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INTERNATIONAL UNION FOR THE PROTECTION OF NEW VARIETIES OF PLANTS

Geneva

DRAFT

TRITICALE

UPOV Code(s):

TRITL

×Triticosecale Witt.

GUIDELINES

FOR THE CONDUCT OF TESTS

FOR DISTINCTNESS, UNIFORMITY AND STABILITY

prepared by experts from Australia to be considered by the Technical Working Party for Agricultural Crops at its forty-seventh session, to be held in Naivasha, Kenya, from 2018-05-21 to 2018-05-25

Disclaimer: this document does not represent UPOV policies or guidance

Alternative names:* Botanical name	English	French	German	Spanish
× <i>Triticosecale</i> Witt.	Triticale	Triticale	Triticale	Triticale

The purpose of these guidelines ("Test Guidelines") is to elaborate the principles contained in the General Introduction (document TG/1/3), and its associated TGP documents, into detailed practical guidance for the harmonized examination of distinctness, uniformity and stability (DUS) and, in particular, to identify appropriate characteristics for the examination of DUS and production of harmonized variety descriptions.

ASSOCIATED DOCUMENTS

These Test Guidelines should be read in conjunction with the General Introduction and its associated TGP documents.

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1. <u>Subject of these Test Guidelines</u>

These Test Guidelines apply to all varieties of ×Triticosecale Witt..

2. <u>Material Required</u>

- 2.1 The competent authorities decide on the quantity and quality of the plant material required for testing the variety and when and where it is to be delivered. Applicants submitting material from a State other than that in which the testing takes place must ensure that all customs formalities and phytosanitary requirements are complied with.
- 2.2 The material is to be supplied in the form of seeds and ears (if requested).
- 2.3 The minimum quantity of plant material, to be supplied by the applicant, should be:

Seeds: 3 kg Ears (if requested): 200

The seed should meet the minimum requirements for germination, species and analytical purity, health and moisture content, specified by the competent authority. In cases where the seed is to be stored, the germination capacity should be as high as possible and should, be stated by the applicant.

The ear should be well developed and should contain a sufficient number of viable seeds to establish a satisfactory row of plants for observation.

- 2.4 The plant material supplied should be visibly healthy, not lacking in vigor, nor affected by any important pest or disease.
- 2.5 The plant material should not have undergone any treatment which would affect the expression of the characteristics of the variety, unless the competent authorities allow or request such treatment. If it has been treated, full details of the treatment must be given.

3. <u>Method of Examination</u>

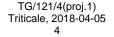
3.1 Number of Growing Cycles

The minimum duration of tests should normally be two independent growing cycles.

3.2 Testing Place

Tests are normally conducted at one place. In the case of tests conducted at more than one place, guidance is provided in TGP/9 "Examining Distinctness".

- 3.3 Conditions for Conducting the Examination
- 3.3.1 The tests should be carried out under conditions ensuring satisfactory growth for the expression of the relevant characteristics of the variety and for the conduct of the examination.
- 3.3.2 The optimum stage of development for the assessment of each characteristic is indicated by a number in the Table of Characteristics. The stages of development denoted by each number are described in Chapter 8.
- 3.4 Test Design
- 3.4.1 Each test should be designed to result in a total of at least 2000 plants, which should be divided between at least 2 replicates.
- 3.4.2 The design of the tests should be such that plants or parts of plants may be removed for measurement or counting without prejudice to the observations which must be made up to the end of the growing cycle.
- 3.4.3 If tests on ear rows are conducted, at least 100 ear rows should be observed. The assessment of the characteristic "Seasonal type" should be carried out on at least 300 plants.



3.5 Additional Tests

Additional tests, for examining relevant characteristics, may be established.

4. Assessment of Distinctness, Uniformity and Stability

4.1 Distinctness

4.1.1 General Recommendations

It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding distinctness. However, the following points are provided for elaboration or emphasis in these Test Guidelines.

To assess distinctness of hybrids, the parent lines and the formula may be used according to the following recommendations:

(i) description of parent lines according to the Test Guidelines;

(ii) check of the originality of the parent lines in comparison with the variety collection, based on the characteristics in Chapter 7, in order to identify similar parent lines;

(iii) check of the originality of the hybrid formula in relation to the hybrids in the variety collection, taking into account the most similar lines; and

(iv) assessment of the distinctness at the hybrid level for varieties with a similar formula.

