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Lactuca sativa L.

GUIDELINES

FOR THE CONDUCT OF TESTS

FOR DISTINCTNESS, UNIFORMITY AND STABILITY

*prepared by experts from the Netherlands
 to be considered by the
 Enlarged Editorial Committee
 at its meeting, to be held in Geneva,
 from 2017-01-11 to 2017-01-12*

Disclaimer: this document does not represent UPOV policies or guidance

Alternative names:*

Botanical name	English	French	German	Spanish
<i>Lactuca sativa</i> L.	Lettuce	Laitue	Salat	Lechuga

The purpose of these guidelines ("Test Guidelines") is to elaborate the principles contained in the General Introduction (document TG/1/3), and its associated TGP documents, into detailed practical guidance for the harmonized examination of distinctness, uniformity and stability (DUS) and, in particular, to identify appropriate characteristics for the examination of DUS and production of harmonized variety descriptions.

ASSOCIATED DOCUMENTS

These Test Guidelines should be read in conjunction with the General Introduction and its associated TGP documents.

* These names were correct at the time of the introduction of these Test Guidelines but may be revised or updated. [Readers are advised to consult the UPOV Code, which can be found on the UPOV Website (www.upov.int), for the latest information.]

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1. Subject of these Test Guidelines

These Test Guidelines apply to all varieties of *Lactuca sativa* L.

2. Material Required

2.1 The competent authorities decide on the quantity and quality of the plant material required for testing the variety and when and where it is to be delivered. Applicants submitting material from a State other than that in which the testing takes place must ensure that all customs formalities and phytosanitary requirements are complied with.

2.2 The material is to be supplied in the form of seed.

2.3 The minimum quantity of plant material, to be supplied by the applicant, should be:

15,000 seeds

The seed should meet the minimum requirements for germination, species and analytical purity, health and moisture content, specified by the competent authority. In cases where the seed is to be stored, the germination capacity should be as high as possible and should, be stated by the applicant.

2.4 The plant material supplied should be visibly healthy, not lacking in vigor, nor affected by any important pest or disease.

2.5 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If it has been treated, full details of the treatment must be given.

3. Method of Examination

3.1 *Number of Growing Cycles*

The minimum duration of tests should normally be two independent growing cycles.

3.2 *Testing Place*

Tests are normally conducted at one place. In the case of tests conducted at more than one place, guidance is provided in TGP/9 "Examining Distinctness".

3.3 *Conditions for Conducting the Examination*

The tests should be carried out under conditions ensuring satisfactory growth for the expression of the relevant characteristics of the variety and for the conduct of the examination.

3.4 *Test Design*

3.4.1 Each test should be designed to result in a total of at least 60 plants, which should be divided between at least 2 replicates.

3.4.2 The design of the tests should be such that plants or parts of plants may be removed for measurement or counting without prejudice to the observations which must be made up to the end of the growing cycle.

3.5 *Additional Tests*

Additional tests, for examining relevant characteristics, may be established.

4. Assessment of Distinctness, Uniformity and Stability

4.1 *Distinctness*

4.1.1 General Recommendations

It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding distinctness. However, the following points are provided for elaboration or emphasis in these Test Guidelines.

4.1.2 Consistent Differences

The differences observed between varieties may be so clear that more than one growing cycle is not necessary. In addition, in some circumstances, the influence of the environment is not such that more than a single growing cycle is required to provide assurance that the differences observed between varieties are sufficiently consistent. One means of ensuring that a difference in a characteristic, observed in a growing trial, is sufficiently consistent is to examine the characteristic in at least two independent growing cycles.

4.1.3 Clear Differences

Determining whether a difference between two varieties is clear depends on many factors, and should consider, in particular, the type of expression of the characteristic being examined, i.e. whether it is expressed in a qualitative, quantitative, or pseudo-qualitative manner. Therefore, it is important that users of these Test Guidelines are familiar with the recommendations contained in the General Introduction prior to making decisions regarding distinctness.

4.1.4 Number of plants or parts of plants to be Examined

Unless otherwise indicated, for the purposes of distinctness, all observations on single plants should be made on 20 plants or parts of plants taken from each of 20 plants and any other observations made on all plants in the test, disregarding any off-type plants.

4.1.5 Method of Observation

The recommended method of observing the characteristic for the purposes of distinctness is indicated by the following key in the second column of the Table of Characteristics (see document TGP/9 "Examining Distinctness", Section 4 "Observation of characteristics"):

MG: single measurement of a group of plants or parts of plants

MS: measurement of a number of individual plants or parts of plants

VG: visual assessment by a single observation of a group of plants or parts of plants

VS: visual assessment by observation of individual plants or parts of plants

Type of observation: visual (V) or measurement (M)

"Visual" observation (V) is an observation made on the basis of the expert's judgment. For the purposes of this document, "visual" observation refers to the sensory observations of the experts and, therefore, also includes smell, taste and touch. Visual observation includes observations where the expert uses reference points (e.g. diagrams, example varieties, side-by-side comparison) or non-linear charts (e.g. color charts). Measurement (M) is an objective observation against a calibrated, linear scale e.g. using a ruler, weighing scales, colorimeter, dates, counts, etc.

Type of record: for a group of plants (G) or for single, individual plants (S)

For the purposes of distinctness, observations may be recorded as a single record for a group of plants or parts of plants (G), or may be recorded as records for a number of single, individual plants or parts of plants (S). In most cases, "G" provides a single record per variety and it is not possible or necessary to apply statistical methods in a plant-by-plant analysis for the assessment of distinctness.

In cases where more than one method of observing the characteristic is indicated in the Table of Characteristics (e.g. VG/MG), guidance on selecting an appropriate method is provided in document TGP/9, Section 4.2.

4.2 *Uniformity*

- 4.2.1 It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding uniformity. However, the following points are provided for elaboration or emphasis in these Test Guidelines:
- 4.2.2 For the assessment of uniformity of seed-propagated varieties, a population standard of 1% and an acceptance probability of at least 95% should be applied. In the case of a sample size of 60 plants, 2 off-types are allowed.

4.3 *Stability*

- 4.3.1 In practice, it is not usual to perform tests of stability that produce results as certain as those of the testing of distinctness and uniformity. However, experience has demonstrated that, for many types of variety, when a variety has been shown to be uniform, it can also be considered to be stable.
- 4.3.2 Where appropriate, or in cases of doubt, stability may be further examined by testing a new seed stock to ensure that it exhibits the same characteristics as those shown by the initial material supplied.

5. Grouping of Varieties and Organization of the Growing Trial

- 5.1 The selection of varieties of common knowledge to be grown in the trial with the candidate varieties and the way in which these varieties are divided into groups to facilitate the assessment of distinctness are aided by the use of grouping characteristics.
- 5.2 Grouping characteristics are those in which the documented states of expression, even where produced at different locations, can be used, either individually or in combination with other such characteristics: (a) to select varieties of common knowledge that can be excluded from the growing trial used for examination of distinctness; and (b) to organize the growing trial so that similar varieties are grouped together.
- 5.3 The following have been agreed as useful grouping characteristics:
- (a) Seed: color (characteristic 1)
 - (b) Leaf: anthocyanin coloration (characteristic 11)
 - (c) Time of beginning of bolting (characteristic 35)
 - (d) Resistance to *Bremia lactucae* (Bl) isolate Bl: 16 (characteristic 38)

In the first place, the collection should be divided according to types as mentioned in the Table 1 on the next page below. In cases of doubt to which type a variety belongs to, it should be tested in all relevant types.

- 5.4 Guidance for the use of grouping characteristics, in the process of examining distinctness, is provided through the General Introduction and document TGP/9 "Examining Distinctness".

