



BMT/15/6

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INTERNATIONAL UNION FOR THE PROTECTION OF NEW VARIETIES OF PLANTS

Geneva

**WORKING GROUP ON BIOCHEMICAL AND MOLECULAR
TECHNIQUES AND DNA PROFILING IN PARTICULAR**

Fifteenth Session

Moscow, Russian Federation, May 24 to 27, 2016

VARIETY IDENTIFICATION OF BARLEY USING KASP GENOTYPING



Document prepared by an expert from the United Kingdom

Disclaimer: this document does not represent UPOV policies or guidance

The Annex to this document contains a copy of a presentation "Variety identification of barley using KASP genotyping" to be made at its fifteenth session of the Working Group on Biochemical and Molecular Techniques and DNA-Profiling in particular (BMT).



Alex Reid, Molecular Biologist, Science and Advice for Scottish Agriculture (SASA), the United Kingdom

[Annex follows]

Variety identification of barley using KASP genotyping

Alex Reid

SNP development

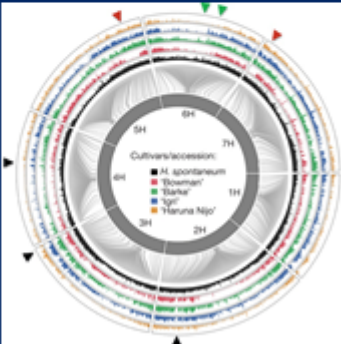
The barley genome sequencing project identified over a hundred thousand SNPs.

We have been working closely with James Hutton Institute in Dundee on the development of a set of SNPs to identify varieties.

JHI screen over 1200 SNPs against 940 varieties.

In conjunction with BLOSS we have narrowed this down to a set of 33 SNPs.

DNA extracted from bulked samples.



KASP genotyping assay

Developed by LGC.



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Uses a unique type of Kompetitive Allele-Specific PCR which can yield highly specific bi-allelic discrimination of SNPs and InDels.

There are a couple of very nice YouTube videos that explains how it works

<https://www.youtube.com/watch?v=Uq9HhmzOqUQ>

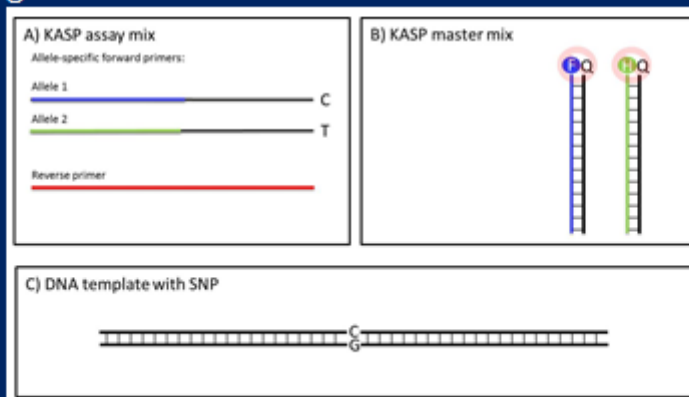
<https://www.youtube.com/watch?v=GJbM7UbE7ZI>

KASP genotyping requires 3 components

A) Assay mix containing 2 allele-specific primers (one for each SNP). Each has a unique unlabelled tail at the 5' end. One common (reverse) primer.

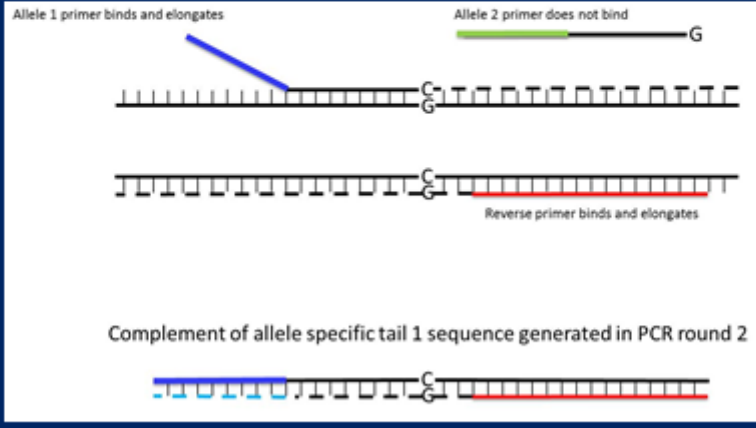
B) FAM and HEX FRET cassettes.

C) Target DNA.



KASP how it works

As PCR continues products are produced which contain one of the SNP alleles with a complimentary tail to the specific FRET cassette.