Further guidance is provided in documents TGP/9 "Examining Distinctness" and TGP/8 "Trial Design and Techniques Used in the Examination of Distinctness, Uniformity and Stability".

4.1.2 Consistent Differences

The differences observed between varieties may be so clear that more than one growing cycle is not necessary. In addition, in some circumstances, the influence of the environment is not such that more than a single growing cycle is required to provide assurance that the differences observed between varieties are sufficiently consistent. One means of ensuring that a difference in a characteristic, observed in a growing trial, is sufficiently consistent is to examine the characteristic in at least two independent growing cycles.

4.1.3 Clear Differences

Determining whether a difference between two varieties is clear depends on many factors, and should consider, in particular, the type of expression of the characteristic being examined, i.e. whether it is expressed in a qualitative, quantitative, or pseudo-qualitative manner. Therefore, it is important that users of these Test Guidelines are familiar with the recommendations contained in the General Introduction prior to making decisions regarding distinctness.

4.1.4 Number of Plants or Parts of Plants to be Examined

Unless otherwise indicated, for the purposes of distinctness, all observations on single plants should be made on 10 plants or parts of plants taken from each of 10 plants and any other observations made on all plants in the test, disregarding any off-type plants.

In the case of observations of parts taken from single plants, the number of parts to be taken from each of the plants should be 1.

4.1.5 Method of Observation

The recommended method of observing the characteristic for the purposes of distinctness is indicated by the following key in the Table of Characteristics (see document TGP/9 "Examining Distinctness", Section 4 "Observation of characteristics"):

MG: single measurement of a group of plants or parts of plants MS: measurement of a number of individual plants or parts of plants VG: visual assessment by a single observation of a group of plants or parts of plants VS: visual assessment by observation of individual plants or parts of plants

Type of observation: visual (V) or measurement (M)

"Visual" observation (V) is an observation made on the basis of the expert's judgment. For the purposes of this document, "visual" observation refers to the sensory observations of the experts and, therefore, also includes smell, taste and touch. Visual observation includes observations where the expert uses reference points (e.g. diagrams, example varieties, side-by-side comparison) or non-linear charts (e.g. color charts). Measurement (M) is an objective observation against a calibrated, linear scale e.g. using a ruler, weighing scales, colorimeter, dates, counts, etc.

Type of record: for a group of plants (G) or for single, individual plants (S)

For the purposes of distinctness, observations may be recorded as a single record for a group of plants or parts of plants (G), or may be recorded as records for a number of single, individual plants or parts of plants (S). In most cases, "G" provides a single record per variety and it is not possible or necessary to apply statistical methods in a plant-by-plant analysis for the assessment of distinctness.

In cases where more than one method of observing the characteristic is indicated in the Table of Characteristics (e.g. VG/MG), guidance on selecting an appropriate method is provided in document TGP/9, Section 4.2.

- 4.2 Uniformity
- 4.2.1 It is of particular importance for users of these Test Guidelines to consult the General Introduction prior to making decisions regarding uniformity. However, the following points are provided for elaboration or emphasis in these Test Guidelines:
- 4.2.2 These Test Guidelines have been developed for the examination of self-pollinated varieties. For varieties with other types of propagation, the recommendations in the General Introduction and document TGP/13 "Guidance for new types and species" Section 4.5 "Testing Uniformity" should be followed.
- 4.2.3 The assessment of uniformity for hybrid varieties depends on the type of hybrid and should be according to the recommendations for hybrid varieties in the General Introduction.
- 4.2.4 Where the assessment of a hybrid variety involves the parent lines, the uniformity of the hybrid variety should, in addition to an examination of the hybrid variety itself, also be assessed by examination of the uniformity of its parent lines.
- 4.2.5 The recommended sample size for the assessment of uniformity is indicated by the following key in the table of characteristics:
 - A sample size of 100 plants/parts of plants
 - B sample size of 2000 plants or parts of plants
- 4.2.6 For the assessment of uniformity of self-pollinated varieties, a population standard of 0.6% and an acceptance probability of at least 95% should be applied. In the case of a sample size of 2000 plants, 18 off-types are allowed.

4.2.7 For the assessment of uniformity in a sample of 100 ear-rows, plants or parts of plants, a population standard of 1% and an acceptance probability of at least 95% should be applied. In the case of a sample size of 100 ear-rows, plants or parts of plants, 3 off-types are allowed. An ear-row is considered to be an off-type ear-row if there is more than 1 off-type plant within that ear-row.