Table 1

Type	Example varieties	Plant: degree of overlapping of upper part of leaves (Char. 3)	Leaf: number of divisions (Char. 6)	Leaf: thickness (Char. 17)	Leaf: undulation of margin (Char. 20)	Leaf: venation (Char. 25)	Only varieties with degree of overlapping of upper part of leaves medium or strong: Head: shape in longitudinal section (Char. 27)
Butterhead type	Clarion, Maikönig, Sartre	medium to strong	absent or very few	thin to thick	absent to weak	not flabellate	circular or narrow oblate
Novita type	Norvick	absent or weak	absent or very few	thin to medium	very weak to medium	flabellate	-
Iceberg type	Great Lakes 659, Roxette, Saladin, Vanguard 75	strong	absent or very few	thick	absent to medium	flabellate	circular or narrow oblate
Batavia type	Aquarel, Curtis, Funnice, Felucca, Grand Rapids, Masaida, Visyon	absent or weak to strong	absent or very few	medium to thick	weak to very strong	flabellate	broad elliptic, circular or narrow oblate
Frisée d'Amérique type	Bijou, Blonde à couper améliorée	absent or weak	absent or very few	thin	absent to strong	flabellate or not flabellate or semi	-
Lollo type	Lollo rossa, Revolution	absent or weak	absent or very few	thin	strong to very strong	flabellate	-
Oakleaf type	Catalogna, Kipling, Muraï, Salad Bowl	absent or weak	few to many	thin	absent to weak	flabellate or not flabellate or semi	-
Multi-divided type	Curletta, Duplex, Jadigon, Rodagio	absent or weak	medium to very many	thin	weak to very strong	flabellate	-
Frillice type	Frilett	absent or weak	absent or very few	thick	weak to strong	flabellate	-
Cos type	Actarus, Blonde maraîchère, Pinokkio	absent or weak to medium	absent or very few	medium to thick	absent to weak	not flabellate	narrow elliptic
Gem type	Craquerelle du Midi, Sucrine, Xanadu	absent or weak to medium	absent or very few	medium to thick	absent to weak	not flabellate	broad elliptic, circular or narrow oblate
Stem type	Celtuce, Guasihong	absent or weak	absent or very few	thin to medium	absent to weak	not flabellate	-

6. Introduction to the Table of Characteristics

6.1 *Categories of Characteristics*

6.1.1 Standard Test Guidelines Characteristics

Standard Test Guidelines characteristics are those which are approved by UPOV for examination of DUS and from which members of the Union can select those suitable for their particular circumstances.

6.1.2 Asterisked Characteristics

Asterisked characteristics (denoted by *) are those included in the Test Guidelines which are important for the international harmonization of variety descriptions and should always be examined for DUS and included in the variety description by all members of the Union, except when the state of expression of a preceding characteristic or regional environmental conditions render this inappropriate.

6.2 *States of Expression and Corresponding Notes*

6.2.1 States of expression are given for each characteristic to define the characteristic and to harmonize descriptions. Each state of expression is allocated a corresponding numerical note for ease of recording of data and for the production and exchange of the description.

6.2.2 In the case of qualitative and pseudo-qualitative characteristics (see Chapter 6.3), all relevant states of expression are presented in the characteristic. However, in the case of quantitative characteristics with 5 or more states, an abbreviated scale may be used to minimize the size of the Table of Characteristics. For example, in the case of a quantitative characteristic with 9 states, the presentation of states of expression in the Test Guidelines may be abbreviated as follows:

State	Note
small	3
medium	5
large	7

However, it should be noted that all of the following 9 states of expression exist to describe varieties and should be used as appropriate:

State	Note
very small	1
very small to small	2
small	3
small to medium	4
medium	5
medium to large	6
large	7
large to very large	8
very large	9

6.2.3 Further explanation of the presentation of states of expression and notes is provided in document TGP/7 "Development of Test Guidelines".

6.3 *Types of Expression*

An explanation of the types of expression of characteristics (qualitative, quantitative and pseudo-qualitative) is provided in the General Introduction.

6.4 *Example Varieties*

Where appropriate, example varieties are provided to clarify the states of expression of each characteristic.

6.5 Legend

	English			français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
1	2	3	4	5	6	7		
	Name of characteristics in English			Nom du caractère en français	Name des Merkmals auf Deutsch	Nombre del carácter en español		
	states of expression			types d'expression	Ausprägungsstufen	tipos de expresión		

- 1 Characteristic number
- 2 (*) Asterisked characteristic – see Chapter 6.1.2
- 3 Type of expression
 QL Qualitative characteristic – see Chapter 6.3
 QN Quantitative characteristic – see Chapter 6.3
 PQ Pseudo-qualitative characteristic – see Chapter 6.3
- 4 Method of observation (and type of plot, if applicable)
 MG, MS, VG, VS – see Chapter 4.1.5
- 5 (+) See Explanations on the Table of Characteristics in Chapter 8.2
- 6 (a)-(c) See Explanations on the Table of Characteristics in Chapter 8.1
- 7 Not applicable

7. Table of Characteristics/Tableau des caractères/Merkmalstabelle/Tabla de caracteres

	English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
1. (*)	PQ VG					
	Seed: color					
	white				Verpia	1
	yellow				Durango	2
	brown				Oaklin	3
	black				Kagraner Sommer 2	4
2. (*)	QN MS/VG					
	Plant: diameter	Plante : diamètre	Pflanze: Durchmesser	Planta: diámetro		
	very small	très petit	sehr klein	muy pequeño	Tom Thumb	1
	small	petit	klein	pequeño	Gotte à graine blanche	3
	medium	moyen	mittel	medio	Clarion, Verpia	5
	large	grand	groß	grande	Great Lakes 659	7
	very large	très grand	sehr groß	muy grande	El Toro	9
3. (*)	QN VG	(+)	(a), (b)			
	Plant: degree of overlapping of upper part of leaves					
	absent or weak				Actarus, Aquarel, Blonde à couper améliorée, Curtis, Lollo rossa	1
	medium				Augusta, Clarion, Fiorella	2
	strong				Roxette, Vanguard 75	3
4.	QN MS/VG	(+)	(b)			
	Only varieties with degree of overlapping of upper part of leaves absent or weak: Plant: number of leaves					
	few				Lollo rossa	3
	medium				Muraï	5
	many				Felucca, Sartre, Xandra	7

	English		français		deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
5.	QN	VG	(+)	(c)				
	Leaf: attitude		Feuille : port		Blatt: Haltung	Hoja: porte		
	erect		dressé		aufrecht	erecto	Feria, Pinokkio	1
	semi-erect		demi-dressé		halbaufrecht	semierecto	Expedition, Sartre	3
	horizontal		horizontal		waagerecht	horizontal	Divina	5
6. (*)	QN	VG	(+)	(a), (c)				
	Leaf: number of divisions							
	absent or very few						Fiorella, Lollo rossa	1
	few						Curletta, Rodagio	3
	medium						Ezabel, Jadigon	5
	many						Expedition, Multired 54	7
	very many						Excite, Ezfrill, Telex	9
7.	PQ	VG	(+)	(c)				
	Only varieties with divisions absent or very few: Leaf: shape							
	triangular							1
	lanceolate						Qingyuanewoju	2
	medium oblate						Stylist	3
	narrow oblate						Commodore, Fiorella	4
	circular						Verpia	5
	broad elliptic						Amadeus	6
	medium elliptic						Xanadu	7
	narrow elliptic						Verte maraîchère	8
	linear						Hongwoju	9
	broad obtrullate							10
	obovate						Raisa	11
	oblanceolate						Xiangshengcai	12

	English		français		deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota	
8.	PQ	VG	(+)	(c)					
	Only varieties with divisions absent or very few: Leaf: shape of apex								
	acute						Celtuce	1	
	obtuse						Actarus	2	
	rounded						Blonde maraîchère, Maserati	3	
	obcordate						PS 6545691	4	
9.	QN	VG		(c)					
	Only varieties with divisions absent or very few: Leaf: cross section		Feuille : forme en section transversale		Blatt: Form im Querschnitt		Hoja: forma en sección transversal		
	concave		concave		konkav		cóncava	Sunstar	1
	flat		plate		flach		plana	Clarion, Lollo rossa	3
	convex							Tiago	5
10.	QN	VG	(+)	(c)					
	Only Oakleaf type varieties: Leaf: width of lobes								
	narrow							Kibrille, Rougini	3
	medium							Bandolin, Ribaï	5
	broad							Horix, Starix, Vizir	7
11. (*)	QN	VG	(+)	(c)					
	Leaf: anthocyanin coloration		Feuille : pigmentation anthocyanique		Blatt: Anthocyanfärbung		Hoja: pigmentación antocianica		
	absent or very weak		absente ou très faible		fehlend oder sehr gering		ausente o muy débil	Clarion	1
	weak		faible		gering		débil	Du bon jardinier	3
	medium		moyenne		mittel		media	Lollo rossa, Luana	5
	strong		forte		stark		fuerte	Merveille des quatre saisons	7
	very strong		très forte		sehr stark		muy fuerte	Iride, Revolution	9

