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|  |  | ETC/51/27**ORIGINAL:** EnglishDATE: February 20, 2015 |
| INTERNATIONAL UNION FOR THE PROTECTION OF NEW VARIETIES OF PLANTS  |
| Geneva |

Technical Committee

Fifty-First Session
Geneva, March 23 to 25, 2015

PARTIAL REVISION of the Test Guidelines for French Bean
(document TG/12/9 Rev.)

*Document prepared by the Office of the Union

Disclaimer: this document does not represent UPOV policies or guidance*

 At its forty-eighth session held in Paestum, Italy, from June 23 to 27, 2014, the Technical Working Party for Vegetables (TWV) considered a partial revision of the Test Guidelines for French Bean on the basis of documents TG/12/9 Rev. and TWV/48/29 “Partial Revision of the Test Guidelines for French Bean (Document TG/12/9 Rev.)” and proposed to revise the Test Guidelines for French Bean as follows (see document TWV/48/43 “Report”, paragraph 97):

 (a) Proposal to revise characteristics 49 to 52

 (b) Proposal to include a revised format for disease resistance characteristics under Chapter 8.2

 The proposed revisions are presented in the Annex to this document.

[Annex follows]

Proposal to Revise Characteristics 49 to 52

*Current wording:*

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| **49.(+)** |  | **Resistance to Bean anthracnose (*Colletotrichum lindemuthianum*)** | **Résistance à l’anthracnose du Haricot (*Colletotrichum lindemuthianum*)** | **Resistenz gegen Brennfleckenkrankheit (*Colletotrichum lindemuthianum*)** | **Resistencia a la antracnosis de la judía (*Colletotrichum lindemuthianum*)** |  |  |
| **49.1(\*)** | **VS/VG** | **Race 6** | **Pathotype 6** | **Pathotyp 6** | **Patotipo 6** |  |  |
| **QL** |  | absent | absente | fehlend | ausente | Goldrush, Masaï, Michelet | 1 |
|  |  | present | présente | vorhanden | presente | Booster, Pastoral | 9 |
| **49.2** | **VS/VG** | **Race Kappa** | **Pathotype Kappa** | **Pathotyp Kappa** | **Patotipo Kappa** |  |  |
| **QL** |  | absent | absente | fehlend | ausente | Goldrush, Masaï, Michelet | 1 |
|  |  | present | présente | vorhanden | presente | Booster, Pastoral | 9 |

*Proposed new wording:*

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| **49.(+)** |  | **Resistance to “Colletotrichum lindemuthianum” (Cl)** | **Résistance à “Colletotrichum lindemuthianum” (Cl)** | **Resistenz gegen “Colletotrichum lindemuthianum” (Cl)** | **Resistencia a “Colletotrichum lindemuthianum” (Cl)** |  |  |
| **49.1(\*)** | **VS/VG** | **Race 6** | **Pathotype 6** | **Pathotyp 6** | **Patotipo 6** |  |  |
| **QL** |  | absent | absente | fehlend | ausente | Goldrush, Masai, Michelet à longue cosse | 1 |
|  |  | present | présente | vorhanden | presente | Booster, Pastoral | 9 |
| **49.2** | **VS/VG** | **Race Kappa** | **Pathotype Kappa** | **Pathotyp Kappa** | **Patotipo Kappa** |  |  |
| **QL** |  | absent | absente | fehlend | ausente | Goldrush, Masai, Michelet à longue cosse | 1 |
|  |  | present | présente | vorhanden | presente | Booster, Pastoral | 9 |

*Current wording:*

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| **50.(\*)(+)** | **VS/VG** | **Resistance to Bean Common Mosaic Necrosis Virus (BCMNV)** | **Résistance au virus de la mosaïque nécrotique commune du Haricot (BCMNV)** | **Resistenz gegen Gewöhnliches nekrotisches Bohnenmosaikvirus (BCMNV)** | **Resistencia al virus del mosaico necrotico común de la judía (BCMNV)** |  |  |
| **PQ** |  | absent | absente | fehlend | ausente | Dufrix, Flandria | 1 |
|  |  | present with necrosis | présente avec nécroses | vorhanden mit Nekrose | presente con necrosis | Booster, Odessa | 2 |
|  |  | present without symptoms | présente sans symptômes | vorhanden ohne Symptome | presente sin síntomas | Bizet | 3 |

