

**Working Group on Biochemical and Molecular Techniques  
and DNA-Profiling in Particular**

**BMT/17/15 Add.**

**Seventeenth Session  
Montevideo, Uruguay, September 10 to 13, 2018**

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**ADDENDUM TO  
A DNA DATABASE FOR ROSE: DEVELOPMENT AND VALIDATION OF A SNP MARKER SET**

*Document prepared by experts from the Netherlands, the International Rose Breeders Association (IRBA) and the International Community of Breeders of Asexually Reproduced Ornamental and Fruit-Tree Varieties (CIOPORA)*


*Disclaimer: this document does not represent UPOV policies or guidance*

The Annex to this document contains a copy of a presentation on “A DNA database for Rose: Development and validation of a SNP marker set”, prepared by experts from the Netherlands, the International Rose Breeders Association (IRBA) and the International Community of Breeders of Asexually Reproduced Ornamental and Fruit-Tree Varieties (CIOPORA), which was made at the seventeenth session of the Working Group on Biochemical and Molecular Techniques and DNA-Profiling in Particular (BMT).

[Annex follows]

A DNA DATABASE FOR ROSE: DEVELOPMENT AND VALIDATION OF A SNP MARKER SET

Presentation prepared by experts from the Netherlands, the International Rose Breeders Association (IRBA) and the International Community of Breeders of Asexually Reproduced Ornamental and Fruit-Tree Varieties (CIOPORA)




BMT/17/15

# A DNA database for Rose

*Development and validation of a SNP marker set*

Hedwich Teunissen




## History and Timeline

2011:  
First CIOPORA / IRBA Meeting – idea of a DNA database was introduced  
Not mature at that time


2011-2016:  
CPVO project on Extraction and Storage of DNA samples of Rose Varieties

2014:  
Previous initiatives (project proposals) to construct DNA databases using SSR markers

2017:  
New initiative to construct DNA databases using SNP markers  
CPVO R&D proposal – Not mature


	<h2 data-bbox="581 199 1130 241">Title and scope of the project</h2>
	<p data-bbox="475 294 548 325">Title:</p> <p data-bbox="475 346 1214 420"><b>Development and validation of a SNP set for cut rose</b></p> <p data-bbox="475 493 1221 598">Ultimate goal: - construction of a database - implementation of DNA in DUS testing of cut rose (management of reference collection)</p> <p data-bbox="475 630 787 667">First step: <u>this project</u></p> <p data-bbox="475 682 1144 714">Dutch Board for Plant Varieties &amp; Naktuinbouw</p>

	<h2 data-bbox="682 966 1031 1008">Project Partners</h2>
	<ul data-bbox="487 1092 1079 1228" style="list-style-type: none"><li data-bbox="487 1092 722 1123">• Naktuinbouw</li><li data-bbox="487 1144 1079 1228">• Rose Breeders (IRBA / CIOPORA) (10 international breeding companies)</li></ul>




## Why Cut Rose?

- Model for ornamental crops with a complex polyploid genome.
- Intensively studied in scientific world (also previous work by Ben Vosman)
- Relevant data on molecular markers and DNA sequences in public domain
- Recently, two papers describing two reference genome sequences (Hibrand Saint-Oyant et al., 2018 and Raymond et al., 2018)




## Why Cut Rose?

- World wide breeding activities and trade. No restriction to regions or countries
- Number of existing varieties (common knowledge) is high
- Very important ornamental crop with many new applications each year
- There is a need to manage the variety collection to
  - Safely exclude comparing varieties from the trial
  - Reduce workload and save costs
  - Avoid an extra year of testing



## Challenges and Risks

- No full overview and availability of common knowledge:
  - ✓ Rapid development of new varieties
  - ✓ Global character of breeding
- Phytosanitary restriction of importing living reference material:
  - ✓ Risk for plant health
  - ✓ Very costly
- Stability of the variety description
  - ✓ Phenotype is influenced by environment
  - ✓ Risk on wrong comparing varieties based on photos and TQ
  - ✓ DUS trials will take more than 1 year



## Benefits and Applications

DNA profiles in databases overcome all risks

Future perspective

More reliable decision on Distinctness: objective comparison of genetic distance for each application with all varieties in the database

Fits with approved UPOV model 2 'combining phenotypic and molecular distances in the management of variety collections'

## Project Objectives and Tasks

- **Develop Cut Rose-specific SNP panel**
  - Identification of cut rose-specific SNP markers
  - Selection of SNP markers based on quality criteria and performance
  - Test and Optimization of the genotyping method, discard badly performing SNPs
  - Genotyping method validation
  - Fit for Purpose Validation of SNP marker set
  - Publication of the results