	English		français		deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
12. (*)	PQ	VG	(+)	(c)				
	Leaf: hue of anthocyanin coloration							
	reddish						Lollo rossa	1
	purplish						Iride	2
	brownish						Luana, Maravilla de Verano	3
13.	QN	VG	(+)	(c)				
	Leaf: area covered by anthocyanin coloration		Feuille : surface couverte par la pigmentation anthocyanique	Blatt: Größe der Anthocyanfärbung	Hoja: área cubierta por la pigmentación antocianica			
	very small						Steirer Krauthauptel	1
	small		petite	klein	pequeña		Diablo	3
	medium		moyenne	mittel	media		Luana	5
	large		grande	groß	grande		Merveille des quatre saisons	7
	very large						Bijou, Revolution	9
14. (*)	PQ	VG	(+)	(c)				
	Leaf: color							
	green						Verpia	1
	yellowish green						Dorée de printemps	2
	greyish green						Celtuce, Du bon jardinier	3
15. (*)	QN	VG	(+)	(c)				
	Leaf: intensity of green color		Feuille: intensité de couleur verte	Blatt: Intensität der Grünfärbung	Hoja: intensidad del color verde			
	very light		très claire	sehr hell	muy clara			1
	light		claire	hell	clara		Blonde maraîchère, Lollo Bionda	3
	medium		moyenne	mittel	media		Aquarel, Clarion	5
	dark		foncée	dunkel	oscura		Expedition, Verpia	7
	very dark		très foncée	sehr dunkel	muy oscura		Pascal, Verdatrix	9

	English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
16.	QN	VG	(c)				
	Leaf: glossiness of upper side		Feuille: brillance de la face supérieure	Blatt: Glanz der Oberseite	Hoja: brillo de la parte superior		
	absent or very weak		nulle ou très faible	fehlend oder sehr gering	ausente o muy débil	Divina, Du bon jardinier	1
	weak		faible	gering	débil	Duplex, Fiorella, Sartre	3
	medium		moyenne	mittel	medio	Funnice	5
	strong		forte	stark	fuerte	Noisette, Redair	7
	very strong		très forte	sehr stark	muy fuerte	Bijou	9
17. (*)	QN	VG	(a), (c)				
	Leaf: thickness		Feuille: épaisseur	Blatt: Dicke	Hoja: espesor		
	thin		mince	dünn	delgado	Bijou, Lollo rossa, Raisa	3
	medium		moyen	mittel	medio	Curtis, Expedition	5
	thick		épais	dick	grueso	Frilett, Roxette	7
18. (*)	QN	VG	(c)				
	Leaf: blistering		Feuille : cloqûre	Blatt: Blasigkeit	Hoja: abullonado		
	absent or very weak		nulle ou très faible	fehlend oder sehr gering	ausente o muy débil	Duplex, Sartre	1
	weak		faible	gering	débil	Fiorella	3
	medium		moyenne	mittel	medio	Commodore	5
	strong		forte	stark	fuerte	Blonde de Paris, Xanadu	7
	very strong		très forte	sehr stark	muy fuerte	Blonde de Doulon, Iride, Karioka	9
19.	QN	VG/VS	(+)	(c)			
	Leaf: size of blisters						
	small					Dorée de printemps, Rodagio	3
	medium					Clarion	5
	large					Fiorella	7
20. (*)	QN	VG/VS	(+)	(a), (c)			
	Leaf: undulation of margin		Feuille : ondulation du bord	Blatt: Randwellung	Hoja: ondulación del borde		
	absent or very weak		absente ou très faible	fehlend oder sehr gering	ausente o muy débil	Tiago	1
	weak		faible	gering	débil	Commodore	3
	medium		moyenne	mittel	media	Noisette, Pentared	5
	strong		forte	stark	fuerte	Calmar, Invicta	7
	very strong		très forte	sehr stark	muy fuerte	Lollo rossa	9

	English		français		deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
21.	PQ	VG	(+)	(c)				
	Leaf: type of incisions of margin							
	crenate						Gloire du Dauphiné	1
	regularly dentate						Soliflore	2
	irregularly dentate						Rodagio	3
	bidentate						Great Lakes 118	4
	tridentate						Expedition	5
22.	QN	VG	(+)	(c)				
	Leaf: depth of incisions of margin							
	absent or very shallow						Actarus, Clarion, Tiago	1
	shallow						Pentared, Unicum	3
	medium						Santarinas	5
	deep						Expedition	7
	very deep							9
23.	QN	VG	(+)	(c)				
	<u>Only varieties with type of incisions irregularly dentate, bi- or tridentate:</u> Leaf: depth of secondary incisions of margin							
	shallow						Great Lakes 659	3
	medium						Expedition	5
	deep							7
24.	QN	VG	(+)	(c)				
	Leaf: density of incisions of margin							
	very sparse							1
	sparse						Maravilla de Verano	3
	medium						Calmar	5
	dense						Grand Rapids	7
	very dense						Locarno	9

	English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
25. (*)	QN	VG	(+)	(a), (c)			
	Leaf: venation						
	not flabellate					Verpia, Xanadu	1
	semi flabellate					Kibrille, Muraï	2
	flabellate					Locarno, Roxette	3
26.	QN	MS/VG		(b)			
	Only varieties with degree of overlapping of upper part of leaves medium or strong: Head: size						
	very small					Tom Thumb	1
	small					Xanadu	3
	medium					Fiorella, Soraya	5
	large					Great Lakes 659	7
	very large					Blonde maraîchère, El Toro	9
27. (*)	QN	MS/VG	(+)	(a), (b)			
	Only varieties with degree of overlapping of upper part of leaves medium or strong: Head: shape in longitudinal section						
	narrow elliptic					Verte maraîchère	1
	broad elliptic					Amadeus, Sucrine	2
	circular					Verpia	3
	narrow oblate					Ametist	4
28.	QN	VG		(b)			
	Only varieties with degree of overlapping of upper part of leaves medium or strong: Head: density						
	loose					Nanda	3
	medium					Daguan, Delice	5
	dense					Atella, Islandia	7
	very dense					Rubette	9

	English		français		deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
29.	QN	MS/VG	(+)	(b)				
	Only Stem type varieties: Stem: length							
	short						Wuweijianye	3
	medium						Zipixiang	5
	long						Guasihong	7
30.	QN	MS/VG	(+)	(b)				
	Only Stem type varieties: Stem: width							
	small						Ailaowoju	1
	medium						Guasihong, Zipixiang	2
	large						Guasihong	3
31.	PQ	VG	(+)	(b)				
	Only Stem type varieties: Stem: shape in longitudinal section							
	cylindrical						Chiwoju	1
	conical						Guasihong	2
	fusiform						Zipixiang	3
32.	PQ	VG		(b)				
	Only Stem type varieties: Stem: color							
	whitish green						Wuweijianye	1
	light green						Chiwoju	2
	medium green						Yangwoju	3
	greenish purple						Guasihong	4
	purplish red						Hongwosun	5
33.	PQ	VG		(b)				
	Only Stem type varieties: Stem: color of flesh							
	yellowish white						Wuweijianye	1
	whitish green						Chiwoju	2
	light green						Yangwoju	3
	medium green						Guasihong	4
	dark green						Chiwosun	5

	English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
34.	QN MG/VG					
	Only varieties with degree of overlapping of upper part of leaves medium or strong: Time of harvest maturity					
	very early				Gotte jaune d'or	1
	early				Pantlika, Sucrine	3
	medium				Clarion	5
	late				Blonde maraîchère, Calmar	7
	very late				El Toro, Pinokkio	9
35. (*)	QN MG/VG	(+)				
	Time of beginning of bolting					
	very early				Blonde à couper améliorée	1
	early				Gotte à graine blanche	3
	medium				Pantlika	5
	late				Hilde II	7
	very late				Erika, Roxette	9
36.	QN VG	(+)				
	Axillary sprouting					
	absent or weak				Claridia, Shotter, Valmaine, Xanadu	1
	medium				Actarus	2
	strong				Amible, Bassoon	3
37.	QN VG	(+)				
	Bolting stem: fasciation					
	absent or very weak				Aquarel, Gotte à graine blanche	1
	weak				Verte maraîchère	3
	medium				Amadeus	5
	strong				Rougini	7
	very strong				Sartre, Verdatrix	9

