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**USE OF A MOLECULAR MARKER-BASED SYSTEM FOR IDENTIFICATION OF  
VARIETIES WITHIN THE GENUS *EUCALYPTUS***


*Document prepared by experts from Brazil*




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**USE OF A MOLECULAR MARKER-BASED SYSTEM  
FOR IDENTIFICATION OF VARIETIES WITHIN THE  
GENUS *Eucalyptus***

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## INTRODUCTION

The genus *Eucalyptus* comprises the most commonly used species for production of **short fiber cellulose** in Brazil.

- Brazil has the largest area with *Eucalyptus* in the world - over 3 million ha
- 6,3 million tons of **cellulose**/year

## INTRODUCTION

- **Alogamous** plant **but** vegetatively propagated (rooted cuttings and micropropagation)
- Clones selected within families
- Sister lines** – low morphological variability
- **Long cycle**

## Characteristics of molecular descriptors - *Eucalyptus*

**Twenty-five** loci markers using **microsatellite sequences** are recommended, and might be considered as **complementary** descriptors for the identification of clones, hybrids and varieties of *Eucalyptus*.

Among these 25, **six** are considered **mandatory** in order to allow the standardization and comparison of genetic profiles generated by different laboratories and to different clones.

## SSR Markers - *Eucalyptus*

Loci	Allele size (base pairs)	Sequence 5'-3' of direct primer	Sequence 5'-3' of reverse primer	Linkage Group
Embra01	100-145	gatagaacttctattgatcg	gtaggattgatgtctgcaa	8
Embra02	103-148	cgfgacaccaggacattac	acaaatgcaaattcaaatga	11
Embra05	78-142	atgctggtccaaactaagatt	tgagcctaaaagcccaac	5
Embra06	120-170	agagaattgctctcatgga	gaaaagtctgcaaatctctge	1
Embra10	110-152	gtaagacatagtgagacattcc	agacagtacgttctctagetc	10
Embra11	123-165	gcttagaatttgcctaaacc	gtaaaatccatgggcaag	1
Embra12	104-162	aggatttgggggcaagt	gttccccatttcatgtcc	1
Embra15	90-125	tttgtrggatgaggactt	caacatgttctccgaaaag	8
Embra16	110-165	caacgtcccccctttctc	atgttagcccaaccag	1
Embra17	120-170	aggatactcgtgagagaagc	gtagatctgtctcatgttg	9
Embra19	55-145	gacgggtgattctctgatt	gtggtgctctctctctct	4
Embra23	118-145	ggttggttcatctttccatg	agcgaaggcaatgtgtt	10
Embra26	112-200	cccacaacaaagggaag	agaggtgtctgattcaatc	11
Embra27	100-170	ataaccacacaaatctgca	tatagctcgaacgtcaac	2
Embra28	180-300	caagacatgatttctgagt	actcttgatgtgacgagaca	6
Embra34	100-160	tcaaaacccctctctcat	aataaacattttctgaacaga	3
Embra37	115-165	caactctccaaactacacaa	ctctctctcttccaccatc	5
Embra42	115-170	gagtaaaaattggtttgagtg	ccctcttttattttgtctt	7
Embra44	205-225	gggggtttgtctgcttag	caaaagagttcagctgtg	4
Embra46	90-130	gaagtcacatctgtgattgc	accattattctttgtgagc	7
Embra49	125-195	attattggttcattatgaaaacc	agatagagattgagtgagacc	3
Embra51	95-200	gatgcattccttttttcc	cattctctgcatctggac	6
Embra58	140-245	caccaactgtgactatgaggat	ttgcttagggtagaacact	9
Embra63	175-230	catctggagatcgaggaa	gagagaaggatcatgccca	2
Embra72	118-170	ctggtcaacgtccgaaag	atgctgcagagggcataa	10

## Reliability

The molecular characterization of these loci have been already published in the **literature** and were validated in a ring test in **several laboratories** around the world:

Scotland - 1;

Brazil -3;

South Africa - 1;

Portugal - 1;

Australia - 2;

Argentina - 1.

## Linkage Groups

At least two molecular markers are listed for each of the eleven linkage groups, corresponding to the eleven chromosomes of *Eucalyptus*.

## Methods

- ▣ 1. Extraction and quantification of DNA:
- ▣ Genomic DNA from plant tissues (leaves, cambium, flowers, etc.).
- ▣ Protocol described by Ferreira & Grattapaglia (1998).
- ▣ The DNA must be quantified by electrophoresis in 0.8% agarose gel followed by ethidium bromide staining.



## Methods

### 2. PCR (Polymerase chain reaction):

#### Reagents:

- 2 to 50 ng of genomic DNA;
- 1.5 mM of Mg<sup>++</sup>;
- 0.25 µM of direct and reverse primers;
- 200 µM of each nucleotide;
- 0.2 mg/ml BSA;
- 1 x buffer PCR with 50 mM KCL;
- 10 mM TRIS-HCL pH 9.0;
- 0.1% Triton X-100;
- 1 polymerase unit of *Taq* DNA in a total volume of 15 µl.