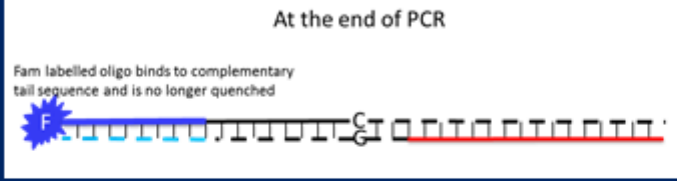
The diagram illustrates the PCR process in three stages. In the first stage, 'Allele 1 primer binds and elongates' (indicated by a blue arrow) and 'Allele 2 primer does not bind' (indicated by a green line). The DNA template has a C-G SNP site. In the second stage, 'Reverse primer binds and elongates' (indicated by a red arrow). The final stage shows the 'Complement of allele specific tail 1 sequence generated in PCR round 2', which is a DNA strand with a blue tail and a red tail.

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Inspiration for the future

KASP how it works

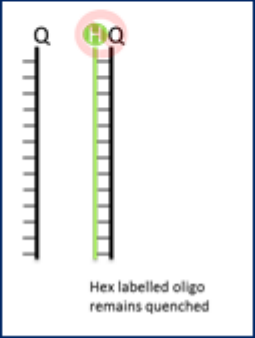
At the end of PCR

Fam labelled oligo binds to complementary tail sequence and is no longer quenched



The diagram shows a FAM-labelled oligo (blue star) binding to the blue tail of the PCR product. The FAM label is now free from the quencher.

The binding of the FRET to its complement frees the fluorophore from its quencher.
The other SNP FRET remains quenched.
End point fluorescence reading is taken.



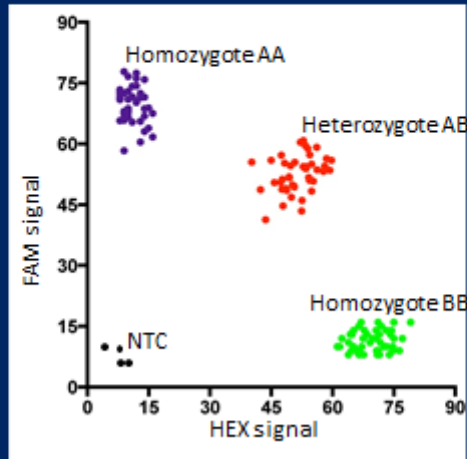
The diagram shows a HEX-labelled oligo (green star) bound to a red tail, which remains quenched.