For "A" characteristics, with the exception of characteristic 2 and 3, the assessment of uniformity can be done in 2 steps. In a first step, 20 plants are observed. If no off-types are observed, the variety is considered to be uniform. If more than 3 off-types are observed, the variety is considered not to be uniform. If 1 to 3 off-types are observed, an additional sample of 80 plants or parts of plants must be observed.

For the assessment of uniformity of hybrid varieties, a population standard of 10% and an acceptance probability of at least 95% should be applied. In case of characteristics indicated by B, the sample size for the assessment of uniformity may be reduced to 200 plants. In case of a sample size of 200 plants, 27 off-types are allowed. In case of a sample size of 100 ear-rows, plants or parts of plants, 15 off-types are allowed.

- 4.3 Stability
- 4.3.1 In practice, it is not usual to perform tests of stability that produce results as certain as those of the testing of distinctness and uniformity. However, experience has demonstrated that, for many types of variety, when a variety has been shown to be uniform, it can also be considered to be stable.
- 4.3.2 Where appropriate, or in cases of doubt, stability may be further examined by testing a new seed stock to ensure that it exhibits the same characteristics as those shown by the initial material supplied.
- 4.3.3 Where appropriate, or in cases of doubt, the stability of a hybrid variety may, in addition to an examination of the hybrid variety itself, also be assessed by examination of the uniformity and stability of its parent lines.

5. <u>Grouping of Varieties and Organization of the Growing Trial</u>

- 5.1 The selection of varieties of common knowledge to be grown in the trial with the candidate varieties and the way in which these varieties are divided into groups to facilitate the assessment of distinctness are aided by the use of grouping characteristics.
- 5.2 Grouping characteristics are those in which the documented states of expression, even where produced at different locations, can be used, either individually or in combination with other such characteristics: (a) to select varieties of common knowledge that can be excluded from the growing trial used for examination of distinctness; and (b) to organize the growing trial so that similar varieties are grouped together.
- 5.3 The following have been agreed as useful grouping characteristics:
 - (a) Time of ear emergence (characteristic 5)
 - (b) Lower glume: hairiness on external surface (characteristic 16)
 - (c) Ear: color (characteristic 18)
 - (d) Seasonal type (characteristic 25)
- 5.4 Guidance for the use of grouping characteristics, in the process of examining distinctness, is provided through the General Introduction and document TGP/9 "Examining Distinctness".

6. Introduction to the Table of Characteristics

6.1 Categories of Characteristics

6.1.1 Standard Test Guidelines Characteristics

Standard Test Guidelines characteristics are those which are approved by UPOV for examination of DUS and from which members of the Union can select those suitable for their particular circumstances.

6.1.2 Asterisked Characteristics

Asterisked characteristics (denoted by *) are those included in the Test Guidelines which are important for the international harmonization of variety descriptions and should always be examined for DUS and included in the variety description by all members of the Union, except when the state of expression of a preceding characteristic or regional environmental conditions render this inappropriate.

- 6.2 States of Expression and Corresponding Notes
- 6.2.1 States of expression are given for each characteristic to define the characteristic and to harmonize descriptions. Each state of expression is allocated a corresponding numerical note for ease of recording of data and for the production and exchange of the description.
- 6.2.2 In the case of qualitative and pseudo-qualitative characteristics (see Chapter 6.3), all relevant states of expression are presented in the characteristic. However, in the case of quantitative characteristics with 5 or more states, an abbreviated scale may be used to minimize the size of the Table of Characteristics. For example, in the case of a quantitative characteristic with 9 states, the presentation of states of expression in the Test Guidelines may be abbreviated as follows:

State	Note
small	3
medium	5
large	7

However, it should be noted that all of the following 9 states of expression exist to describe varieties and should be used as appropriate:

State	Note
very small	1
very small to small	2
small	3
small to medium	4
medium	5
medium to large	6
large	7
large to very large	8
very large	9

- 6.2.3 Further explanation of the presentation of states of expression and notes is provided in document TGP/7 "Development of Test Guidelines".
- 6.3 Types of Expression

An explanation of the types of expression of characteristics (qualitative, quantitative and pseudoqualitative) is provided in the General Introduction.

6.4 Example Varieties

Where appropriate, example varieties are provided to clarify the states of expression of each characteristic.