	English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
38.	QL	VG	(+)				
	Resistance to <i>Bremia lactucae</i> (BI) isolate BI: 16						
	absent					Green Towers	1
	present					Argelès	9
39.	QL	VG	(+)				
	Resistance to <i>Bremia lactucae</i> (BI) isolate BI: 17						
	absent					Green Towers	1
	present					Argelès	9
40.	QL	VG	(+)				
	Resistance to <i>Bremia lactucae</i> (BI) isolate BI: 20						
	absent					Green Towers	1
	present					FrRsal-1	9
41.	QL	VG	(+)				
	Resistance to <i>Bremia lactucae</i> (BI) isolate BI: 21						
	absent					Green Towers	1
	present					Argelès, Colorado	9
42.	QL	VG	(+)				
	Resistance to <i>Bremia lactucae</i> (BI) isolate BI: 22						
	absent					Green Towers	1
	present					FrRsal-1	9
43.	QL	VG	(+)				
	Resistance to <i>Bremia lactucae</i> (BI) isolate BI: 23						
	absent					Green Towers	1
	present					Colorado	9

	English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
44.	QL	VG	(+)				
	Resistance to <i>Bremia lactucae</i> (BI) isolate BI: 24						
	absent					Argelès, Colorado	1
	present					Dandie, NunDm15, UCDm14	9
45.	QL	VG	(+)				
	Resistance to <i>Bremia lactucae</i> (BI) isolate BI: 25						
	absent					Colorado	1
	present					Argelès	9
46.	QL	VG	(+)				
	Resistance to <i>Bremia lactucae</i> (BI) isolate BI: 26						
	absent					Colorado	1
	present					Balesta, Bedford	9
47.	QL	VG	(+)				
	Resistance to <i>Bremia lactucae</i> (BI) isolate BI: 27						
	absent					Balesta, Colorado	1
	present					FrRsal-1	9
48.	QL	VG	(+)				
	Resistance to <i>Bremia lactucae</i> (BI) isolate BI: 29						
	absent					Argelès	1
	present					Balesta	9
49.	QL	VG	(+)				
	Resistance to <i>Bremia lactucae</i> (BI) isolate BI: 30						
	absent					Argelès, Colorado	1
	present					Balesta	9

	English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
50.	QL	VG	(+)				
	Resistance to <i>Bremia lactucae</i> (BI) isolate BI: 31						
	absent					Colorado, RYZ910457	1
	present					Argelès, Balesta	9
51.	QL	VG	(+)				
	Resistance to <i>Lettuce mosaic virus</i> (LMV) pathotype II						
	absent					Bijou, Hilde II, Sprinter, Sucrine	1
	present					Capitan, Corsica	9
52.	QL	MS/VG	(+)				
	Resistance to <i>Nasonovia ribisnigri</i> (Nr) biotype Nr: 0						
	absent					Abel, Green Towers, Nadine	1
	present					Barcelona, Bedford, Dynamite, Silvinas	9
53.	QN	MS/VG	(+)				
	Resistance to <i>Fusarium oxysporum</i> f.sp. <i>lactucae</i> (Fol) race 1						
	susceptible					Cobham Green, Patriot	1
	moderately resistant					Affic, Fuzila, Natexis	2
	highly resistant					Costa Rica No. 4, Romasol	3

8. Explanations on the Table of Characteristics

8.1 *Explanations covering several characteristics*

Characteristics containing the following key in the second column of the Table of Characteristics should be examined as indicated below:

(a) See also 5.3 for a table to determine the type using several characteristics.



Heading; thin to rather thick, tender leaves with a clear midrib; leaf shape circular to transverse broad elliptic; in general no incised margin; head shape ranging from broad elliptic to transvers elliptic.

Butterhead type



Cross between Butterhead and Iceberg type for glasshouse growing. Open heading; leaf structure like Butterhead, incisions of the margin as Iceberg.

Novita type



Heading with strong or very strong overlapping of upper part of leaves; thick and crispy leaves, predominantly green and greyish green, leaf margin hardly to rather strongly incised, no clear midrib but with flabellate venation.

Iceberg type



Open to strong heading; generally medium thick, rather strongly blistered leaves, predominately yellowish or medium green; leaf margin with weak to strong undulation.

Batavia type



Frisée d'Amérique type

Non-heading, loose, generally quite large plant; thin leaves. Compared to Lollo type in general less undulating margin and showing more leaf blade. Compared to Batavia type, leaves are thinner. Mainly used for babyleaf production.



Lollo type

Non-heading; thin leaves with strongly undulated leaf margin. The plant as a whole shows mainly the undulating leaf margins. In general strongly blistered leaves, blisters are rather small.



Oakleaf type

Thin, divided leaves; divisions have an oakleaf or lobed shape with in general a rounded tip. Radichetta or Catalogna with acute tip of the division. Heart can be loose to dense.



Multi-divided type

Non-heading; thin, medium to very strong divided leaves. Tip of divisions can be undulated and incised. Plant may look as a Lollo type, but leaves are always divided.



Frillice type

Non-heading; thick, crispy leaves, sometimes weakly divided. Clearly incised leaf margin.



Cos type

Elongated and rather tough leaves with a clear midrib, head shape in longitudinal section elliptic, length of head $>1.5 \times$ diameter; heading can be very late.



Gem type

Tough leaves with clear midrib, head shape short elliptic to slightly obovate. Some types only have a tightly filled heart, others are more similar to a short Cos type. Suitable for semi-arid conditions.



Stem type

Forms a fleshy stem before bolting, at least under (semi-)short day conditions; leaves are mainly tough and have a clear midrib. Leaves and/or stem are consumed.

- (b) Plant, head and stem: Observations should be made at harvest maturity. For varieties with degree of overlapping absent or weak observations should be made just before deterioration and before bolting.
- (c) Leaf: For varieties with degree of overlapping medium or strong observations should be made on the largest outer leaves, at harvest maturity. For varieties with degree of overlapping absent or weak observations should be made on the largest leaves, just before deterioration and before bolting. For Stem type varieties observations should be made on leaves at the middle third of the stem, just before deterioration and before bolting.

8.2 Explanations for individual characteristics

Ad. 3: Plant: degree of overlapping of upper part of leaves

Observations should be made on leaves at the heart of the plant to form a head.



1
absent or weak



2
medium



3
strong

Ad. 4: Only varieties with degree of overlapping of upper part of leaves absent or weak: Plant: number of leaves

In case of doubt, observations can be made by cutting the plant in half.



3
few



5
medium



7
many

Ad. 5: Leaf: attitude



1
erect



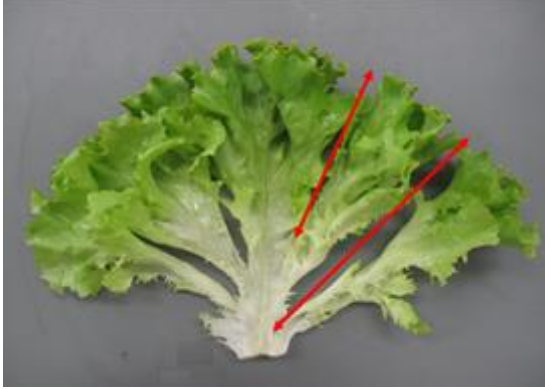
3
semi-erect



5
horizontal

Ad. 6: Leaf: number of divisions

Observations should be made only on the incisions more than halfway to the midrib of the whole leaf.



1
absent or very few



3
few



5
medium






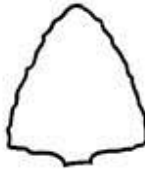


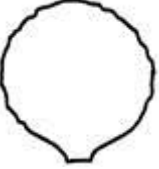
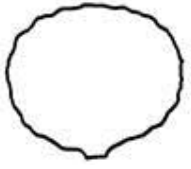
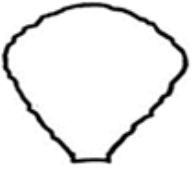
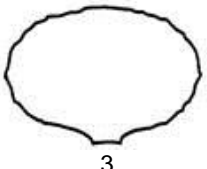