*Proposed new wording:*

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| **50.(\*)(+)** | **VS/VG** | **Resistance to “Bean common mosaic necrosis virus” (BCMNV)** | **Résistance au “Bean common mosaic necrosis virus” (BCMNV)** | **Resistenz gegen “Bean common mosaic necrosis virus” (BCMNV)** | **Resistencia al “Bean common mosaic necrosis virus” (BCMNV)** |  |  |
| **PQ** |  | absent | absente | fehlend | ausente | Dufrix, Flandria | 1 |
|  |  | present with necrosis | présente avec nécroses | vorhanden mit Nekrose | presente con necrosis | Booster, Odessa | 2 |
|  |  | present without symptoms | présente sans symptômes | vorhanden ohne Symptome | presente sin síntomas | Bizet | 3 |

*Current wording:*

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| **51.(+)** | **VS/VG** | **Resistance to Halo Blight (*Pseudomonas syringae* pv. *phaseolicola*)** | **Résistance à la graisse à halo (*Pseudomonas syringae* pv. *phaseolicola*)** | **Resistenz gegen Fettfleckenkrankheit (*Pseudomonas syringae* pv. *phaseolicola*)** | **Resistencia a la grasa (*Pseudomonas syringae* pv. *phaseolicola*)** |  |  |
|  |  | **Race 6** | **Pathotype 6** | **Pathotyp 6** | **Patotipo 6** |  |  |
| **QL** |  | absent | absente | fehlend | ausente | Michelet (D) | 1 |
|  |  | present | présente | vorhanden | presente | Masai (D), Vaillant (D) | 9 |

*Proposed new wording:*

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| **51.(+)** | **VS/VG** | **Resistance to “Pseudomonas savastanoi pv. phaseolicola” (Psp)** | **Résistance à “Pseudomonas savastanoi pv. phaseolicola” (Psp)** | **Resistenz gegen “Pseudomonas savastanoi pv. phaseolicola” (Psp)** | **Resistencia a “Pseudomonas savastanoi pv. phaseolicola” (Psp)** |  |  |
|  |  | **Race 6** | **Pathotype 6** | **Pathotyp 6** | **Patotipo 6** |  |  |
| **QL** |  | absent | absente | fehlend | ausente | Michelet à longue cosse (D) | 1 |
|  |  | present | présente | vorhanden | presente | Masai (D), Vaillant (D) | 9 |

*Current wording:*

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| **52.(+)** | **VG** | **Resistance to Common Blight (*Xanthomonas campestris* pv. *phaseoli*), Isolate 422** | **Résistance à la graisse commune (*Xanthomonas campestris* pv. *phaseoli*), Isolate 422** | **Resistenz gegen Bohnenbrand (*Xanthomonas campestris* pv. *phaseoli*), Isolat 422** | **Resistencia a la grasa común (*Xanthomonas campestris* pv. *phaseoli*), Isolate 422** |  |  |
| **QL** |  | absent | absente | fehlend | ausente  | Echo (D), Keygold (D) | 1 |
|  |  | present | présente | vorhanden | presente | Walley (US line) (D) | 9 |

*Proposed new wording:*

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| **52.(+)** | **VG** | **Resistance to “Xanthomonas axonopodis pv. phaseoli” (Xap)** | **Résistance à “Xanthomonas axonopodis pv. phaseoli” (Xap)** | **Resistenz gegen “Xanthomonas axonopodis pv. phaseoli” (Xap)** | **Resistencia a “Xanthomonas axonopodis pv. phaseoli” (Xap)** |  |  |
| **QL** |  | absent | absente | fehlend | ausente  | Echo (D), Keygold (D) | 1 |
|  |  | present | présente | vorhanden | presente | Walley (US line) (D) | 9 |

Proposal to Include a Revised Format for Disease Resistance Characteristics

*Current wording:*

Ad. 49: Resistance to Bean anthracnose (*Colletotrichum lindemuthianum*)