Legend 6.5

	English	1	françai	S	deutsch español		Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
1 2	3	4	5	6	7			
	Name of characteristics in English		Nom o caract frança	ère en	Name des Merkmals auf Deutsch	Nombre del carácter en español		
	states expres		types of	d'expression	Ausprägungsstufen	tipos de expresión		

1 Characteristic number

2	(*)	Asterisked characteristic	- see Chapter 6.1.2
3	Type of expression QL QN PQ	Qualitative characteristic Quantitative characteristic Pseudo-qualitative characteristic	 see Chapter 6.3 see Chapter 6.3 see Chapter 6.3
4	Method of observation (and type MG, MS, VG, VS	e of plot, if applicable)	- see Chapter 4.1.5

- See Explanations on the Table of Characteristics in Chapter 8.1 5 (+)
- Not applicable 6
- 7 Growth stage key See Explanations on the Table of Characteristics in Chapter 8

 - A sample size of 100 plants/parts of plantsB sample size of 2000 plants or parts of plants

7. <u>Table of Characteristics/Tableau des caractères/Merkmalstabelle/Tabla de caracteres</u>

		English	fra	nçais	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
1.	QN	VG A	(+)		9-11			
	Colec antho colora	ocyanin						
	abser	nt or very weak					Coral Sea	1
	weak						Yowie	3
	mediu	ım					Tickit	5
	strong]						7
	very s	strong						9
2.	QN	VG B	(+)		25-29	·		
	Plant	growth habit						
	erect						Prime 322	1
	semi-	erect					Crackerjack	3
	interm	nediate					Chopper	5
	semi-	prostrate					Forerunner	7
	prostr	prostrate					Tobruk	9
3.	QN	VG B	(+)		47-51			
	Plant plant flag le	: frequency of s with recurved eaves						
	abser	nt or very low					Tuckerbox	1
	low						Crackerjack	3
	mediu	ım					Austute	5
	high						Forerunner	7
	very h	nigh					Madonna	9
4.	QN	VG A	(+)		47-51			
	Flag I colora	eaf: anthocyanin ation of auricles						
	abser	nt or weak					Austute	1
	weak						Hawkeye	3
	mediu	ım					Coral Sea	5
	strong)					Heritage Zephyr	7
	Very s	strong	[Crackerjack 2	9

		English	français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
5. (*)	QN	MG B	(+)				
	Time	of ear emergence					
	very e	arly				Chopper	1
	early					Prime 322	3
	mediu	ım				Coral Sea	5
	late					Crackerjack	7
	very la	ate				Pacific Falcon	9
6.	QN	VG B		55-65	-		
	Flag I sheat	eaf: glaucosity of h					
	absen	t or very weak				Tobruk	1
	weak					Endeavour	3
	mediu	ım				Forerunner	5
	strong)				Tickit	7
	very s	trong				Heritage Zephyr	9
7.	QN	VG B		65			
	Anther: anthocyanin coloration						
	absen	it or very weak				Tobruk	1
	weak						2
	strong)				Maiden	3
8. (*)	QN	VG A		80-92			
	Awn: colora	anthocyanin ation					
	absen	t or very weak				Crackerjack	1
	weak					Fusion	2
	mediu	ım				Yowie	3
	strong)					4
	very strong						5
9.	QN	MS A		60-69			
	Flag I blade	eaf: length of					
	short					Crackerjack	3
	mediu	ım				Chopper	5
	long					Endeavour	7

		English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
10.	QN	MS A			60-69			
<u>.</u>	Flag I blade	eaf: width of		•				
	narro	N					Tobruk	3
	mediu	ım					Yowie	5
	broad						Chopper	7
11. (*)	QN	VG B	(+)		60-69			
	Stem: hairin	density of less of neck		:				
		absent of very weak					Maiden	1
	weak						Tuckerbox	3
	mediu						Fusion	5
	strong						Austute	7
	very s	trong					Coral Sea	9
12.	QN	VG B			60-69			
	Ear: glaucosity							
	abser	absent or very weak					Tobruk	1
	weak						Coral Sea	3
	mediu	ım					Hawkeye	5
	strong)					Tuckerbox	7
	very s						Chopper	9
13.	QN	VG A	(+)		90-92			
	Straw sectio	r: pith in cross on						
	thin						Chopper	1
	mediu	ım					Kosciuszko	3
	thick							5
14. (*)	QN	VG A	(+)		80-92			
	Lowe of firs	r glume: length st beak						
	very s	hort						1
	short						Chopper	3
	mediu	ım					Tobruk	5
	long						Fusion	7
	very lo	ong					Treat	9

