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| International Union for the Protection of New Varieties of Plants |  |

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Partial revision of the Test Guidelines for PEA

Document prepared by an expert from the European Union

Disclaimer: this document does not represent UPOV policies or guidance

 The purpose of this document is to present a proposal for a partial revision of the Test Guidelines for Pea (*Pisum sativum* L.) (document TG/7/10 Rev.).

 The Technical Working Party for Vegetables (TWV), at its fifty-first session, held in Roelofarendsveen, Netherlands, from July 3 to 7, 2017, considered a proposal for a partial revision of the Test Guidelines for Pea (*Pisum sativum* L.) on the basis of documents TG/7/10 Rev. and TWV/51/6 “Partial Revision of the Test Guidelines for Pea” and proposed the following revisions to the Test Guidelines for Pea (see document TWV/51/16 “Report”, paragraphs 104 and 105):

1. To add new example varieties for Characteristic 60 “Resistance to *Ascochyta pisi* Race”
2. To change the methodology for Characteristic 60 under Ad. 60

 The proposed changes are presented below in highlight and underline (insertion).

Proposed change to add new example varieties for Characteristic 60 “Resistance to Ascochyta pisi Race”

*Current wording*

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| 60.(+) | VG | Resistance to *Ascochyta pisi*, Race C | Résistance à *Ascochyta pisi*, race C | Resistenz gegen *Ascochyta pisi*, Pathotyp C | Resistencia a *Ascochyta pisi*, Raza C |  |  |
| QL |  | absent | absente | fehlend | ausente | Kelvedon Wonder | 1 |
|  |  | present | présente | vorhanden | presente | Rondo | 9 |

*Proposed new wording*

|  |  |  |  |  |  |  |  |
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| 60.(+) | VG | Resistance to *Ascochyta pisi*, Race C | Résistance à *Ascochyta pisi*, race C | Resistenz gegen *Ascochyta pisi*, Pathotyp C | Resistencia a *Ascochyta pisi*, Raza C |  |  |
| QL |  | absent | absente | fehlend | ausente | Crecerelle, Kelvedon Wonder | 1 |
|  |  | present | présente | vorhanden | presente | Madonna, Nina, Rondo | 9 |

Proposed change to the methodology for Characteristic 60 under Ad. 60

*Current wording*

Ad. 60: Resistance to *Ascochyta pisi*, Race C (Ascochyta Leaf and Pod Spot)

Resistant and Susceptible varieties

Kelvedon Wonder (susceptible = resistance absent (1))

Rondo (resistant = resistance present (9))

Isolates and isolate identity

Isolate used in the test: Tezier Strain

Isolate identity is determined by testing against a host differential set.

Maintenance of isolates

Maintain on Mathur medium at ambient temperature. Isolate identity is determined by testing against a host differential set.

Source for isolates: GEVES SNES

Station Nationale d’Essais de Semences

Rue George Morel, B.P.24

49071 Beaucouzé Cedex France

Preparation of inoculum

Add 0.4% Tween 80 wetting agent to aid dispersal of spores. Remove hyphal fragments by straining solution through muslin. Concentration of 106 spores/ml

Inoculation and assessment of disease

Grow seedlings in glasshouse under natural daylength at 20oC and high humidity. Spray inoculum on young seedlings 10-15 days after emergence; mist spray 2 or 3 times per day for 15 minutes. Alternatively, inoculation can be made at the apex of enclosed leaves. This method does not require conditions of high humidity.

Plants are assessed about 5 days after inoculation. Infection is very clear when present: necrotic lesions are slightly sunken, brown and sharply delineated. Lesions are circular on pods and elongated on stems. Two replicates of 10 plants are grown; a third replicate is grown if any problems arise.

Genetic background

The expression of resistance to Race C (also known as BP2) is controlled by a single dominant gene Rap2. At least five pathotypes and four resistance alleles are known.