|  |  |  |
| --- | --- | --- |
| Maintenance of races |  | In a test tube on glucose-peptone agar |
| Pre-germination of seed (about 4 to 5 days) |  | At least twice, 10 seeds are placed at 20°C in petri-dishes on moist vermiculite. After the start of germination (1 to 2 cm root length) the seed coat is removed. |
| Inoculum and inoculation |  | Growth on GPA in 1 liter glass bottles for 12 to 14 days. Removal of inoculum with a scraper. The germinated seeds are dipped in a suspension of spores of *Colletotrichum lindemuthianum* for 2 minutes. The concentration of spores should be 1 million spores per ml |
| Sowing: |  | Sowing in pots with sand, covering of seed with sand to 1 cm. |
| Culture of plants: |  | The pots are placed in a Phytotron at 20°C with 16 hours of daylight. Regular watering is needed, no special air humidity requirements. |
| Observation: |  | The symptoms are visible during sprouting of the plants or up to 10 days thereafter. The observations can be made after 10 to 14 days. |
| Scheme of observation: |  | Resistance present: healthy plants with no symptoms, or weak reaction with small superficial necroses in the form of dots or stripesResistance absent: reaction with up to 5 necrotic flecks on stem, or strong reaction with necroses larger than 3 mm, sunk deeply into the tissue, or dying plants with strong formation of necroses during sprouting or thereafter. |

*proposed new wording:*

Ad. 49: Resistance to “Colletotrichum lindemuthianum” (Cl)

|  |  |  |
| --- | --- | --- |
| 1. | Pathogen | “Colletotrichum lindemuthianum”(Cl) |
| 2. | Quarantine status | No |
| 3. | Host species | *Phaseolus vulgaris* |
| 4. | Source of inoculum | GEVES (FR), Naktuinbouw (NL), INIA (ES) |
| 5. | Isolate | 6, Kappa  |
| 6. | Establishment isolate identity | On differentials: |

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
|   | Old race name: |   |   | - | (no longer in TG) Lambda  | Kappa |
|   | Binary race name: |   |   | 6 | 55 | 31 |
| **Differential** | Gene | Binary |  |  |  |
| A | Michelite |   | 1 | R | S | S |
| B | Michigan Dark Red Kidney | Co-1 | 2 | S | S | S |
| C | Perry Marrow | Co-13 | 4 | S | S | S |
| D | Cornell 49242 | Co-2 (Are) | 8 | R | R | S |
| E | Widusa | Co-15 | 16 | R | S | S |
| F | Kaboon | Co-12 | 32 | R | S | R |
| G | Mexico 222 | Co-3 | 64 | R | R | R |
| H | PI 207262 |   | 128 | R | R | R |
| I | TO | Co-4 | 256 | R | R | R |
| J | TU | Co-5 | 512 | R | R | R |
| K | AB 136 | Co-6 | 1024 | R | R | R |
| L | G 2333 | Co-4-2/5/7 | 2048 | R | R | R |

|  |  |  |
| --- | --- | --- |
| 7. | Establishment pathogenicity | On susceptible variety |
| 8. | Multiplication inoculum |  |
| 8.1 | Multiplication medium | PDA (Potato Dextose Agar) or Mathur medium (20-25°C) |
| 8.2 | Multiplication variety | - |
| 8.3 | Plant stage at inoculation | Seed for soaking 5 days old seedlings for spraying |
| 8.4 | Inoculation medium | - |
| 8.5 | Inoculation method | Soaking or spraying seedlings |
| 8.6 | Harvest of inoculum | Scrape spores with scraper from 7-20 days old plates grown at 20-25°C |
| 8.7 | Check of harvested inoculum | Count spores and adjust to 106 spores per mL |
| 8.8 | Shelflife/viability inoculum | About 4 hoursLong term storage of strains: at -80°C in 20% glycerol |
| 9. | Format of the test |  |
| 9.1 | Number of plants per genotype | At least 20 plants |
| 9.2 | Number of replicates | - |
| 9.3 | Control varieties |  |
|  | Susceptible: | Goldrush, Michelet à longue cosse, Masai  |
|  | Resistant for race 6 and race Lambda: | Booster, Pastoral  |
| 9.4 | Test design | - |
| 9.5 | Test facility | Climate cell |
| 9.6 | Temperature | 20-22°C |
| 9.7 | Light | - |
| 9.8 | Season | - |
| 9.9 | Special measures | Plants are placed in high humidity |
| 10. | Inoculation |  |
| 10.1 | Preparation inoculum | Culture on PDA or Mathur medium |
| 10.2 | Quantification inoculum | Count spores and adjust to 106 spores per mL |
| 10.3 | Plant stage at inoculation | Pre-germinated seed for soaking 5 days old seedlings for spraying |
| 10.4 | Inoculation method | One of two methods may be applied: - Soaking pre-germinated seeds in a spore suspension for 2 minutes. Seeds are planted in soil after inoculation - Spraying cotyledons with inoculum suspension 5 days after sowing |
| 10.5 | First observation | 7 days after inoculation |
| 10.6 | Second observation | 12 days after inoculation |
| 10.7 | Final observations | 14 days after inoculation |
| 11. | Observations |  |
| 11.1 | Method | Visual observation of symptoms |
| 11.2 | Observation scale | 0: no symptoms1: weak reaction with small superficial necrosis (dots or stripes)2: necrotic lesions larger than 3 mm and/or deeply sunk into the tissue of hypocotyls and/or stems3: dying plants |
| 11.3 | Validation of test | Control varieties must show expected symptoms |
| 11.4 | Off-types | - |
| 12. | Interpretation of data in terms of UPOV characteristic states | - |
|  | For soaking seeds: | Resistant [9]: class 0 and 1Susceptible [1]: class 2 and 3 |
|  | For spraying cotyledons: | Some flecks of necrosis can occur in the stem and some in the cotyledons of resistant varieties |
| 13. | Critical control points | Monitor the inoculation pressure with a suitable variety e.g. with Pastoral. This variety has a weaker resistance and can give an indication of aggressiveness of the test. |

