without VG B vernalization)</b>						
<b>QN</b>	<b>(a)</b>	semi-erect		<b>■</b>		<b>■</b>	3	
		medium				Lofa, Paulita	5	
		semi-prostrate				Sulino	7	
<b>3.</b>	<b>20-29</b>	<b>Leaf : length (at vegetative stage)</b>						
		<b>VG B</b>						
		<b>[NZ - to provide example varieties]</b>						
<b>QN</b>		short				<b>Lesana</b>	3	
		medium				<b>Prior</b>	5	
		broad				<b>Felopa</b>	7	
<b>4</b>	<b>20-29</b>	<b>Leaf : width (at vegetative stage)</b>						
		<b>VG B</b>						
		<b>[NZ - to provide example varieties]</b>						
<b>QN</b>		narrow				<b>Lesana</b>	3	
		medium				<b>Prior</b>	5	
		broad				<b>Felopa</b>	7	



Char No.	Method of Examination	English	français	deutsch	español	Example Varieties/ Exemples/ Beispielssorten/ Variedades ejemplo	Note/ Nota
<b>5.</b>	<b>30</b>	<b>Plant: width (after vernalization)</b>		<b>[ ESA – to check stability of the characteristic]</b>			
(+)	MS A VS A						
QN		narrow				Prior	3
		medium				Sulino	5
		wide				Perun	7
<b>6.</b>	<b>30-39</b>	<b>Plant: vegetative growth habit (after vernalization)</b>					
	VS A VG B						
QN	(a)	semi-erect				Paulita	3
		medium				Lofa	5
		semi-prostrate				Prior	7
<b>7.</b>	<b>30-39</b>	<b>Plant : height (after vernalization)</b>					
	VG B						
QN		short				Prior	3
		medium				Perun	5
		tall					7
<b>8.</b>	<b>50</b>	<b>Plant: time of inflorescence emergence (after vernalization)</b>					
(*) (+)	MSA MGB						
QN		early				Sulino	3
		medium				Prior	5
		late					7

Char No.	Method of Examination	English	français	deutsch	español	Example Varieties/ Exemples/ Beispielssorten/ Variedades ejemplo	Note/ Nota	
<b>9.</b>	<b>50</b>	<b>Plant: natural height at inflorescence emergence</b>						
<b>(+)</b>	<b>MS A</b>							
<b>QN</b>	<b>(b)</b>	short				Prior	3	
		medium				Perun	5	
		tall					7	
<b>10.</b>	<b>50</b>	<b>Flag leaf: length</b>						
<b>(*)</b>								
<b>(+)</b>	<b>MS A</b>							
<b>QN</b>	<b>(b)</b>	short				Prior	3	
		medium				Sulino	5	
		long				Perun	7	
<b>11.</b>	<b>50</b>	<b>Flag leaf: width</b>						
<b>(*)</b>								
<b>(+)</b>	<b>MS A</b>							
<b>QN</b>	<b>(b)</b>	narrow				Prior	3	
		medium				Lofa	5	
		broad					7	
<b>12.</b>	<b>60-68</b>	<b>Plant: length of longest stem, inflorescence included (when fully expanded)</b>						
<b>(*)</b>								
<b>(+)</b>	<b>MS A</b>							
<b>QN</b>	<b>(c)</b>	short				Prior	3	
		medium				Sulino	5	
		long					7	
<b>13.</b>	<b>60-68</b>	<b>Plant: length of upper internode</b>						
<b>(*)</b>								
<b>(+)</b>	<b>MSA</b>							
<b>QN</b>	<b>(c)</b>	short					3	
		medium					5	
		long					7	

[NZ - to provide example varieties]

Char No.	Method of Examination	English	français	deutsch	español	Example Varieties/ Exemples/ Beispielssorten/ Variedades ejemplo	Note/ Nota
14.	60-68 MS A	<b>Inflorescence: degree of branching</b>		[TWA 2006 – suggested as a new characteristic]			
QN	(c)	few				Perun	3
		medium					5
		many				Felina	7
15	60-68 VS A	<b>Inflorescence: length</b>					
QN	(c)	long					3
		medium				Prior	5
		short				Perun	7
16	60-68 MS A	<b>Inflorescence: number of spikelets</b>		[UK – this characteristic only works for varieties with unbranched inflorescences – delete?]			
QN	(c)	few				Sulino	3
		medium				Prior	5
		many				Lofa	7
17.	60-68 (+) MS A	<b>Inflorescence: density</b>		[UK – this characteristic only works for varieties with unbranched inflorescences – delete?]			
QN	(c)	lax				Sulino	3
		medium				Perun	5
		dense					7