#### PCR Program:

1. Initial denaturation at 95° C for 4 minutes;
2. 30 cycles of denaturation at 95° C for 1 minute;
3. Annealing at 52° C for 1 minute;
4. Extension at 65° C for 1 minute;
5. Final extension step at 65° C for 10 minutes.

## Methods

### 3. Polymorphism detection and genotype determination:

- automatic DNA sequencer is recommended;
- the primers for microsatellite loci must be marked with fluorochromes (blue (FAM); green (HEX); or yellow (NED)) and a specific spectrum filter, according with technology widely used in individual identification in human beings, animals and cultivated plants.
- each locus can be analyzed **individually**, or in "**multiplex**" combinations for simultaneous analyses of several loci.
- an internal standard marked with a fluorescent **TAMRA** or a red color **ROX** must be used for definition of fragment sizes.

## Methods

- ❑ 4. **Interpretation of results**
- ❑ - for each of the analyzed descriptor loci, the observed genotype should be identified and registered.
- ❑ - the alleles will be visualized as peaks in the electropherogram and will be identified by their size in base pairs.
- ❑ - genotypes should be described with the alleles identified in **number of base pairs**.
- ❑ - the analysis should include, as **control check**, the DNA of a well characterized *Eucalyptus* clone, to be identified by the laboratory, to be used for comparison of allele size in base pairs among laboratories or between different experiments within the same laboratory.

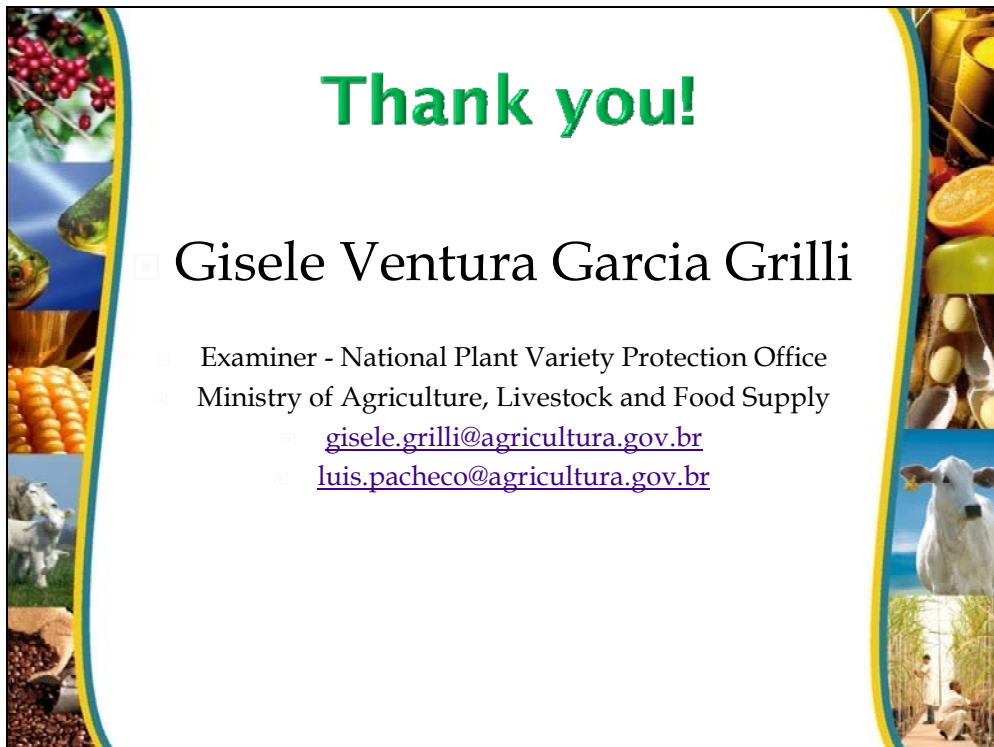
## Database Model

		Genotypes in pairs of bases							
Loci EMBRA		2		28		11		15	
Sample	Number	Allele 1	Allele 2	Allele 1	Allele 2	Allele 1	Allele 2	Allele 1	Allele 2
DF-EG112	1	108.42	113.12	190.52	209.15	133.73	133.73	117.92	142.22
DF-EG113	2	108.30	113.12	190.39	209.13	133.68	133.68	117.92	142.32
DF-EG114	3	108.30	<b>124.54</b>	190.53	<b>221.26</b>	<b>117.73</b>	<b>124.59</b>	<b>126.07</b>	<b>138.22</b>
DF-EG115	4	108.30	113.00	190.41	209.13	133.69	133.69	117.69	142.11
Control Check	C	124.43	145.13	197.06	221.31	117.40	119.30	125.92	136.16



## Researchers involved (Brazil)

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## Thank you!

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