Hex labelled oligo remains quenched

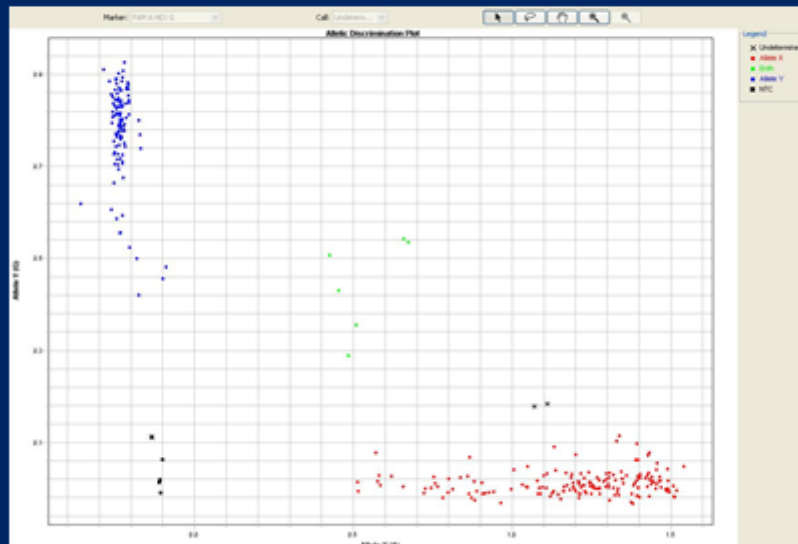
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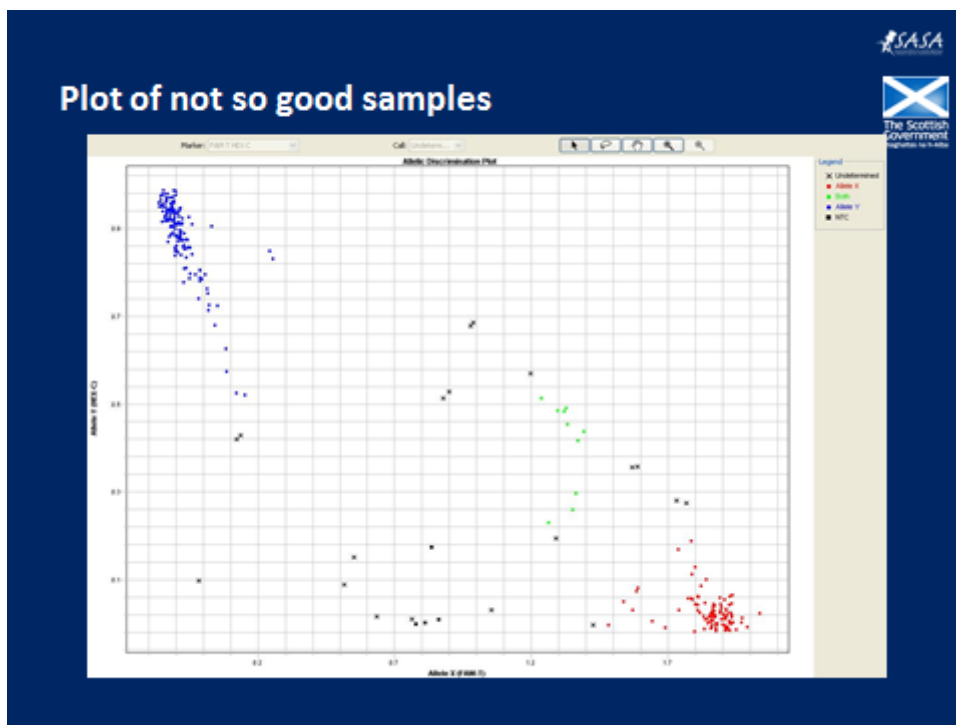
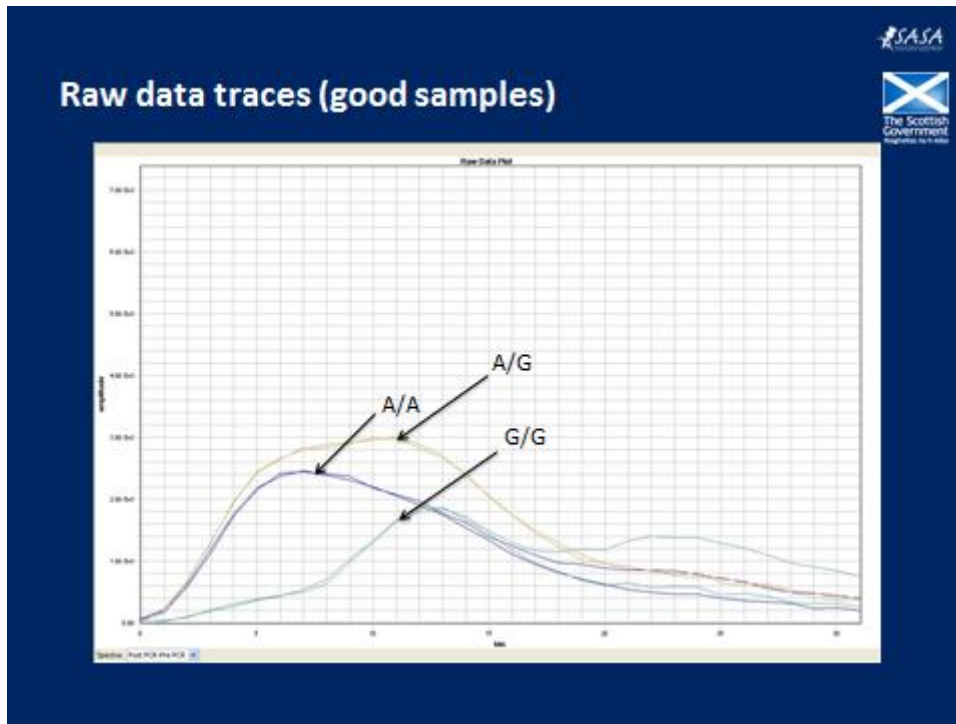
KASP results