7
many



9
very many

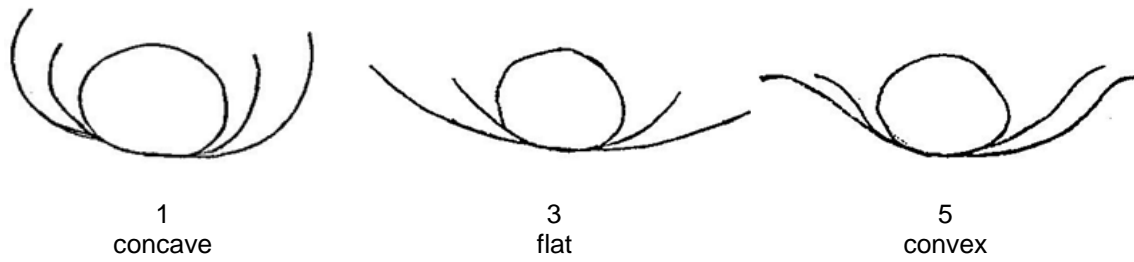
Ad. 7: Only varieties with divisions absent or very few: Leaf: shape

width (ratio length/width)	< broadest part >		
	below middle	at middle	above middle
narrow (high)		 9 linear	
	 2 lanceolate	 8 narrow elliptic	 12 oblanceolate
		 7 medium elliptic	
	 1 triangular	 6 broad elliptic	 11 obovate
medium (medium)		 5 circular	
		 4 narrow oblate	 10 broad obtrullate
broad (low)		 3 medium oblate	

Ad. 8: Only varieties with divisions absent or very few: Leaf: shape of apex



Ad. 9: Only varieties with divisions absent or very few: Leaf: cross section



Ad. 10: Only Oakleaf type varieties: Leaf: width of lobes



Ad. 11: Leaf: anthocyanin coloration

Ad. 12: Leaf: hue of anthocyanin coloration

Anthocyanin coloration (Char. 11)	Hue of anthocyanin coloration (Char. 12)		
	1 reddish	2 purplish	3 brownish
1 absent or very weak	Clarion		
3 weak	Du bon jardinier, Steirer Krauthauptel		Brauner Troztkopf, Diablo, Maravilla de Verano
5 medium	Lollo rossa		Frisée d'Amérique, Luana, New Red Fire, Salad bowl rossa
7 strong	Jadigon		Duplex, Merveille des quatre saisons
9 very strong	Revolution	Iride	Multired 54

Ad. 13: Leaf: area covered by anthocyanin coloration

Observations should be made on the total area of diffused and/or localised anthocyanin coloration.



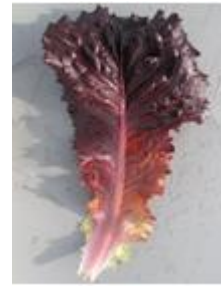
3
small



5
medium



7
large



9
very large

Ad. 14: Leaf: color

Ad. 15: Leaf: intensity of green color

Only to observe for green varieties and for two-colored varieties with 'Leaf: area covered by anthocyanin coloration' less than large (less than note 7 to 9), so the green color of the leaf can be observed without picking a leaf from the plant.

Intensity of green color (Char. 15)	Color (Char. 14)		
	1 green	2 yellowish green	3 greyish green
1 very light			
3 light	Blonde maraîchère, New Red Fire	Lollo Bionda, Steirer Krauthauptel	Celtuce
5 medium	Ballerina	Aquarel, Australische Gele, Dorée de printemps	Clarion, Du bon jardinier, Durango
7 dark	Actarus, Baby Star, Expedition, Verpia		Webbs Wonderful
9 very dark	Pascal, Verdatrix		

Ad. 19: Leaf: size of blisters

Observations should be made on the whole leaf.



3
small



5
medium



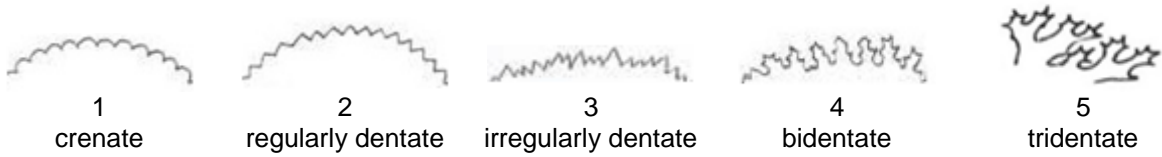
7
large

Ad. 20: Leaf: undulation of margin

Observations should be made on undulation of margin of apical part; also apical part in case of divided leaves.

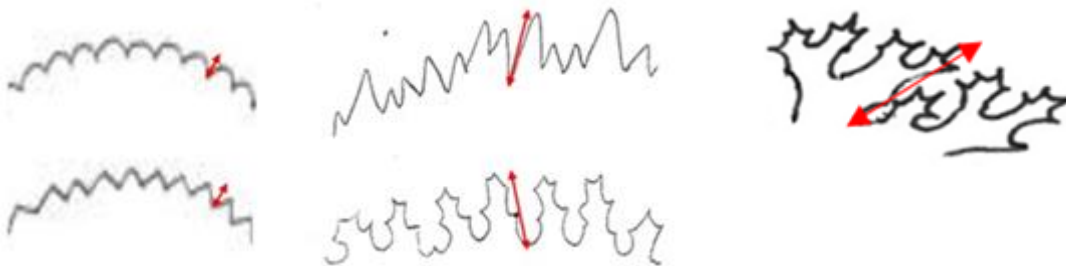
Ad. 21: Leaf: type of incisions of margin

Observations should be made on incisions of the margin at distal half.



Ad. 22: Leaf: depth of incisions of margin

Observations should be made on incisions of the margin at distal half. For varieties with irregularly dentate, bidentate or tridentate incisions describe the deepest incisions and use Char. 23 for the secondary incisions.



Ad. 23: Only varieties with type of incisions irregularly dentate, bi- or tridentate: Leaf: depth of secondary incisions of margin

Observations should be made on secondary incisions of the margin at distal half. In case of tridentate incisions observations should not be made on tertiary incisions of the margin (the most shallow ones).



Ad. 24: Leaf: density of incisions of margin

Observations should be made on all incisions of the margin at distal half, so in case of irregularly dentate or bidentate both primary and secondary incisions, in case or tridentate also tertiary incisions.

Ad. 25: Leaf: venation



1
not flabellate

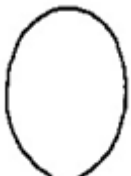


2
semi flabellate



3
flabellate

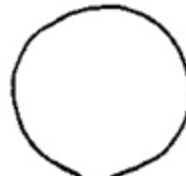
Ad. 27: Only varieties with degree of overlapping of upper part of leaves medium or strong: Head: shape in longitudinal section



1
narrow elliptic



2
broad elliptic

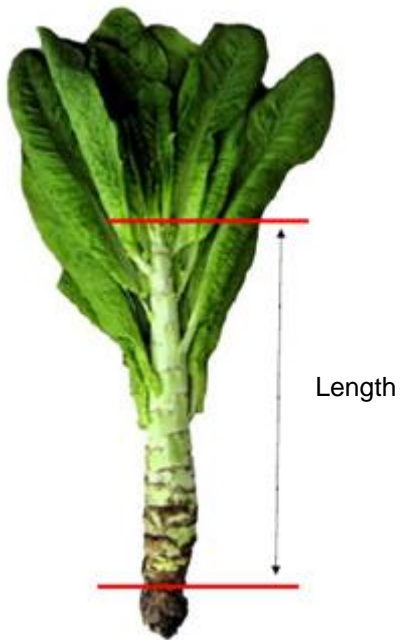


3
circular



4
narrow oblate

Ad. 29: Only Stem type varieties: Stem: length

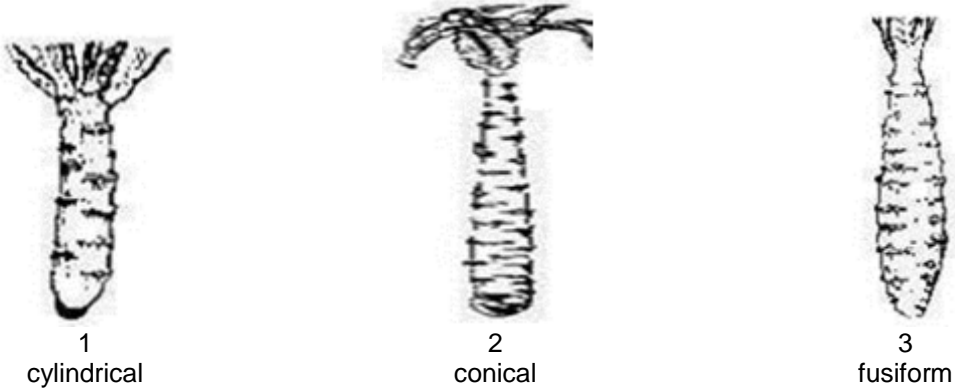


Ad. 30: Only Stem type varieties: Stem: width

Observations should be made on the broadest part of the stem.