		English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
15.	QN	VG A	(+)		80-92			
	Lowe secor	r glume: size of Id beak						
	absen	t or very small					Treat	1
	small						Forerunner	3
	mediu	m						5
	large						Crackerjack 2	7
	very la	arge						9
16. (*)	QL	VG A			80-92	·	·	
	Lowe hairin surfac	r glume: ess on external ce						
	absen	t					Chopper	1
	prese	nt					Fusion	9
17. (*) I	QN	MG B	(+)		75-92		•	
	Plant: length			·				
	very s	hort						1
	short						Chopper	3
	mediu	m					Endeavour	5
	long						Forerunner	7
	very lo	ong						9
18. (*)	PQ	VG B	(+)		90-92	•	•	
i	Ear: c	olor		:				
	white						Austute	1
	slightly	y colored					Forerunner	2
	strong	ly colored						3
19.	QN	MS A/VG A	(+)		90-92	•	•	
·	Ear: d	lensity		•				
	very la	ax						1
	lax						Treat	3
	mediu	m					Coral Sea	5
	dense		1			+	Forerunner	7
	very d	ense	+				Tobruk	9

		English		français	deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
20. (*)	QN	VG A	(+)		80-92			1
	Ear: c awns	distribution of		1				
	tip aw	ned						1
	half a	awned					Coral Sea	2
	fully a	wned					Austute	3
21.	QN	VG B			90-92	1		
	Ear: v view	width in profile		ŝ				
	very r	narrow						1
	narro	W					Heritage Zephyr	3
	mediu						Austute	5
	broad						Hillary	7
	very b	broad						9
22. (*)	QN	MS A/VG A	(+)		80-92			
	Ear: I awns	ength of scurs or						
	very s	short					Forerunner	1
	short						Fusion	3
	mediu						Tobruk	5
	long						Yowie	7
	very le	ong					Maiden	9
23. (*)	QN	MS A/VG B	(+)		90-92	•		
	Ear: length							
	very s	short						1
	short						Crackerjack	3
	mediu	JM					Yowie	5
	long						Tuckerbox	7
	very le	ong	Ι					9

		English	français	s deutsch	español	Example Varieties Exemples Beispielssorten Variedades ejemplo	Note/ Nota
24.	QN	VG A	(+)	92			
	Grain pheno	: coloration with					
	absen	t or very light				Coral Sea	1
	light					Tobruk	3
	mediu	m				Tuckerbox	5
	dark					Credit	7
	very d	ark				Hawkeye	9
25. (*)	PQ	VG A	(+)		-		
	Seasonal type						
	winter	type				Coral Sea	1
	alterna	ative type				Breakwell	2
	spring	ı type				Austute	3

8.1 Explanations for individual characteristics

Ad. 1: Coleoptile: anthocyanin coloration

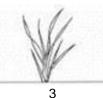
Method for the Determination of Anthocyanin Coloration Number of seeds per test: 100 seeds Preparation of seeds: Set up non-dormant seeds on moistened filter paper covered with a Petri dish lid during germination Place: Laboratory or greenhouse Light: After the coleoptiles have reached a length of about 1 cm in the dark, they are placed in artificial light (daylight equivalent) at 13000 to 15000 lux continuously for 3-4 days Temperature: 15 to 20°C Time of recording: Coleoptiles fully developed (about 1 week) at stage 09-11 Note: At least two example varieties should be included as a control Any alternative method may be used if it gives the same results

Ad. 2: Plant: growth habit

The growth habit should be assessed visually from the attitude of the leaves and tillers at tillering stage (growth stages 25-29). The angle formed by the outer leaves and the tillers with an imaginary middle axis should be used.



erect



semi-erect

5 intermediate

7

semi-prostrate

9 prostrate

Ad. 3: Plant: frequency of plants with recurved flag leaves

1 (absent or very low): all or almost all flag leaves are rectilinear

- 3 (low): about 1/4 of the plants with recurved flag leaves
- 5 (medium): about 1/2 of the plants with recurved flag leaves
- 7 (high): about 3/4 of the plants with recurved flag leaves