## Identification of Cut Rose specific SNPs


Strategies for the Identification of SNP  
(genetic diversity in training set)


€ Select SNPs from publically available genome sequences that represent genetic diversity

€€ Screen commercial available SNP arrays to obtain SNPs from our selection of varieties

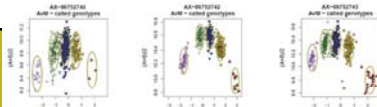
€€€ Re-sequence selection of varieties to obtain SNPs

68,983 SNPs





From tetraploid cut rose and garden rose varieties



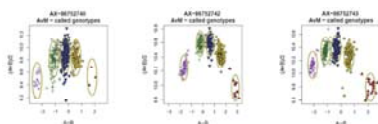
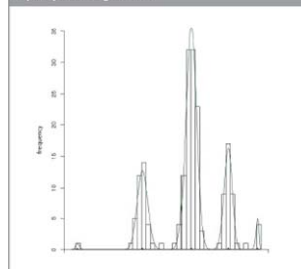


## Identification of Cut Rose specific SNPs

### Quality criteria for SNP selection


- ✓ Allele frequencies / dosage
- ✓ PIC value (highly discriminative)
- ✓ Single locus
- ✓ Equal distribution
- ✓ Whole genome covered
- ✓ Easy to score

Figure 1: The typical graphical output of fitetra is a histogram of the measured ratios of the two signals: (allele a)/(allele a + allele b), with the model that is fitted to the observed ratios superimposed as a green line.



## Identification of Cut Rose specific SNPs


- Define **training set** of varieties that is representative for broad genetic diversity. Focus on cut rose (70%), other types (30%). Species used as rootstock are excluded (not relevant)
- This set is used for the identification of SNPs
- Define **test set** of varieties that represent difficult samples in respect to DUS and controls (e.g. same variety different origins; mutants and/or derived varieties; seedlings and parents).
- This set is used for the validation



## Results of Identification

First selection of SNPs (~384)

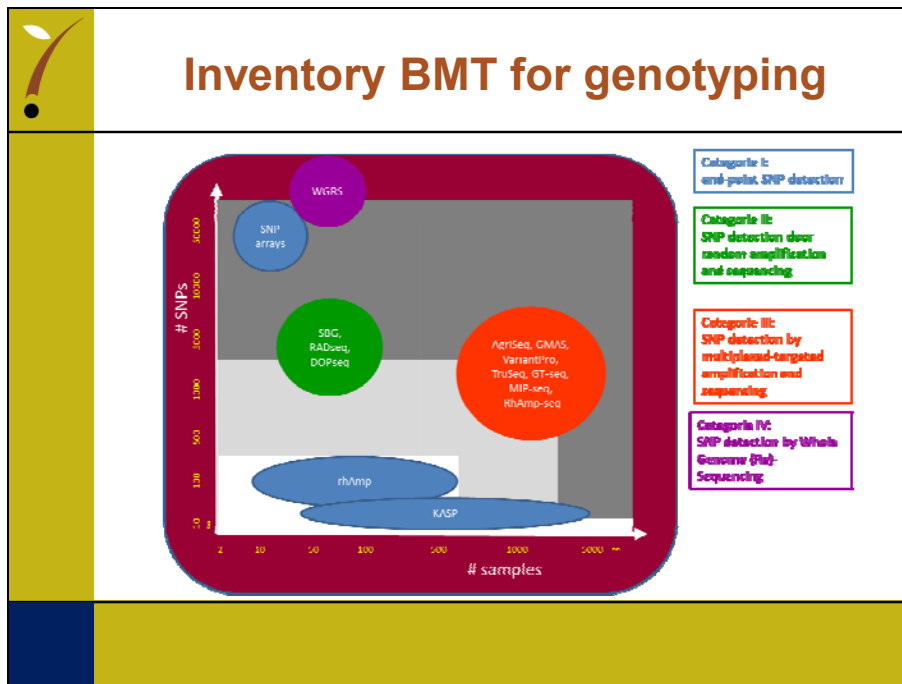
- On performance based on Quality Criteria
- On performance on Distinctness of varieties in training set



## Choice on SNP detection technology

- Inventory and benchmark of genotyping technologies in 2017
- Promising Genotyping technologies:
  - ✓ SNP Genotyping by Multiplexed Targeted Amplicon Sequencing Approach (e.g. GT-Seq, MIP-seq)
  - ✓ Single SNP assays - End-Point measurements (e.g. RhAmp, KASP)