Char No.	Method of Examination	English	français	deutsch	español	Example Varieties/ Exemples/ Beispielssorten/ Variedades ejemplo	Note/ Nota
<b>18.</b>	<b>60-68</b>	<b>Inflorescence: length of outer MS A glume at basal spikelet</b>		<b>[TWA 2006 – very labour intensive – NZ to consider if useful]</b>			
<b>QN</b>	<b>(c)</b>	short				Sulino	3
		medium				Prior	5
		long				Lofa	7
<b>19.</b>	<b>60-68</b>	<b>Inflorescence: length of basal MS A spikelet excluding awn</b>		<b>[TWA 2006 – very labour intensive – NZ to consider if useful]</b>			
<b>QN</b>	<b>(c)</b>	short				Lofa	3
		medium				Perun	5
		long				Sulino	7

8. Explanations on the Table of Characteristics

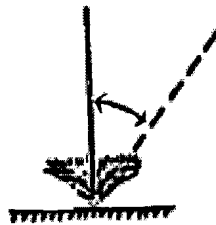
8.1 *Explanations covering several characteristics*

Characteristics containing the following key in the second column of the Table of Characteristics should be examined as indicated below:

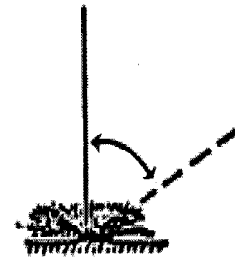
- (a) For the estimation of growth habit, characteristics 2 and 6, the observations should be made visually from the attitude of the leaves of the plant as a whole. The angle formed by the imaginary line through the region of greatest leaf density and the vertical should be used. Characteristic 2 may be recorded during the growing season in which the trials are planted



3  
semi-erect



5  
medium



7  
semi-prostrate

- (b) To be recorded on each individual plant at the time of inflorescence emergence (Growth Stage DC 54), that is at the same time as Characteristic 8
- (c) Measurements for characteristics 12 to 19 should be made on the longest stem.

## 8.2 *Explanations for individual characteristics*

### Ad. 1: Plant: ploidy

The ploidy of the plant can be determined either by standard cytological methods.

### Ad. 5: Plant: width

To allow for irregular plant shapes (for example due to wind shaping effects) the plant width is determined by taking two measurements (MS A) or by making two visual observations (VS A) of the diameter across the plant at right angles to each other and then using the average of these two figures as the plant width.

### Ad. 8: Plant: Time of inflorescence emergence

Timing of observations will depend upon time of planting.

Spaced plants or row plots should be observed at least twice per week.

#### Plots with spaced plants

The date of inflorescence emergence of each single plant should be observed. A single plant is considered to have headed when the tip of three inflorescences can be seen protruding from the flag leaf sheath (Growth Stage DC 50). From the single plant data, a mean date per plot and a mean date per variety is obtained.

#### Row plots

The time of inflorescence emergence is the date at which the average plot stage DC 54 has been reached. This date should – if necessary – be obtained by interpolation. At each observation date, the average plot stage should be expressed in one of the following growth stages:

- |     |       |   |
|-----|-------|---|
| (1) | DC 50 | First spikelet of inflorescence just visible        |
| (2) | DC 52 | 25% of the inflorescence emerged (across all stems) |
| (3) | DC 54 | 50% of the inflorescence emerged (across all stems) |
| (4) | DC 56 | 75% of the inflorescence emerged (across all stems) |

### Ad. 9: Plant: natural height at inflorescence emergence

To be recorded by measuring the average height of the foliage in the centre of the plant.

### Ad. 10: Flag leaf: length

To be measured from the ligule to the tip of the flag leaf on the longest stem.

Ad. 11: Flag leaf: width

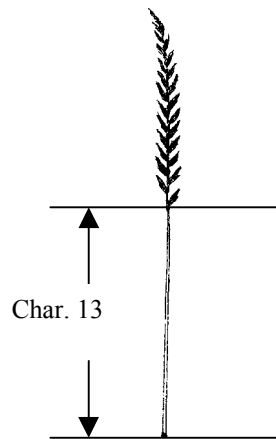
To be measured one third of the way from the base to the tip of the flag leaf on the longest stem

Ad. 12: Plant: length of longest stem, inflorescence included (when fully expanded)

To be recorded in the field from ground level, when the inflorescence is fully expanded.