*Current wording:*

Ad. 50: Resistance to Bean Common Mosaic Necrosis Virus (BCMNV)

|  |  |  |
| --- | --- | --- |
| Production of infection material |  |  |
| Nature of medium: |  | Plants or dry leaves |
| Special conditions: |  | Glasshouse culture (plants) or deep-frozen leaves |
| Identification: |  | Use of virus strain “NL 3” |
| Conduct of trials |  |  |
| Plant stage: |  | Two-leaf |
| Temperature: |  | Culture at 20 to 25°C, following inoculation 30°C for a period of 8 days |
| Light: |  | Normal daylight, if necessary shaded |
| Culture: |  | Glasshouse |
| Type of inoculation: |  | Mechanical, by rubbing the inoculum on the leaves |
| Duration of trials |  |  |
| - Sowing to inoculation: |  | 8 to 9 days |
| - Inoculation to observation: |  | 6 to 21 days |
| Number of plants tested: |  | 60 (20 pots with 3 plants each) |

|  |
| --- |
| Description of the Method |
|  |
| (1)  Obtaining the inoculation material.-  The virus strain “NL 3” is used for the tolerance testing since it covers practically all the groups of strains of Bean Common Mosaic Virus. To begin with, dwarf bean plants of the variety “Dufrix” or of another variety highly sensitive to the virus are infected, around the beginning of Spring, by rubbing with pressed juice containing the virus, obtained from own maintenance culture or from freeze-dried leaves (provided for instance by the Institute for Biochemistry and Virus Diseases of the Federal Biological Institute in Brunswick (= strain “NL 3”)). These infected plants are then used, around two months later, for producing pressed juice containing the virus with which the test plants are inoculated. |
| (2)  Inoculation.-  The pressed juice containing the virus is diluted for inoculation (approximately one part juice to two parts water). After the two leaves have been strewn with carborundum or celite, the diluted juice is lightly rubbed on using a firm sponge. The leaves are then rinsed with water some 15 to 20 minutes later using a watering can with a fine spout.(3)  Incubation.-  Following inoculation, the air temperature in the glasshouse must be kept at 30°C for at least one week. (Important!!! The temperature must be maintained throughout the day and also at night). First lesions may already occur after 3 to 4 days. Top necrosis will already become visible one week after inoculation. Varieties with tolerance absent demonstrate the typical mosaic symptoms after approximately two weeks. The final observations can be made some three weeks after inoculation.(4)  Observation:  The first assessment should be made on the sixth day following the day of innoculation. The mosaic symptoms and the necrotic symptoms can be distinguished as follows: (i)  Mosaic symptoms: pale-colored leaves; light and dark green mosaic; dark green areas between veins blistered; narrow chlorotic bands along veins and leaf margin rolling downwards. Various symptoms may be expressed in various degrees. The mosaic symptoms may be recorded using a scale from 1 to 9 to assess the reaction of the candidate variety(1 = no symptoms, 9 = strongest stage of expression). If a candidate variety does not show any mosaic symptoms, while the susceptible standard varieties do so, that candidate variety should be regarded as being resistant to mosaic. (ii)  Blackroot symptoms: there are two types of necrosis (especially when tested with strain “NL3”), which are to be classified as “Blackroot.” Local necrosis (local hypersensitivity): characterized by brown necrotic netting (the veins) localized on a part of the leaf blade; Systemic necrosis (top necrosis): characterized by a rapid development of necrosis through out the stem, the petiole and the roots, resulting in top necrosis or even complete necrosis of the plant. (The vascular bundles of the stem, the petiole and finally the roots, if innoculated at a young plant stage, turn brown, hence the term “Blackroot”).Varieties or strains showing blackroot symptoms (both local hypersensitivity and top necrosis) generally prove to be resistant to mosaic in the field. During the resistance testing most local necroses develop into top necroses.  Remarks:  The genetics of resistance to Bean Common Mosaic Virus (BCMV) and/or Blackroot is based on a number of a-specific and specific recessive genes of which some are allelic. Drijfhout found at least 4 genes; e.g.: bc-u bc-1/bc-12 bc-2/bc-22 and bc-3. A dominant necrosis gene ‘I’ interferes with these resistance genes. The recessive form ‘I+’ in combination with bc-3 and bc-22 gives complete resistance to both BCMV and Blackroot (Example variety: Great Northern 31). (for more details, see Drijfhout (1978)) |