9 (very high): almost all or all flag leaves are recurved

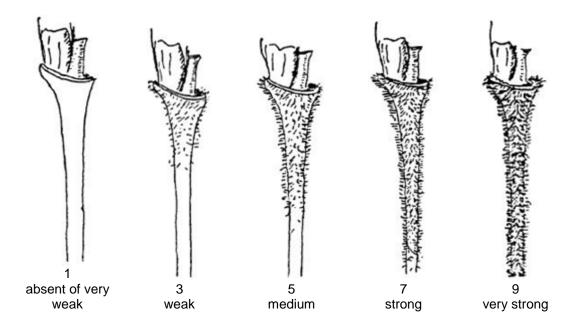
Ad. 4: Flag leaf: anthocyanin coloration of auricles

The appropriate scoring time between stages 49 and 60 should be determined depending on the location. All varieties should be assessed at the same stage

Ad. 5: Time of ear emergence

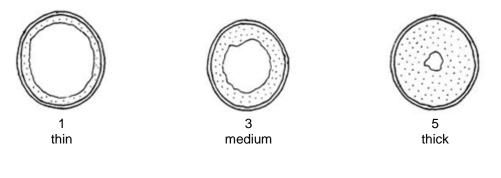
Time of ear emergence is reached when the first spikelet is visible on 50% of ears.

Ad. 11: Stem: density of hairiness of neck



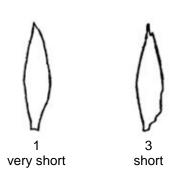
Ad. 13: Straw: pith in cross section

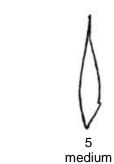
Pith in cross section should be observed half way between base of ear and uppermost node. All stems of the plant should be checked and the highest score per plant recorded



5

Ad. 14: Lower glume: length of first beak

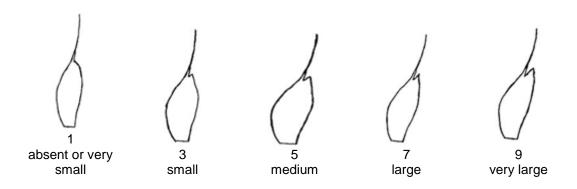








Ad. 15: Lower glume: size of second beak



Ad. 17: Plant: length

The length of plant includes stem, ear, awns and scurs

Ad. 18: Ear: color

White ear varieties may be slightly colored due to environmental conditions

Ad. 19: Ear: density

The density is the ratio of the number of spikelets per ear length

Ad. 20: Ear: distribution of awns





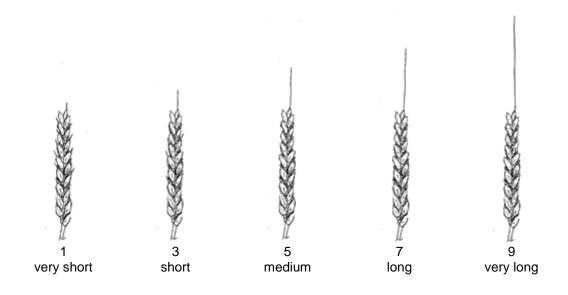


3 fully awned

1 tip awned

2 half awned

Ad. 22: Ear: length of scurs or awns



Ad. 23: Ear: length

Length of ear should be observed excluding awns and scurs

Ad. 24: Grain: coloration with phenol

Method for Determination of Phenol Reaction: Number of seeds per test: 100 seeds. The seeds should not have been treated chemically. Preparation of seeds: Soak in tap water for 16 to 20 hours, drain and remove surface water, place the seeds with crease downwards, cover dish with lid Concentration of solution: 1 per cent Phenol-solution (freshly made up) Amount of solution: The seeds should be about 3/4 covered Place: Laboratory Light: Daylight - out of direct sunshine Temperature: 18 to 20°C Time of recording: 4 hours (after adding solution) Note: At least two example varieties should be included as a control Any alternative method may be used if it gives the same results

Ad. 25: Seasonal type

The seasonal type (need of vernalization) should be assessed on plots sown in springtime. Example varieties should always be included in the trial. When the example varieties behave according to their descriptions, candidate varieties can be described. At the time when the latest spring type variety is fully mature (stage 91/92 of the Zadoks decimal code) growth stage reached by the respective variety should be assessed. The states of expression are defined as follows:

1- Winter type (high need of vernalization): the plants have reached stage 45 of the Zadoks decimal code (boots swollen) at maximum.