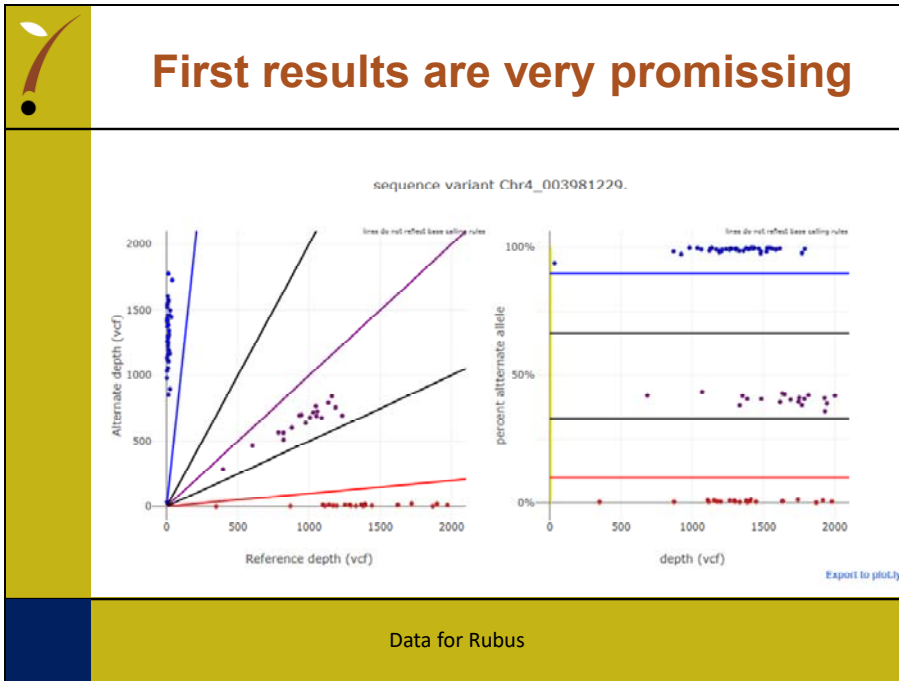


## SNP genotyping by multiplexed targeted amplicon sequencing


- GT seq**
  - Genotyping-in-thousands by sequencing
  - Cost effective SNP genotyping method based on multiplexed targeted amplicon sequencing
  - Combining multiple loci and multiple samples in 1 sequencing run (even samples of different crops and experiments)
  - Open source method

GT-SEQ: SNP GENOTYPING BY

**PCR1:** Targeted multiplex PCR adds Illumina sequencing primer sites to amplicons.  
**PCR2:** Targeted PCR adds unique barcode sequences and Illumina capture sites to targets.  
**Sequencing:** Mass parallelization, normalization and pool sample amplicons.  
**Illumina Sequencing:** Single and dual 150 base reads with dual 5 base index sequencing.  
**SNP Sequences into Individual Files:** 25 clusters identify genes and its sequence identifies each position.  
**Genotype samples:** Count occurrences of each allele at each locus and use allele calls to generate genotypes.




- ## Optimization of the method
- PCR components and reaction conditions
  - Primer-dimers (inefficient amplification)
  - PCR duplicates (inefficient amplification and hamper allele dosage determination)
  - Normalisation
  - Multiplex factor (# samples x # SNPs)
  - Sequence coverage needed for reliable dosage calling?
  - Bioinformatic pipelines



## Method validation

- After optimization the protocol is fixed
- Test how well the method performs:
  - Accuracy
  - Reproducibility
  - Repeatability
  - Robustness



## Fit for Purpose Validation

- Validation of selected SNPs using defined varieties from the **test set** (samples that should be distinct and samples that should not be distinct)
  - ✓ Genetically very similar varieties or lines
  - ✓ Parents and off-spring
  - ✓ Genetically close but morphologically distinct varieties
  - ✓ Series of mutants e.g.
- Blind tests with new sets of varieties, replica's and controls
- Select final SNPs set




## Publically available SNP set

- Publish the results in peer reviewed scientific paper. SNP set is publically available. Also for other DUS testing authorities to use.
- The choice on SNP detection method is open for every lab
- Validation between labs is required



## Memorandum of Understanding

- Breeders provide well defined plant material for both training set and test set
- Naktuinbouw performs the laboratory work
- Screening of the Axiom Array is outsourced to service provider in NL
- All partners have signed the MoU: legal and practical guidance during the project



## State of Affairs

- Training set of 164 varieties and 28 control samples obtained from 9 breeding companies
- DNA was extracted (is challenging when old leaf material was provided)
- DNA was send to service provider. Currently screening of Axiom Array takes place

***Quality in Horticulture***