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**INTERNATIONAL UNION
FOR THE PROTECTION
OF NEW VARIETIES OF
PLANTS**

**UNION INTERNATIONALE
POUR LA PROTECTION
DES OBTENTIONS
VÉGÉTALES**

**INTERNATIONALER
VERBAND ZUM SCHUTZ
VON PFLANZEN-
ZÜCHTUNGEN**

**UNIÓN INTERNACIONAL
PARA LA PROTECCIÓN
DE LAS OBTENCIONES
VEGETALES**

DRAFT

GUIDELINES
FOR THE CONDUCT OF TESTS
FOR DISTINCTNESS, UNIFORMITY AND STABILITY

WHITE MUSTARD

(Sinapis alba L.)

These Guidelines should be read in conjunction with document TG/1/2, which contains explanatory notes on the general principles on which the Guidelines have been established.

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I. Subject of these Guidelines

These Test Guidelines apply to all varieties of *Sinapis alba* L.

II. Material Required

1. The competent authorities decide when, where and in what quantity and quality the plant material required for testing the variety is to be delivered. Applicants submitting material from a State other than that in which the testing takes place must make sure that all customs formalities are complied with. The minimum quantity of seed to be supplied by the applicant in one or several samples should be:

500 g

The seed should at least meet the minimum requirements for germination capacity, moisture content and purity for marketing certified seed in the country in which the application is made. The germination capacity should be as high as possible.

2. The plant material must not have undergone any treatment unless the competent authorities allow or request such treatment. If it has been treated, full details of the treatment must be given.

III. Conduct of Tests

1. The minimum duration of tests should normally be two independent growing cycles.
2. The tests should normally be conducted at one place. If any important characteristics of the variety cannot be seen at that place, the variety may be tested at an additional place.
3. The field tests should be carried out under conditions ensuring normal growth. The distance between rows and between plants within the rows should be adjusted to enable observations on individual plants. The size of the plots should be such that plants or parts of plants may be removed for measurement and counting without prejudice to the observations which must be made up to the end of the growing period. Each test should include as a minimum 300 plants which should be divided between three or more replicates. In addition each test should include a replicate of minimum 300 plants for the characteristics assessed by observation of a group of plants.

4. Additional tests for special purposes may be established.

IV. Methods and Observations

1. Unless otherwise stated, all observations for assessment of distinctness and stability should be made on 60 plants or parts taken from each of 60 plants.

2. For the assessment of uniformity

- unless otherwise stated, all observations determined by measurements should be made on 60 plants or parts taken from each of 60 plants (MS)
- all single observations of a group of plants or parts of plants should be made on the total plot of minimum 300 plants.

The variability within the variety should not exceed the variability of comparable varieties already known.

V. Grouping of Varieties

1. The collection of varieties to be grown should be divided into groups to facilitate the assessment of distinctness. Characteristics which are suitable for grouping purposes are those which are known from experience not to vary, or to vary only slightly, within a variety. Their various states of expression should be fairly evenly distributed throughout the collection.

2. It is recommended that the competent authorities use the following characteristics for grouping varieties:

- (a) Seed: erucic acid (characteristic 1)
- (b) Ploidy (characteristic 2)
- (c) Flower: yellow color of petals (characteristic 13)

VI. Characteristics and Symbols

1. To assess distinctness, uniformity and stability, the characteristics and their states as given in the Table of Characteristics should be used.

2. Notes (numbers), for the purposes of electronic data processing, are given opposite the states of expression for each characteristic. For each characteristic it is indicated whether measurements of a number of single plants or parts of plants (MS), measurement of a group of plants or parts of plants (MG) or visual assessments by a single observation of a group of plants or parts of plants (VG) should be used.

3. Legend:

- (*) Characteristics that should be used on all varieties in every growing period over which examinations are made and always be included in the variety descriptions, except when the state of expression of a preceding characteristic or regional environmental conditions render this impossible.
- (+) See Explanations on the Table of Characteristics in Chapter VIII.
- 1) The optimum stage of development for the assessment of each characteristic is indicated by a number in the second column. The stages of development denoted by each number are described at the end of chapter VIII.

MG: measurement of a group of plants or parts of plants

MS: measurement of a number of single plants or parts of plants

VG: visual assessment by a single observation of a group of plants or parts of plants

VIII. Explanations on the Table of Characteristics

Ad. 1: Seed: erucic acid

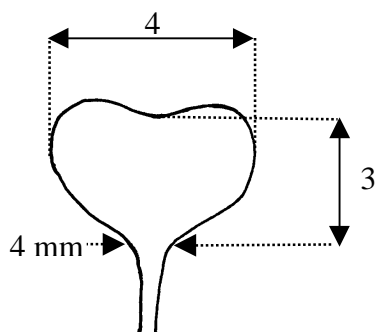
The erucic acid content should be observed on seed sent in by the applicant. It should be expressed as a percentage by mass of methyl esters in accordance with the ISO standard in document 5508, paragraph 6.2.2.1. Seed containing 2% or less of erucic acid will be classified as “absent.”