Ad. 13: Plant: length of upper internode

To be measured from the top node to the base of the inflorescence.



Ad. 17: Inflorescence: density

This characteristic is calculated by dividing Characteristic 15 (Inflorescence: length) by Characteristic 16 (Inflorescence: number of spikelets)

8.3 *Growth stages of grasses derived from the decimal code for the growth stages of cereals (Zadoks, et al., 1974).*

This decimal code is in close conformity with the BBCH-code (Meier, 1997)

*Seedling growth (seedling: one shoot)*

DC 10	First leaf through coleoptile
DC 15	Five leaves unfolded
DC 19	Nine or more leaves unfolded

*Tillering*

DC 20	Main shoot only (beginning of tillering)
DC 23	Main shoot and 3 tillers
DC 25	Main shoot and 5 tillers
DC 29	Main shoot and 9 more tillers

*Stem elongation*

DC 30	Pseudo-stem erection (formed by sheaths of leaves)
DC 31	First node detectable (early stem extension across all stems)
DC 35	Fifth node detectable (50% extension across all stems)
DC 39	Flag leaf ligula/collar just visible (pre-boot stage)

*Booting*

DC 41	Flag leaf sheath extending (little enlargement of the inflorescence, early boot-stage)
DC 45	Boots swollen (late-boot stage)
DC 47	First leaf sheath opening
DC 49	First awns visible (in awned forms only)

*Inflorescence emergence (mostly non-synchronous)*

DC 50	First spikelet of inflorescence just visible
DC 52	25% of the inflorescence emerged (across all stems)
DC 54	50% of the inflorescence emerged (across all stems)
DC 56	75% of the inflorescence emerged (across all stems)
DC 58	Emergence of inflorescence completed

*Anthesis (mostly non-synchronous)*

DC 60	Beginning of anthesis
DC 64	Anthesis half-way
DC 68	Anthesis complete



## 9. Literature

Baltjes, H.J., Klein Geltink, D.J.A., Nienhuis, K.H. and Luesink, B., 1985: "Linking Distinctiveness and Description of Varieties," *Journal of the National Institute of Agricultural Botany*, 17, pages 9-19

Barker, R.E., Kilgore, J.A., Cook, R.L., Garay, A.E. and Warnke, S. E., 2001: Use of flow cytometry to determine ploidy level of ryegrass. *Journal of Seed Science and Technology*, 29, 493-502.

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Hawkins, R.P., 1958, "The Classification of the Strains (Varieties) of Herbage Plants," *Journal of the National Institute of Agricultural Botany*, 9, pages 434-449

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Patterson, H.D. and Weatherup S.T.C., (1984): "Statistical Criteria for Distinctness between Varieties of Herbage Crops," *Journal of Agricultural Science, Cambridge*, 102, pages 59-68

Squire A.M., 1962: A rapid technique for counting chromosomes in grass breeding studies. *Journal of the British Grassland Society*, 21(4), 305-306.

Tyler, B.F., Hayes, J.D. and Ellis Davies, W., 1985: "IBPGR/CEC Descriptive List for Forage Grasses," *International Board for Plant Genetic Resources (IBPGR)*, 83/90

Weatherup, S.T.C., 1980: "Statistical Procedures for Distinctness, Uniformity and Stability Trials," *Journal of Agricultural Science, Cambridge*, 94, page 31-46

Zadoks, J.C., Chang T.T., and C.F. Konzak, 1974: A decimal code for the growth stages of cereals. *Weed Research* 14: 415 – 421.

10. Technical Questionnaire

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
		Application date: (not to be filled in by the applicant)
<b>TECHNICAL QUESTIONNAIRE</b> to be completed in connection with an application for plant breeders' rights		
1. Subject of the Technical Questionnaire:		
<i>Botanical Name</i>	<input type="text" value="×Festulolium Aschers. et Graebn."/>	
Common Name	<input type="text" value="Festulolium"/>	
2. Applicant		
Name	<input type="text"/>	
Address	<input type="text"/>	
Telephone No.	<input type="text"/>	
Fax No.	<input type="text"/>	
E-mail address	<input type="text"/>	
Breeder (if different from applicant)	<input type="text"/>	
3. Proposed denomination and breeder's reference		
Proposed denomination (if available)	<input type="text"/>	
Breeder's reference	<input type="text"/>	