Ad. 31: Only Stem type varieties: Stem: shape in longitudinal section



Ad. 35: Time of beginning of bolting

Observations should be made in a trial with more than 12 hours of day light as lettuce varieties need a long photo period to induce bolting.

Observations should be made when 50% of the plants start to bolt. The top of the bolting stem can be seen or felt at the top of the plant.

Ad. 36: Axillary sprouting

Observations should be made at the start of bolting.

Ad. 37: Bolting stem: fasciation

Observations should be made on the stem of bolted plants, not earlier than when the first flowers are open. Varieties with very late time of beginning of bolting and with strong degree of overlapping: the cover leaves of the head may be incised just before deterioration in order to be able to observe fasciation.



Ad. 38 to 50: Resistance to *Bremia lactucae* (Bl), several isolates

1. Pathogen	<i>Bremia lactucae</i>
2. Quarantine status	no
3. Host species	lettuce - <i>Lactuca sativa</i> L.
4. Source of inoculum	GEVES ² (FR) or Naktuinbouw ³ (NL)
5. Isolate	Bl: 16,17, 20-27, 29-31
6. Establishment isolate identity	test on differentials (see table below)
7. Establishment pathogenicity	test on susceptible varieties
8. Multiplication inoculum	
8.1 Multiplication medium	lettuce plantlets
8.2 Multiplication variety	susceptible variety, for example Green Towers. for higher isolates, a variety with defeated resistance may be preferable to keep the isolate fit.
8.3 Plant stage at inoculation	cotyledon to first leaf
8.4 Inoculation medium	tap water
8.5 Inoculation method	spraying a spore suspension
8.6 Harvest of inoculum	washing off from leaves
8.7 Check of harvested inoculum	counting spores
8.8 Shelf life/viability inoculum	2 hours at room temperature; 2 days in fridge
9. Format of the test	
9.1 Number of plants per genotype	at least 20
9.2 Number of replicates	-
9.3 Control varieties	(informative) differentials (see table below)
9.4 Test design	-
9.5 Test facility	climate room
9.6 Temperature	15°C-18°C
9.7 Light	adequate for good plant growth; seedlings should not etiolate. option: reduced light 24 hours after inoculation
9.8 Season	-
9.9 Special measures	plants may grow on wet blotting paper with or without a nutrient solution, on sand or on potting soil (see point 13). high humidity (>90%) is essential for infection and sporulation.
10. Inoculation	
10.1 Preparation inoculum	washing off from leaves by vigorous shaking in a closed container
10.2 Quantification inoculum	counting spores; spore density should be $3 \cdot 10^4$ - $1 \cdot 10^5$
10.3 Plant stage at inoculation	cotyledon stage
10.4 Inoculation method	spraying till run-off. option: reduced light 24 hours after inoculation
10.5 First observation	beginning of sporulation on susceptible varieties (around 7 days after inoculation)
10.6 Second observation	3-4 days after first observation (around 10 days after inoculation)
10.7 Final observations	14 days after inoculation two of these three observations may be sufficient, the third notation is optional for observation of evolution of symptoms in case of doubt. the day of maximum sporulation should occur in this period.
11. Observations	
11.1 Method	visual observation of sporulation and necrotic reaction to infection

² matref@geves.fr

³ resistentie@naktuinbouw.nl

11.2 Observation scale

resistant:

- 0 no sporulation, no necrosis
- 1 no sporulation, necrosis present
- 2 weak sporulation (much less than susceptible control) with necrosis
- 3 weak sporulation (less than susceptible control and not evolving between second and third observation) with necrosis
- 4 very sparse sporulation (not evolving between second and third observation) without necrosis

susceptible:

- 5 reduced sporulation (compared to susceptible control) without necrosis
- 6 normal sporulation without necrosis

11.3 Validation of test

on standards

in case of normal sporulation (same level as susceptible control) with necrosis another test on bigger plants or other substrate must be undertaken.

12. Interpretation of data in terms of UPOV characteristic states

class 0, 1, 2, 3 and 4: resistant

class 5 and 6: susceptible

13. Critical control points

reaction of standards (the infection pressure may vary between experiments, leading to slight differences in sporulation intensity); when the reactions are not clear the experiment should be repeated. the sowing on soil can be used to see necrosis, but weak sporulation (much less than susceptible control) can appear; when testing on sand, spores can be confused with grains of sand. in case of use of nutritive solution on blotting paper, a fungicide can be added to avoid contamination by saprophytes.

For reference: The International Bremia Evaluation Board (IBEB) produces regular updates of the host differential reaction table. The most recent table is available through ISF at <http://www.worldseed.org/our-work/plant-health/other-initiatives/ibeb/>. The table for isolates mentioned in this guideline and illustrations for the observation scale are given.

Isolates	Differentials	Green Towers	Dandie	R4T57D	UC Dm14	NunDm15	CGDm16	Colorado	FtRsal-1	Argelès	RYZ 2164	RYZ910457	Bedford	Balesta	Bartoli	Design
Bl: 16	+	+	+	-	-	+	-	-	-	-	-	-	-	-	-	-
Bl: 17	+	+	-	+	+	-	+	+	-	-	-	(+)	-	-	-	-
Bl: 20	+	+	+	-	-	+	+	-	-	-	-	-	-	-	-	-
Bl: 21	+	+	+	-	+	+	-	+	-	-	-	-	-	-	-	-
Bl: 22	+	-	+	+	+	-	+	-	-	-	-	-	+	-	-	-
Bl: 23	+	+	+	-	-	+	-	-	+	-	-	-	-	-	-	-
Bl: 24	+	-	+	-	-	+	+	-	+	-	-	-	-	-	-	(-)
Bl: 25	+	-	+	-	-	+	+	+	-	-	-	-	-	-	-	-
Bl: 26	+	+	+	-	-	+	+	+	+	-	-	-	-	-	-	-
Bl: 27	+	+	+	+	+	-	+	-	+	+	-	(-)	+	-	-	-
Bl: 29	+	-	+	+	+	+	+	+	+	+	-	-	-	-	-	-
Bl: 30	+	-	+	+	+	-	+	-	+	+	-	-	-	-	-	+
Bl: 31	+	+	+	+	-	-	+	-	-	+	+	-	-	-	-	+

Ad. 51: Resistance to *Lettuce mosaic virus* (LMV) pathotype II

1. Pathogen	Lettuce mosaic virus
2. Quarantine status	no
3. Host species	lettuce - <i>Lactuca sativa</i> L.
4. Source of inoculum	GEVES ⁴ (FR) or Naktuinbouw ⁵ (NL)
5. Isolate	pathotype II (isolates LMV-0 and Ls1 belong to the same pathotype)
6. Establishment isolate identity	resistant and susceptible controls
7. Establishment pathogenicity	susceptible control inoculation
8. Multiplication inoculum	
8.2 Multiplication variety	susceptible control
8.3 Plant stage at inoculation	2-3 leaves
8.4 Inoculation medium	0,05 M PBS, 0,25% (w/v) Na ₂ SO ₃ 0,5% C ₅ H ₁₀ NNaS ₂ .3H ₂ O, 4% carborundum and 5% active charcoal
8.5 Inoculation method	rubbing; optionally repeat after 4 d; 1-2 h high humidity after inoculation
8.6 Harvest of inoculum	homogenized fresh leaf in buffer (50% w/v); freeze-dried leaves can be kept less than 1 year in storage, long term storage at -80°C
8.7 Check of harvested inoculum	compare with mock inoculation with LMV buffer + carborundum + charcoal
8.8 Shelf life/viability inoculum	2 h at 4°C or on ice
9. Format of the test	
9.1 number of plants per genotype	at least 20
9.2 number of replicates	1
9.3 Control varieties	susceptible: Bijou (red), Hilde II (green), Sprinter (green), Sucrine (green) resistant: Capitan (green), Corsica (green), Diveria (red)
9.4 Test design	several mock-inoculated plants in the same tray
9.5 Test facility	climate chamber
9.6 Temperature	after inoculation 15-22°C
9.7 Light	12-16 h light ca. 5000 lux
10. Inoculation	
10.1 Preparation inoculum	fresh leaf ground in fresh LMV buffer incl. carborundum and active charcoal
10.3 Plant stage at inoculation	1 st leaf well-developed at 1 st inoculation, optionally 4 days later 2 nd inoculation
10.4 Inoculation method	rubbing, rinse carborundum off
10.7 Final observations	21 days post inoculation
11. Observations	
11.1 Method	visual estimate of mosaic severity; compare with standards, preferably with standards of same growth type.
11.2 Observation scale	resistant = no symptoms susceptible = growth retardation, young leaves with mosaic, leaf curling
11.3 Validation of test	standards should conform to description
12. Interpretation of data in terms of UPOV characteristic states	classify resistant or susceptible per plant, see 11.2.
13. Critical control points	Sprinter is less susceptible than many other susceptible varieties, this variety can be used to detect low inoculation pressure in a specific experiment. anthocyanin coloration in leaves may mask mosaic symptoms and an earlier observation date for green varieties may be possible, depending on the reaction of the standard varieties in the test.