Ad. 2: Ploidy

Ploidy should be assessed on at least 100 seedlings.

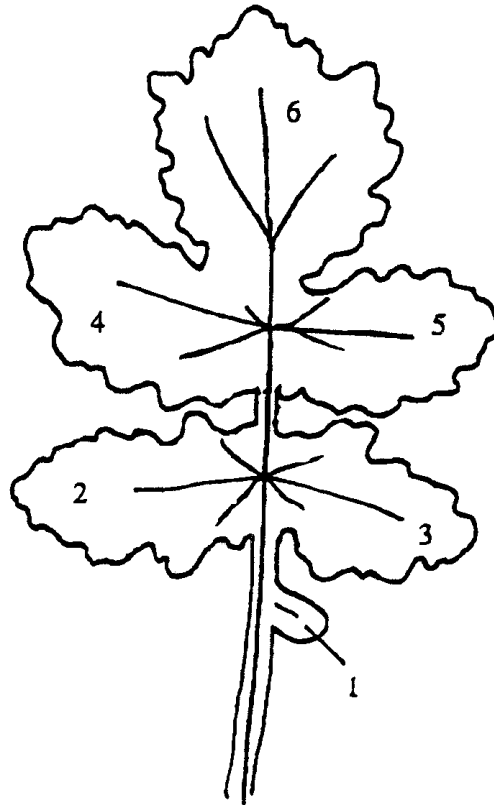
Ad. 3 + 4: Cotyledon: length (3) and width (4)

The measurement should be taken in the glasshouse. If the two cotyledons differ in size, the bigger one should be measured. The length is defined as distance between the inclination at top of the cotyledon and the point where the width of the petiole is about 4 mm. The width of the cotyledon should be measured at the widest point of the cotyledons.



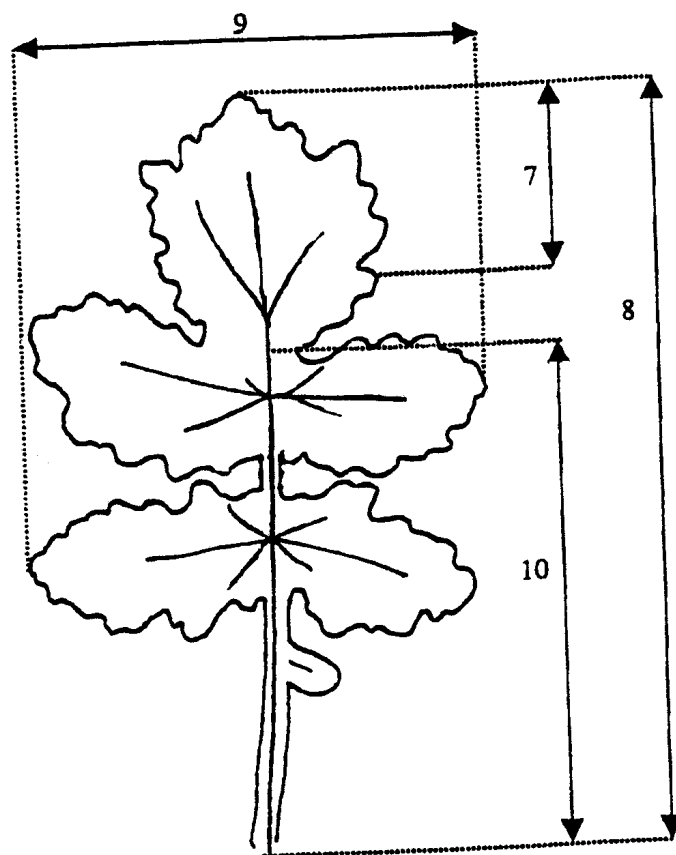
Ad. 6: Leaf: number of lobes

Parts of the leaf blade are considered as lobes if their length is at least equivalent to the width of the leaf petiole at their point of attachment and if the upper notch of the blade has at least half the length of the lobe itself.



Ad. 7-10: Leaf: dentation (7), length (8), width (9), length of petiole (10)

7 = part on which the dentation should be recorded (characteristic 7)



Ad. 11: Time of flowering

The observation should be done at least three times per week and more frequently if there is any need to do so. The date should be calculated - if necessary by interpolation - at which 50% of plants show at least one open flower.

When assessed on the plot as a whole, the recommended percentage is 10%. This characteristic may be useful for arranging the varieties in the collection.

Ad. 12 : Plant: height at flowering

The height of the plants should be assessed when all normally developed plants have opened at least one flower.

Ad. 17-21: Siliqua

All observations on the siliqua should be recorded in the midpart of the inflorescence of the main stem.

Ad. 22: Seed: thousand seed weight

Per replicate a bulk sample of 20 siliques should be taken.