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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#4. Information on the breeding scheme and propagation of the variety

4.1 Breeding scheme

Variety resulting from:

4.1.1 Crossing

(a) controlled cross  [ ]  
(please state parent varieties)

(b) partially known cross  [ ]  
(please state known parent variety(ies))

(c) unknown cross  [ ]

4.1.2 Mutation  [ ]  
(please state parent variety)

4.1.3 Discovery and development  [ ]  
(please state where and when discovered  
and how developed)

4.1.4 Other  [ ]  
(please provide details)

4.2 Method of propagating the variety

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# Authorities may allow certain parts of this information to be given in a confidential section of the Technical Questionnaire.

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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5. Characteristics of the variety to be indicated (the number in brackets refers to the corresponding characteristic in Test Guidelines; please mark the note which best corresponds).

	Characteristics	Example Varieties	Note
<b>5.1</b>	<b>Plant: ploidy</b>		
<b>(1)</b>			
	diploid	Matrix	2[ ]
	tetraploid	Perun, Paulita, Prior	4[ ]
	hexaploid	Felina, Hykor	6[ ]
<b>5.3</b>	<b>Plant: time of inflorescence emergence (after vernalization)</b>		
<b>(8)</b>			
	early	Sulino	3[ ]
	medium	Prior	5[ ]
	late		7[ ]
<b>5.4</b>	<b>Plant: length of longest stem, inflorescence included (when fully expanded)</b>		
<b>(12)</b>			
	short	Prior	3[ ]
	medium	Sulino	5[ ]
	long		7[ ]

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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6. Similar varieties and differences from these varieties

*Please use the following table and box for comments to provide information on how your candidate variety differs from the variety (or varieties) which, to the best of your knowledge, is (or are) most similar. This information may help the examination authority to conduct its examination of distinctness in a more efficient way.*

Denomination(s) of variety(ies) similar to your candidate variety	Characteristic(s) in which your candidate variety differs from the similar variety(ies)	Describe the expression of the characteristic(s) for the <b>similar</b> variety(ies)	Describe the expression of the characteristic(s) for <b>your</b> candidate variety
<i>Example</i>	<i>Plant: length of longest stem, inflorescence included (when fully expanded)</i>	<i>short</i>	<i>medium</i>

Comments:

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#7. Additional information which may help in the examination of the variety

7.1 In addition to the information provided in sections 5 and 6, are there any additional characteristics which may help to distinguish the variety?

Yes [ ] No [ ]

(If yes, please provide details)

7.2 Are there any special conditions for growing the variety or conducting the examination?

Yes [ ] No [ ]

(If yes, please provide details)

7.3 Main use

- (a) forage [ ]  
(b) amenity [ ]  
(c) other [ ]  
(please provide details)

7.4 Type

Festuca- type [ ] Lolium - type [ ]

(Please describe inflorescence type and/or indicate other identifying characteristic/s) to

7.5 Other information

8. Authorization for release

(a) Does the variety require prior authorization for release under legislation concerning the protection of the environment, human and animal health?

Yes [ ] No [ ]

(b) Has such authorization been obtained?

Yes [ ] No [ ]

If the answer to (b) is yes, please attach a copy of the authorization.

# Authorities may allow certain of this information to be provided in a confidential section of the Technical Questionnaire.

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9. Information on plant material to be examined or submitted for examination.

9.1 The expression of a characteristic or several characteristics of a variety may be affected by factors, such as pests and disease, chemical treatment (e.g. growth retardants or pesticides), effects of tissue culture, different rootstocks, scions taken from different growth phases of a tree, etc.

9.2 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If the plant material has undergone such treatment, full details of the treatment must be given. In this respect, please indicate below, to the best of your knowledge, if the plant material to be examined has been subjected to:

- |   |         |        |
|---|---------|--------|
| (a) Microorganisms (e.g. virus, bacteria, phytoplasma)    | Yes [ ] | No [ ] |
| (b) Chemical treatment (e.g. growth retardant, pesticide) | Yes [ ] | No [ ] |
| (c) Tissue culture  | Yes [ ] | No [ ] |
| (d) Other factors   | Yes [ ] | No [ ] |

Please provide details for where you have indicated "yes".

.....

10. I hereby declare that, to the best of my knowledge, the information provided in this form is correct:

Applicant's name	<input type="text"/>		
Signature	<input type="text"/>	Date	<input type="text"/>

[End of document]