⁴ matref@geves.fr

⁵ resistentie@naktuinbouw.nl

Ad 52: Resistance to *Nasonovia ribisnigri* (Nr) biotype Nr: 0

1. Pathogen	<i>Nasonovia ribisnigri</i>
2. Quarantine status	no
3. Host species	lettuce - <i>Lactuca sativa</i> L.
4. Source of inoculum	Naktuinbouw ⁶ (NL)
5. Isolate	Nr: 0, preferably red colored biotype
6. Establishment isolate identity	the ends of the legs are black, size 1.5-2.5 mm
7. Establishment pathogenicity	with susceptible control Abel or Green Towers
8. Multiplication inoculum	
8.2 Multiplication variety	Abel or Green Towers
8.3 Plant stage at inoculation	4 to 6 leaves
8.5 Inoculation method	transfer ~5 aphids per plant
8.6 Harvest of inoculum	transfer to Petri-dish; shake off when aphids are numerous carefully remove aphids using a fine painting brush when only few are available
8.7 Check of harvested inoculum	check the black ends of the aphids legs
8.8 Shelf life/viability inoculum	a few hours in shadow
9. Format of the test	
9.1 number of plants per genotype	at least 20
9.2 number of replicates	no
9.3 Control varieties	susceptible: Abel, Green Towers, Nadine resistant: Barcelona, Bedford, Dynamite, Silvinas
9.4 Test design	
9.5 Test facility	glasshouse
9.6 Temperature	after inoculation: 20-22°C, keep below 26°C
9.7 Light	daylight
9.9 Special measures	containment of winged aphids needs special attention
10. Inoculation	
10.1 Preparation inoculum	transfer by shake-off or with brush into Petri-dish
10.3 Plant stage at inoculation	2 to 3 week old seedlings
10.4 Inoculation method	transfer 5 small or medium sized aphids to each plant
10.7 Final observations	15 to 20 days post inoculation
11. Observations	
11.1 Method	count red aphids per plant; if many aphids are present, strong growth reduction can be observed; for this observation, a separate aphid free tent is necessary for blanks
11.2 Observation scale	0 no aphids 1 1-5 aphids 2 6-10 aphids 3 >10 aphids
11.3 Validation of test	controls should be >95% ok; if >5% plants are in class 2 or off-type, the experiment should be repeated
12. Interpretation of data in terms of UPOV characteristic states	0 or 1 resistant 3 susceptible
13. Critical control points	allow sufficient time for the aphids born after inoculation to mature and turn red; as soon as this is the case, the test must be concluded; this may be before 15 days post inoculation. only adult, red aphids are counted; young aphids are transparent and do not count

⁶ resistentie@naktuinbouw.nl

Ad 53: Resistance to *Fusarium oxysporum* f.sp. *lactucae* (Fol) race 1

1. Pathogen	<i>Fusarium oxysporum</i> f.sp. <i>lactucae</i>									
2. Quarantine status	EPPO alert list									
3. Host species	lettuce - <i>Lactuca sativa</i> L.									
4. Source of inoculum	NIAS Genebank ⁷ (JP), CREA-SCS ⁸ (IT), Naktuinbouw ⁹ (NL), GEVES ¹⁰ (FR)									
5. Isolate	Fol: 1									
6. Establishment isolate identity	use microscope and inoculation to lettuce susceptible standard									
7. Establishment pathogenicity	use lettuce susceptible standard									
8. Multiplication inoculum										
8.1 Multiplication medium	inoculation by sowing on contaminated soil: Wheat bran-soil medium inoculation by soaking seedlings: on synthetic liquid medium (e.g. Potatoes Dextrose Broth)									
8.6 Harvest of inoculum	inoculation by sowing on contaminated soil: 7-10 day-old culture inoculation by soaking seedlings: 15 days									
9. Format of the test										
9.1 Number of plants per genotype	at least 30, in case of doubt 60									
9.2 Number of replicates	at least 2									
9.3 Control varieties	susceptible: Cobham Green, Patriot (Cobham Green is slightly less susceptible than Patriot) moderately resistant: Affic, Fuzila, Natexis (Natexis is the lower level of moderate resistance) resistant: Costa Rica No.4, Romasol									
9.4 Test design	include control varieties									
9.5 Test facility	greenhouse or climate room									
9.6 Temperature	25-28 °C (day) / 20 °C (night)									
9.7 Light	under natural day length									
10. Inoculation	two methods can be used for inoculation:									
	<table border="1"> <thead> <tr> <th>sowing seeds on contaminated soil</th> <th>soaking seedlings</th> </tr> </thead> <tbody> <tr> <td>wheat bran-soil medium culture mixed with sterilized soil</td> <td>soaking of roots and of hypocotyl axis for 5 to 15 min in the inoculum suspension</td> </tr> <tr> <td>soil : culture = 20 : 1</td> <td>spores are harvested and adjusted to 10⁶ to 10⁷ sp/ml</td> </tr> <tr> <td>seeds stimulated to emerge (remark: avoid seeds rotted by factors other than pathogen)</td> <td>cotyledons to 2 or 3 leaves appearing</td> </tr> </tbody> </table>		sowing seeds on contaminated soil	soaking seedlings	wheat bran-soil medium culture mixed with sterilized soil	soaking of roots and of hypocotyl axis for 5 to 15 min in the inoculum suspension	soil : culture = 20 : 1	spores are harvested and adjusted to 10 ⁶ to 10 ⁷ sp/ml	seeds stimulated to emerge (remark: avoid seeds rotted by factors other than pathogen)	cotyledons to 2 or 3 leaves appearing
sowing seeds on contaminated soil	soaking seedlings									
wheat bran-soil medium culture mixed with sterilized soil	soaking of roots and of hypocotyl axis for 5 to 15 min in the inoculum suspension									
soil : culture = 20 : 1	spores are harvested and adjusted to 10 ⁶ to 10 ⁷ sp/ml									
seeds stimulated to emerge (remark: avoid seeds rotted by factors other than pathogen)	cotyledons to 2 or 3 leaves appearing									
10.1 Preparation inoculum										
10.2 Quantification inoculum										
10.3 Plant stage at inoculation										
10.4 Inoculation method	two methods can be used, as described above									
10.5 First observation	7- 10 days post inoculation									
10.6 Second observation	14 days post inoculation									
10.7 Final observations	20-25 days post inoculation (sowing or soaking). One or two of these 3 observations may be sufficient. The observation for inoculation by soaking is destructive since stems are cut for the observation of vessels.									
11. Observations										
11.1 Method	visual and/or counting number of plants with symptom; as information calculate a disease index.									






⁷ genebank@nias.affrc.go.jp

⁸ scs.sa@crea.gov.it

⁹ resistentie@naktuinbouw.nl

¹⁰ matref@geves.fr

11.2 Observation scale

inoculation by sowing seeds on contaminated soil	inoculation by soaking seedlings
0: healthy	0: plant without symptoms and healthy vessels 
1: slightly stunting, growing reduction	1: plant with brown vessels only below the cotyledon without yellowing and wilting 
2: severely stunting	2: plant with brown vessels above the cotyledon, without yellowing and wilting 
3: die	3: plant yellowing and wilting, brown vessels 
	4: dead plant 

11.3 Validation of test

results should be compared with results of controls and are depending of the aggressiveness of the test and the distribution of the plants over the categories.

a disease index may be helpful (example for the method of inoculation by soaking seedlings: $DI = (0A + 1B + 2C + 3D + 4E) / (A + B + C + D + E)$, where A to E are number of plants in each category).

12. Interpretation of data in terms of UPOV characteristic states

compare the distribution over the categories with the result of the controls.