*proposed new wording:*

Ad. 50: Resistance to “Bean common mosaic necrosis virus” (BCMNV)

|  |  |  |
| --- | --- | --- |
| 1. | Pathogen | “Bean common mosaic necrosis virus” (BCMNV) |
| 2. | Quarantine status | No |
| 3. | Host species | *Phaseolus vulgaris* |
| 4. | Source of inoculum | GEVES (FR), Naktuinbouw (NL), INIA (ES) |
| 5. | Isolate | NL3 or NL5 (Pathogenicity group VI) |
| 6. | Establishment isolate identity | On differentials Widusa and Top Crop;Widusa (I) must show top or vein necrosis;Top Crop (bc-1, I) must show only local necrosis |
| 7. | Establishment pathogenicity | On susceptible variety |
| 8. | Multiplication inoculum |  |
| 8.1 | Multiplication medium | - |
| 8.2 | Multiplication variety | Dufrix or Flandria |
| 8.3 | Plant stage at inoculation | First leaf expanded (8-12 days) |
| 8.4 | Inoculation medium | PBS (Phosphate Buffer Saline) and carborundum |
| 8.5 | Inoculation method | Rubbing |
| 8.6 | Harvest of inoculum | Pick leaves with mosaic and/or leaf rolling 14 days after inoculation on susceptible variety |
| 8.7 | Check of harvested inoculum | - |
| 8.8 | Shelflife/viability inoculum | Very long in dry or freeze dried leaves  |
| 9. | Format of the test |  |
| 9.1 | Number of plants per genotype | 20 |
| 9.2 | Number of replicates | 2 |
| 9.3 | Control varieties |  |
|  | Susceptible: | Dufrix, Flandria |
|  | Resistant with necrosis: | Booster, Odessa |
|  | Resistant without necrosis: | Bizet |
| 9.4 | Test design | Glasshouse or climatic chamber |
| 9.5 | Test facility | Glasshouse |
| 9.6 | Temperature | Initial 5-7 days after inoculation:25° day / 18°C night or 30°C day and nightAfter 5-7 days:25°C day and night |
| 9.7 | Light | See remark 13. |
| 9.8 | Season | - |
| 9.9 | Special measures | Rinse leaves after inoculation to reduce damage by carborundum |
| 10. | Inoculation |  |
| 10.1 | Preparation inoculum | Maceration in PBS |
| 10.2 | Quantification inoculum | - |
| 10.3 | Plant stage at inoculation | First leaf expanded (8-12 days after sowing) |
| 10.4 | Inoculation method | Rubbing |
| 10.5 | First observation | 6 days after inoculation |
| 10.6 | Second observation | 9 days after inoculation |
| 10.7 | Final observations | 14 days after inoculation |
| 11. | Observations |  |
| 11.1 | Method | Visual observation |
| 11.2 | Observation scale | 1: mosaic and/or leaf rolling2: top necrosis, vein necrosis and/or small necrotic lesions3: no symptoms |
| 11.3 | Validation of test | Control varieties must show expected symptoms |
| 11.4 | Off-types | - |
| 12. | Interpretation of data in terms of UPOV characteristic states | Classify in three classes corresponding with observation scale:1: resistant absent2: resistant present with necrosis3: resistant present without necrosis |
| 13. | Critical control points | Temperature-dependent expression of symptoms in some varieties, necrosis increasing with temperature. Light may also enhance symptom development. |