2- Alternative type (partial need of vernalization): the plants have exceeded stage 45 of the Zadoks decimal code (they should have normally exceeded stage 75) and have reached stage 90 at maximum.
3- Spring type (no need or very weak need of vernalization): the plants have exceeded stage 90 of the Zadoks decimal code

8.2 The descriptions of the growth stages of the Zadoks decimal code for cereals

00Dry seed40-01Start of imbibition41Flag leaf sheath extending03Imbibition complete43Boots just visibly swollen05Radicle emerged from seed47Flag leaf sheath opening09Leaf just at coleoptile tip49First saft sheath opening09Leaf just at coleoptile tip49First saft spiklelt of inflorescence visible11First leaf through coleoptile50First spiklelt of inflorescence emerged122 leaves unfolded573/4 of inflorescence emerged133 leaves unfolded573/4 of inflorescence completed155 leaves unfolded60Beginning on anthesis166 leaves unfolded60Anthesis completed177 leaves unfolded70-188 leaves unfolded70-199 or more leaves unfolded71Kernel watery ripe20Main shoot and 2 tillers77Late milk21Main shoot and 3 tillers80-24Main shoot and 3 tillers80-25Main shoot and 6 tillers87Hard dough26Main shoot and 3 tillers90-27Main shoot and 7 tillers90-28Main shoot and 7 tillers91Kernel hard (difficult to divide with thumbnail)29Main shoot and 9 or more tillers92Kernel hard (difficult to divide with thumbnail)30Pseudo stem erection93 <td< th=""><th>Zadoks Decimal code</th><th>Description</th><th>Zadoks Decimal code</th><th>Description</th></td<>	Zadoks Decimal code	Description	Zadoks Decimal code	Description
39 Flag leaf ligule/collar just visible	$\begin{array}{c} 01\\ 03\\ 05\\ 07\\ 09\\ 10\\ 11\\ 12\\ 13\\ 14\\ 15\\ 16\\ 17\\ 18\\ 19\\ 20\\ 21\\ 22\\ 23\\ 24\\ 25\\ 26\\ 27\\ 28\\ 29\\ 30\\ 31\\ 32\\ 33\\ 34\\ 35\\ 36\\ 37\\ \end{array}$	Start of imbibition Imbibition complete Radicle emerged from seed Coleoptile emerged from seed Leaf just at coleoptile tip First leaf through coleoptile First leaf unfolded 2 leaves unfolded 3 leaves unfolded 4 leaves unfolded 5 leaves unfolded 6 leaves unfolded 8 leaves unfolded 9 or more leaves unfolded Main shoot only Main shoot and 1 tiller Main shoot and 2 tillers Main shoot and 3 tillers Main shoot and 5 tillers Main shoot and 5 tillers Main shoot and 7 tillers Main shoot and 8 tillers Main shoot and 9 or more tillers Pseudo stem erection 1st node detectable 3rd node detectable 3rd node detectable 5th node detectable 6th node detectable Flag leaf just visible	41 43 45 47 49 50 53 55 57 59 60 65 69 70 71 73 75 77 80 83 85 87 90 91 92 93 94 95 96 97 98	Boots just visibly swollen Boots just swollen Flag leaf sheath opening First awns visible First spikelet of inflorescence visible 1/4 of inflorescence emerged 3/4 of inflorescence emerged Emergence of inflorescence completed Beginning on anthesis Anthesis half-way Anthesis completed - Kernel watery ripe Early milk Medium milk Late milk - Early dough Soft dough Hard dough - Kernel hard (difficult to divide with thumbnail) Kernel hard (no longer dented with thumbnail) Kernel loosening in daytime Overripe, straw dead and collapsing Seed dormant Viable seed giving 50% germination Seed not dormant Secondary dormancy induced

9. <u>Literature</u>



10. <u>Technical Questionnaire</u>

TECH	NICAL Q	UESTIONNAIRE		Page {x} of {y}	Reference Number:	
					Application date: (not to be filled in by the applicant))
		to be completed in c		CHNICAL QUESTIO	IRE for plant breeders' rights	
1.	Subject	of the Technical Questic	onnai	ire		
	1.1	Botanical name	×7	<i>riticosecale</i> Witt.		
	1.2	Common name	Tri	iticale		
2.		s one No. address r (if different from				
3.	Propos (if avail	ed denomination and bre ed denomination able) r's reference		's reference		