Ad. 23: Generative development in year of sowing for late summer sown trials

The observation of the growth stages reached (proportion of plants below bud stage, in bud stage, in flowering stage, in stage of siliqua formation) should be made in autumn, when the development stagnates.

Alternatively the beginning of flowering may be observed in this trial; early flowering would mean strong generative development, late flowering would mean weak development.

Phenological growth stages according to the BBCH-identification keys of oilseed rape (Meier, 1997)

Code	Description
Principal growth stage 0: Germination	
00	Dry seed
01	Beginning of seed imbibition
03	Seed imbibition complete
05	Radicle emerged from seed
07	Hypocotyl with cotyledons emerged from seed
08	Hypocotyl with cotyledons growing towards soil surface
09	Emergence: cotyledons emergence through soil surface
Principal growth stage 1: Leaf development	
10	Cotyledons completely unfolded
11	First leaf unfolded
12	2 leaves unfolded
13	3 leaves unfolded
1.	Stages continuous till...
19	9 or more leaves unfolded
Principal growth stage 2: --	
Principal growth stage 3: Stem elongation	
30	Beginning of stem elongation: no internodes ('rosette')
31	1 visibly extended internodes
32	2 visibly extended internodes
33	3 visibly extended internodes
3.	Stages continuous till...
39	9 or more visibly extended internodes
Principal growth stage 4: --	
Principal growth stage 5: Inflorescence emergence	
50	Flower buds present, still enclosed by leaves
51	Flower buds visible from above ("green bud")
52	Flower buds free, level with the youngest leaves
53	Flower buds raised above the youngest leaves
55	Individual flower buds (main inflorescence) visible but still closed
57	Individual flower buds (secondary inflorescences) visible but still closed
59	First petals visible, flower buds still closed ("yellow bud")

Code	Description
Principal growth stage 6: Flowering	
60	First flowers open
61	10 % of flowers on main raceme open, main raceme elongating
62	20 % of flowers on main raceme open
63	30 % of flowers on main raceme open
64	40 % of flowers on main raceme open
65	Full flowering: 50 % flowers on main raceme open, older petals falling
67	Flowering declining: majority of petals fallen
69	End of flowering
Principal growth stage 7: Development of fruit	
71	10 % of pods have reached final size
72	20 % of pods have reached final size
73	30 % of pods have reached final size
7.	Stages continuous till...
78	80 % of pods have reached final size
79	Nearly all pods have reached final size
Principal growth stage 8: Ripening	
80	Beginning of ripening: seed green, filling pod cavity
81	10 % of pods ripe, seeds dark and hard
82	20 % of pods ripe, seeds dark and hard
83	30 % of pods ripe, seeds dark and hard
8.	Stages continuous till...
88	80 % of pods ripe, seeds dark and hard
89	Fully ripe: nearly all pods ripe, seeds dark and hard

IX. Literature

Growth stages of mono- and dicotyledonous plants: BBCH-Monograph. Federal Biological Research Centre of Agriculture and Forestry (ed.) Ed. by Uwe Meier.-Berlin; Wien [u.a.]: Blackwell Wiss.-Verl., 1997.

X. Technical Questionnaire

	<p>Reference Number (not to be filled in by the applicant)</p>
<p>TECHNICAL QUESTIONNAIRE to be completed in connection with an application for plant breeders' rights</p>	
<p>1. Species <i>Sinapis alba L.</i> WHITE MUSTARD</p>	
<p>2. Applicant (Name and address)</p>	
<p>3. Proposed denomination or breeder's reference</p>	

4. Information on origin, maintenance and reproduction of the variety

4.1 Genetic origin and breeding method

4.2 Other information

5. Characteristics of the variety to be indicated (the number in brackets refers to the corresponding characteristic in Test Guidelines; please mark the state of expression which best corresponds).

Characteristics	Example Varieties	Note
5.1 Seed: erucic acid (1)		
absent	Rizo	1 []
present	Emergo	9 []
5.2 Ploidy (2)		
diploid	Emergo	2 []
tetraploid	Oscar	4 []
5.3 Time of flowering (11)		
very early	Carla	1 []
early	Silenda	3 []
medium	Litember	5 []
late	Sito	7 []
very late		9 []

Characteristics		Example Varieties	Note
5.4	Plant: total length		
(16)			
	short	Silenda	3 []
	medium	Perine	5 []
	long	Litember	7 []
6. Similar varieties and differences from these varieties			
Denomination of similar variety	Characteristic in which the similar variety is different ^{o)}	State of expression of similar variety	State of expression of candidate variety
^{o)} In the case of identical states of expressions of both varieties, please indicate the size of the difference.			

7. Additional information which may help to distinguish the variety

7.1 Resistance to pest and diseases

7.2 Special conditions for the examination of the variety

7.3 Other information

8. Authorization for release

- (a) Does the variety require prior authorization for release under legislation concerning the protection of the environment, human and animal health?

Yes []

No []

- (b) Has such authorization been obtained?

Yes []

No []

If the answer to that question is yes, please attach a copy of such an authorization.

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