*Current wording:*

Ad. 51: Resistance to Halo Blight (*Pseudomonas syringae* pv. *phaseolicola*)

|  |  |  |
| --- | --- | --- |
| Maintenance of strains |  |  |
| Type of medium |  | Infected, dry leaves |
| Identification: |  | On the basis of preliminary trials, the European strains (which probably belong to the African race-by J.D. Taylor, H.R.I. Wellesbourne) have a higher level of virulence than the US race 1 and race 2. The aggressivity of the pathogen is measured by the spot size of the pod of sensitive varieties. The isolates used for the test should cause a grease spot with a minimum diameter of 3 mm. |
| Execution of test |  |  |
| Growth stage of plants: |  | When the first and second trifoliate leaves are 2 to 3 cm in length |
| Temperature: |  | Day: 24°C; night: 18°C |
| Humidity: |  | 100% relative humidity until inoculated leaves are fully developed |
| Growing method: |  | In the glasshouse |
| Inoculum: |  | Bacterial suspension with a concentration of 108 bacterial cells/ml. |
| Method of inoculation: |  | Mechanical, using a camel-hair brush |
| Duration of test |  |  |
| - from inoculation to reading: |  | Until infected leaves are fully developed |
| Number of plants to be tested: |  | 10-20 plants |
| Multiplication/propagation of bacteria: |  | Bouillon-Agar (2 g Na2 HPO4, 2 g NaH2PO4, 3 g NaCl, 25 g Bouillon-Agar/1000 ml distilled water) |
| Remarks: |  | -  Leaf reaction is very commonly studied nowadays. The reaction of the pod is of polygenic character, and there is no genetic linkage between leaf and pod reaction. There are as yet no varieties with pod resistance.-  Resistance means, genetically, that this host has the recessive gene with or without the presence of the modifiers; in the case where the modifiers are present the sources of these genes are: PI 150 414 (USA), CNRA-HW5A (Fr.).It is possible to evaluate the lesions at the stage of the fully developed leaf. The different types of symptom are shown below. |

Legend of illustration following hereafter



healthy tissue     water-soaked lesion without discoloration

toxically chlorotic tissue     water-soaked lesion with discoloration



some cell-size brownish red

necrotic spots

Scheme of observation

Resistance absent



 water-soaked lesion with toxically

 chlorotic halo, systemic chlorosis;

 water-soaked lesion with halo, no

 systemic chlorosis;

 water-soaked lesion without halo,

 no systemic chlorosis



    discoloration of water-soaked lesions

    with halo, systemic chlorosis;

    discoloration of water-soaked lesions with

    halo, no systemic chlorosis

Resistance present



necrotic spots of 1-2 mm diameter, no systemic chlorosis or some cell-size brownish-red hypersensitive necrotic spots or healthy, uninfected plant

*Proposed new wording:*

Ad. 51: Resistance to “Pseudomonas savastanoi pv. phaseolicola” (Psp)