TECHNICAL G	QUESTIONNAIRE	Page {x} of {y}		Reference Number:		
#4. Information on the breeding scheme and propagation of the variety						
4.1	4.1 Breeding scheme					
Variety	resulting from:					
4.1.1	Crossing					
(a)	controlled cross (please state parent varietie	es)		[]		
	()	x	()		
	female parent			male parent		
(b)	partially known cross (please state known parent	variety(ies))		[]		
	()	x	()		
	female parent			male parent		
(c)	unknown cross			[]		
4.1.2	Discovery and development (please state where and wh	: en discovered and hov	v de	[] veloped)		
4.1.3	Mutation (please state parent variety)			[]		
4.1.4	Other (Please provide details)			[]		

TECHNICAL C	QUESTIONNAIRE	Page {x} of {y}	Reference Number:	
4.2 4.2.1	Method of propagating th Other (Please provide details)	ie variety	[]

TECH	NICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:	
	Characteristics of the variety to be characteristic in Test Guidelines; p		prackets refers to the corresponding h best corresponds).	
	Characteristics		Example Varieties	Note
5.1 (5)	Time of ear emergence			
	very early		Chopper	1[]
	early		Prime 322	3[]
	medium		Coral Sea	5[]
	late		Crackerjack	7[]
	very late		Pacific Falcon	9[]
5.2 (17)	Plant: length			
	very short			1[]
	short		Chopper	3[]
	medium		Endeavour	5[]
	long		Forerunner	7[]
	very long			9[]
5.3 (18)	Ear: color			
	white		Austute	1[]
	slightly colored		Forerunner	2[]
	strongly colored			3[]
5.4 (25)	Seasonal type			
	winter type		Coral Sea	1[]
	alternative type		Breakwell	2[]
	spring type		Austute	3[]

TECHNICAL QUESTIONNAIRE	Page {x} of {y}	Reference Number:					
6. Similar varieties and differences from these varieties Please use the following table and box for comments to provide information on how your candidate variety differs from the variety (or varieties) which, to the best of your knowledge, is (or are) most similar. This information may help the examination authority to conduct its examination of distinctness in a more efficient way.							
Denomination(s) of Characteristic variety(ies) similar to your candidate candidate variety from the similar	variety differs the characte	e expression of Describe the expression of eristic(s) for the variety(ies) candidate variety					
Example							
Comments:							

TECHNICAL QUESTIONNAIRE		RE Page {x} of {y	y} Reference Number:						
#7.	Additional information	which may help in the examir	nation of the variety						
7.1	In addition to the inform help to distinguish the	•	and 6, are there any additional characteristics which may						
	Yes []	No	[]						
	(If yes, please provide	details)							
7.2	Are there any special	conditions for growing the va	riety or conducting the examination?						
	Yes []	No	[]						
	(If yes, please provide details)								
7.3	Other information								

TEC	HNICA	L QUESTIONNAIRE	Page {x} of {y}	Referenc	e Number:				
8.	Autho	prization for release							
	(a)	a) Does the variety require prior authorization for release under legislation concerning the protection of the environment, human and animal health?							
		Yes []	No []						
	(b)	Has such authorization I	been obtained?						
		Yes []	No []						
	If the	answer to (b) is yes, pleas	se attach a copy of the aut	norization.					
9. In	formati	on on plant material to be	examined or submitted for	examination					
	s and	disease, chemical treatm	eristic or several character ent (e.g. growth retardan t growth phases of a tree,	ts or pesticides),					
char has	acterist underg	tics of the variety, unless to one such treatment, full d	have undergone any tre the competent authorities a etails of the treatment mus at material to be examined	allow or request s at be given. In this	uch treatment.	If the plant materia			
	(a)	Microorganisms (e.	g. virus, bacteria, phytoplas	sma)	Yes []	No []			
	(b)	Chemical treatment	(e.g. growth retardant, pes	sticide)	Yes []	No []			
	(c)	Tissue culture			Yes []	No []			
	(d)	Other factors			Yes []	No []			
	Ple	ase provide details for wh	ere you have indicated "ye	S".					
10.	l he	ereby declare that, to the b	est of my knowledge, the i	nformation provide	ed in this form is	s correct:			
	Ар	olicant's name							
	Sic	gnature		Date					
L									

[End of document]