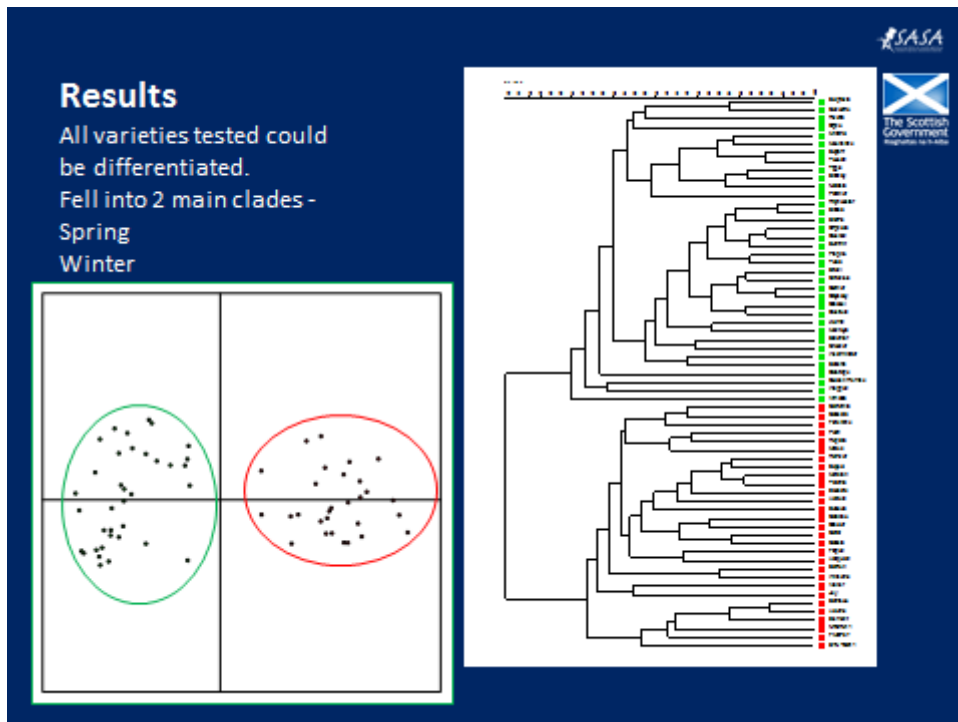
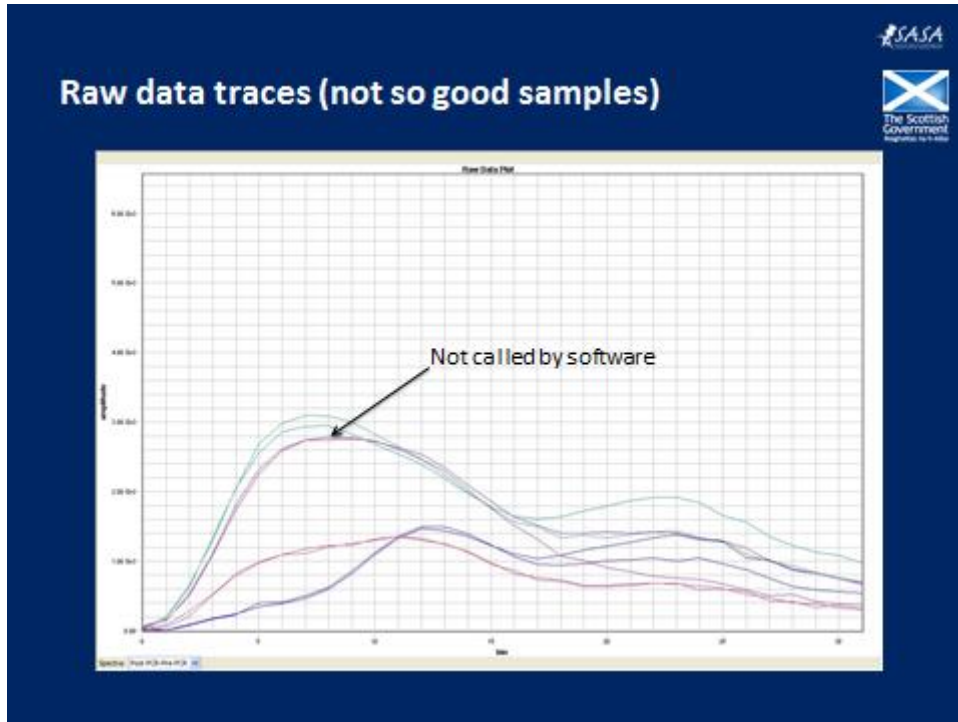
In an ideal world results look like this.



Not always so clear cut (good run)







Not all plane sailing

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The occasional variety yielded odd results.
However, all blind test samples were identified correctly.

Pros and Cons of KASP

Pros

Very cheap (ca. £9/sample for all 33 SNPs). This is for 188 samples in duplicate using 384 well plates.

Can be automated easily (both DNA extraction and PCR set-up).

Automated scoring.

Easy to scaled up to 384 and 1536 well plates.

Cons

If you only want to run one sample with all 33 SNPs this rises to £51.

Each SNP requires several controls so not really feasible to run one sample with 33 SNPs

Not always easy to score (esp. heterozygotes).

There are limits to rogue detection.

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Summary

We have a working KASP genotyping method for barley variety identification.
Using 33 SNP assays.
Initial test of 66 varieties.
Blind test successful.
Two main clades spring and winter varieties.
Largely automated and therefore fairly high throughput.
Future work to expand the number of varieties tested.



Acknowledgements

SASA

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