*Proposed new wording*

Ad. 60: Resistance to *Ascochyta pisi*, Race C

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| 1. | Pathogen | *Ascochyta pisi* |
| 2. | Quarantine status | no |
| 3. | Host species | Pea – *Pisum sativum* L. |
| 4. | Source of inoculum | GEVES[[1]](#footnote-2) (FR) or SASA[[2]](#footnote-3) (GB) |
| 5. | Isolate | *Ascochyta pisi* race C strain 21A.13. (the test protocol has been validated in a European CPVO co-funded project[[3]](#footnote-4) with this isolate). |
| 6. | Establishment isolate identity | genetically defined pea controls (Physiological races of *A. pisi* and differentials, adapted from Gallais et Bannerot, 1992) |

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| Physiological races (Dr Hubbeling)Strains | DN°1 | \_Several isolates | \_N°4 | \_N°14 | CTézier21A.13 | B\_ | E\_ |
| Gullivert | R | R | R | R | S | R | R |
| Rondo | R | R | S | VLS | R | R | S |
| Finale | R | R | S | LS | R | - | - |
| Kelvedon Wonder | R | S | S | S | S | R | R |
| Dark Skin Perfection | S | S | S | S | S | R | S |
| Arabal, Cobri, Starcovert, Sucovert, Vitalis | S | S | S | S | S | S | S |

R = resistant; S = susceptible, VLS = very lightly susceptible, LS = lightly susceptible

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| 7. | Establishment pathogenicity | test on susceptible plants |
| 8. | Multiplication inoculum |  |
| 8.1 | Multiplication medium | V8 agar or Mathur medium or Potato Dextrose Agar or a synthetic medium. |
| 8.2 | Multiplication variety | - |
| 8.3 | Plant stage at inoculation | - |
| 8.4 | Inoculation medium | water, option: add Tween 80 (wetting agent to aid dispersal of spores, e.g. 0.4%) |
| 8.5 | Inoculation method | - |
| 8.6 | Harvest of inoculum | see 10.1 |
| 8.7 | Check of harvested inoculum | see 10.2 |
| 8.8 | Shelflife/viability inoculum | 4/8h, keep cool to prevent spores’ germination |
| 9. | Format of the test |  |
| 9.1 | Number of plants per genotype | at least 20 plants and 5 non inoculated plants per variety |
| 9.2 | Number of replicates | - |
| 9.3 | Control varieties |  |
|  | Susceptible | Crecerelle, Kelvedon Wonder |
|  | Resistant | Nina and Madonna or Rondo |
| 9.4 | Test design | - |
| 9.5 | Test facility | climate room or greenhouse |
| 9.6 | Temperature | 20°C |
| 9.7 | Light | 12 hours or longer |
| 9.8 | Season | - |
| 9.9 | Special measures | high humidity or watering by spraying 2 or 3 times per day |
| 10. | Inoculation |  |
| 10.1 | Preparation inoculum | remove hyphen fragments by straining solution through muslin |
| 10.2 | Quantification inoculum | 106 spores/mL (to adapt depending conditions of tests) |
| 10.3 | Plant stage at inoculation | 2 weeks old seedlings (i.e. 2-3 node stage) |
| 10.4 | Inoculation method | spraying on green leaves without surface moisture |
| 10.5 | First observation | - |
| 10.6 | Second observation | - |
| 10.7 | Final observations | 10-18 days post-inoculation |
| 11. | Observations |  |
| 11.1 | Method | visual |
| 11.2 | Observation scale | 0: no symptoms1: few small superficial necrosis2: bigger darker and deep necrosis3: necrosis at each level of the plant or serious symptoms surrounding the stemMadonna, Nina and Rondo will be resistant controls; varieties with same level of resistance as Madonna/Rondo and/or Nina will be interpreted as resistant. Crecerelle and Kelvedon Wonder will be susceptible controls, varieties with a lower level of resistance than Nina as well as Madonna/Rondo will be interpreted as susceptible. |



Resistant

Crecerelle

Madonna

Nina

Susceptible

Kelvedon Wonder

Rondo

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| 11.3 | Validation of test | evaluation of variety resistance should be calibrated with results of resistant and susceptible controls |
| 11.4 | Off-types | - |
| 12. | Interpretation of data in terms of UPOV characteristic states |  |
|  | absent [1]present [9] | susceptibleresistant |
| 13. | Critical control points | - |

[End of document]

1. matref@geves.fr [↑](#footnote-ref-2)
2. Marian.McEwan@sasa.gsi.gov.uk [↑](#footnote-ref-3)
3. Harmores 2 CPVO project (<http://www.cpvo.europa.eu/main/en/home/documents-and-publications/technical-projects-reports>) [↑](#footnote-ref-4)