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10. Technical Questionnaire

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
		Application date: (not to be filled in by the applicant)
TECHNICAL QUESTIONNAIRE to be completed in connection with an application for plant breeders' rights		
1.	Subject of the Technical Questionnaire	
1.1	Botanical name	<input type="text" value="Lactuca sativa L."/>
1.2	Common name	<input type="text" value="Lettuce"/>
2.	Applicant	
	Name	<input type="text"/>
	Address	<input type="text"/>
	Telephone No.	<input type="text"/>
	Fax No.	<input type="text"/>
	E-mail address	<input type="text"/>
	Breeder (if different from applicant)	<input type="text"/>
3.	Proposed denomination and breeder's reference	
	Proposed denomination (if available)	<input type="text"/>
	Breeder's reference	<input type="text"/>

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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#4. Information on the breeding scheme and propagation of the variety

4.1 Breeding scheme

Variety resulting from:

4.1.1 Crossing

(a) controlled cross []

(b) partially known cross []

(c) unknown cross []

4.1.2 Mutation []

(please state parent variety)

4.1.3 Discovery and development []

(please state where and when discovered and how developed)

4.1.4 Other []

(please provide details)

Authorities may allow certain of this information to be provided in a confidential section of the Technical Questionnaire.

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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4.2	Method of propagating the variety	
4.2.1	Seed-propagated varieties	
(a)	Self-pollination	[]
(b)	Other (please provide details)	[]
	<input type="text"/>	
4.2.2	Other (Please provide details)	[]
	<input type="text"/>	

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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5. Characteristics of the variety to be indicated (the number in brackets refers to the corresponding characteristic in Test Guidelines; please mark the note which best corresponds).

Characteristics	Example Varieties	Note
5.1 Seed: color (1)		
white	Verpia	1 []
yellow	Durango	2 []
brown	Oaklin	3 []
black	Kagraner Sommer 2	4 []
5.2 Leaf: anthocyanin coloration (11)		
absent or very weak	Clarion	1 []
very weak to weak		2 []
weak	Du bon jardinier	3 []
weak to medium		4 []
medium	Lollo rossa, Luana	5 []
medium to strong		6 []
strong	Merveille des quatre saisons	7 []
strong to very strong		8 []
very strong	Iride, Revolution	9 []
5.3 Leaf: intensity of green color (15)		
very light		1 []
very light to light		2 []
light	Blonde maraîchère, Lollo Bionda	3 []
light to medium		4 []
medium	Aquarel, Clarion	5 []
medium to dark		6 []
dark	Expedition, Verpia	7 []
dark to very dark		8 []
very dark	Pascal, Verdatrix	9 []

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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Characteristics	Example Varieties	Note
5.4 Time of beginning of bolting (35)		
very early	Blonde à couper améliorée	1 []
very early to early		2 []
early	Gotte à graine blanche	3 []
early to medium		4 []
medium	Pantlika	5 []
medium to late		6 []
late	Hilde II	7 []
late to very late		8 []
very late	Erika, Roxette	9 []

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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6. Similar varieties and differences from these varieties

Please use the following table and box for comments to provide information on how your candidate variety differs from the variety (or varieties) which, to the best of your knowledge, is (or are) most similar. This information may help the examination authority to conduct its examination of distinctness in a more efficient way.

Denomination(s) of variety(ies) similar to your candidate variety	Characteristic(s) in which your candidate variety differs from the similar variety(ies)	Describe the expression of the characteristic(s) for the similar variety(ies)	Describe the expression of the characteristic(s) for your candidate variety
<i>Example</i>	<i>Plant: diameter</i>	<i>medium</i>	<i>medium to large</i>
Comments:			

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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#7. Additional information which may help in the examination of the variety

7.1 In addition to the information provided in sections 5 and 6, are there any additional characteristics which may help to distinguish the variety?

Yes No

(If yes, please provide details)

7.2 Are there any special conditions for growing the variety or conducting the examination?

Yes No

(If yes, please provide details)

7.3 Other information

Type (see 5.3 and 8.1 in the Test Guidelines for Lettuce (document TG/13/11) for explanations):

Type	Example varieties	
Butterhead type	Clarion, Maikönig, Sartre	<input type="checkbox"/>
Novita type	Norvick	<input type="checkbox"/>
Iceberg type	Great Lakes 659, Roxette, Saladin, Vanguard 75	<input type="checkbox"/>
Batavia type	Aquarel, Curtis, Funnice, Felucca, Grand Rapids, Masaida, Visyon	<input type="checkbox"/>
Frisée d'Amérique type	Bijou, Blonde à couper améliorée	<input type="checkbox"/>
Lollo type	Lollo rossa, Revolution	<input type="checkbox"/>
Oakleaf type	Catalogna, Kipling, Murai, Salad Bowl	<input type="checkbox"/>
Multi-divided type	Curletta, Duplex, Jadigon, Rodagio	<input type="checkbox"/>
Frillice type	Frilett	<input type="checkbox"/>
Cos type	Actarus, Blonde maraîchère, Pinokkio	<input type="checkbox"/>
Gem type	Craquerelle du Midi, Sucrine, Xanadu	<input type="checkbox"/>
Stem type	Celtuice, Guasihong	<input type="checkbox"/>

Resistances:

(38) Resistance to *Bremia lactucae* (BI) isolate BI: 16
not tested 0 absent 1 present 9

(39) Resistance to *Bremia lactucae* (BI) isolate BI: 17
not tested 0 absent 1 present 9

(40) Resistance to *Bremia lactucae* (BI) isolate BI: 20
not tested 0 absent 1 present 9

(41) Resistance to *Bremia lactucae* (BI) isolate BI: 21
not tested 0 absent 1 present 9

(42) Resistance to *Bremia lactucae* (BI) isolate BI: 22
not tested 0 absent 1 present 9

(43) Resistance to *Bremia lactucae* (BI) isolate BI: 23
not tested 0 absent 1 present 9

- (44) Resistance to *Bremia lactucae* (Bl) isolate Bl: 24
not tested 0 [] absent 1 [] present 9 []
- (45) Resistance to *Bremia lactucae* (Bl) isolate Bl: 25
not tested 0 [] absent 1 [] present 9 []
- (46) Resistance to *Bremia lactucae* (Bl) isolate Bl: 26
not tested 0 [] absent 1 [] present 9 []
- (47) Resistance to *Bremia lactucae* (Bl) isolate Bl: 27
not tested 0 [] absent 1 [] present 9 []
- (48) Resistance to *Bremia lactucae* (Bl) isolate Bl: 29
not tested 0 [] absent 1 [] present 9 []
- (49) Resistance to *Bremia lactucae* (Bl) isolate Bl: 30
not tested 0 [] absent 1 [] present 9 []
- (50) Resistance to *Bremia lactucae* (Bl) isolate Bl: 31
not tested 0 [] absent 1 [] present 9 []
- (51) Resistance to *Lettuce mosaic virus* (LMV) pathotype II
not tested 0 [] absent 1 [] present 9 []
- (52) Resistance to *Nasonovia ribisnigri* (Nr) biotype Nr: 0
not tested 0 [] absent 1 [] present 9 []
- (53) Resistance to *Fusarium oxysporum* f. sp. *lactucae* (Fol) race 1
not tested 0 [] susceptible 1 [] moderately resistant 2 [] highly resistant 3 []

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:
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8. Authorization for release

(a) Does the variety require prior authorization for release under legislation concerning the protection of the environment, human and animal health?

Yes No

(b) Has such authorization been obtained?

Yes No

If the answer to (b) is yes, please attach a copy of the authorization.

9. Information on plant material to be examined or submitted for examination

9.1 The expression of a characteristic or several characteristics of a variety may be affected by factors, such as pests and disease, chemical treatment (e.g. growth retardants or pesticides), effects of tissue culture, different rootstocks, scions taken from different growth phases of a tree, etc.

9.2 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If the plant material has undergone such treatment, full details of the treatment must be given. In this respect, please indicate below, to the best of your knowledge, if the plant material to be examined has been subjected to:

(a) Microorganisms (e.g. virus, bacteria, phytoplasma)	Yes <input type="checkbox"/>	No <input type="checkbox"/>
(b) Chemical treatment (e.g. growth retardant, pesticide)	Yes <input type="checkbox"/>	No <input type="checkbox"/>
(c) Tissue culture	Yes <input type="checkbox"/>	No <input type="checkbox"/>
(d) Other factors	Yes <input type="checkbox"/>	No <input type="checkbox"/>

Please provide details for where you have indicated "yes".

.....

10. I hereby declare that, to the best of my knowledge, the information provided in this form is correct:

Applicant's name

Signature Date

[End of document]