|  |  |  |
| --- | --- | --- |
| 1. | Pathogen | “Pseudomonas savastanoi pv. phaseolicola” (Psp) |
| 2. | Quarantine status | No |
| 3. | Host species | *Phaseolus vulgaris* |
| 4. | Source of inoculum | GEVES (FR), Naktuinbouw (NL), HRI (GB), INIA (ES) |
| 5. | Isolate | Race 6 |
| 6. | Establishment isolate identity | All differentials should be susceptible (Canadian Wonder, A52, Red Mexican UI3, Mesunka, A53, A43, Guatemala 196-B) |
| 7. | Establishment pathogenicity | On susceptible variety |
| 8. | Multiplication inoculum |  |
| 8.1 | Multiplication medium | King’s B or Yeast Dextrose Agar at 27°C |
| 8.2 | Multiplication variety | - |
| 8.3 | Plant stage at inoculation | First leaf (9-14 days after sowing) |
| 8.4 | Inoculation medium | Tap water or saline solution (0.85% NaCl) |
| 8.5 | Inoculation method | - |
| 8.6 | Harvest of inoculum | 4 days after start of pure culture |
| 8.7 | Check of harvested inoculum | - |
| 8.8 | Shelflife/viability inoculum | The number of subculturing before inoculation shall not exceed 2 and the inoculation shall be done within 2-3 days. |
| 9. | Format of the test |  |
| 9.1 | Number of plants per genotype | 20 |
| 9.2 | Number of replicates | 2 |
| 9.3 | Control varieties |  |
|  | Susceptible | Michelet à longue cosse |
|  | Resistant | Masai, Vaillant |
| 9.4 | Test design | - |
| 9.5 | Test facility | Glasshouse or climate cell |
| 9.6 | Temperature | 22/20°C day/night or 20°C day and night |
| 9.7 | Light | - |
| 9.8 | Season | - |
| 9.9 | Special measures | High humidity required during first 1-3 days after inoculation |
| 10. | Inoculation |  |
| 10.1 | Preparation inoculum | Rinse bacteria from plate with tap water and add 2 g carborundum per 100 ml or rinse bacteria with saline solution (0.85% NaCl). |
| 10.2 | Quantification inoculum | 108 cfu/ ml or 1-2 full-grown plates per 100 ml water for 100 plants |
| 10.3 | Plant stage at inoculation | First pair of leaves spreading (9-14 days after sowing) |
| 10.4 | Inoculation method | Rubbing with sponge or inoculation by spraying leaves with pressure (2 bars) until runoff. For this purpose several types of equipment may be used: atomizer or paint brush with a pressure supplier. |
| 10.5 | First observation | 7 days after inoculation |
| 10.6 | Second observation | 14 days after inoculation |
| 10.7 | Final observations | - |
| 11. | Observations |  |
| 11.1 | Method | Visual observation |
| 11.2 | Observation scale |  |
|  | Resistant [9] | No symptoms or necrotic pinpoints |
|  | Susceptible [1] | Light green halo around minute lesionsWater soaked (“oily”) lesions (few or many)Water soaked lesions, later turning necroticDeformation and chlorosis on first trifoliate leavesNecrosis on stemsDying plants |
| 11.3 | Validation of test | Control varieties must show expected symptoms |
| 11.4 | Off-types | - |
| 12. | Interpretation of data in terms of UPOV characteristic states | 11.2 |
| 13. | Critical control points | Inoculation may produce some damage on susceptible and resistant plants.Maintenance of isolate: beware that the colony may die after keeping 3 weeks on plate. |

*Current wording:*

Ad. 52: Resistance to Common Blight (*Xanthomonas campestris* pv. *phaseoli*), Isolate 422

|  |  |  |
| --- | --- | --- |
| Maintenance of races |  |  |
| Type of medium: |  | Infected, dry leaves |
| Execution of test |  |  |
| Growth stage of plants:  |  | When the first and second trifoliate leaves are 2 to 3 cm in length |
| Temperature: |  | Day: 26°C; night: 20°C |
| Humidity: |  | 100% relative humidity during, and 1 to 2 days after, inoculation, thereafter normal relative humidity |
| Growing method: |  | In the glasshouse |
| Inoculum: |  | Bacterial suspension with a concentration of 108 bacterial cells/ml. |
| Method of inoculation: |  | Mechanical, using a camel-hair brush |
| Duration of test |  |  |
| - from inoculation to reading: |  | Until infected leaves are fully developed |
| Number of plants tested: |  | 10-20 plants |
| Multiplication/propagation of bacteria: |  | 20 g extract of yeast powder, 20 g glucose, 20 g CaCO3, 20 g agar-agar/1000 ml distilled water) |
| Remarks: |  | - Isolate 422 can be obtained from the Vegetable Research Institute, 1775 Budapest, P.O. Box 95, Hungary.- The reaction of pods to *X. phaseoli* is not yet clear enough today. |

Legend of illustration following hereafter





 healthy tissue (2)  dying tissues

 (1)  chlorotic tissue (3)  some cell-size brownish red hypersensitive

  necrotic spots

Scheme of observation

If chlorotic tissues (1) and/or dying tissue (2) are observed, the variety should be regarded as non-resistant.

If only some cell-size brownish red hypersensitive necrotic spots (3) are observed, the variety should be regarded as resistant.

Possible combinations of symptoms

Resistance absent



Resistance present



*Proposed new wording:*

Ad. 52: Resistance to “Xanthomonas axonopodis pv. phaseoli” (Xap)

|  |  |  |
| --- | --- | --- |
| 1. | Pathogen | “Xanthomonas axonopodis pv. phaseoli” (Xap) |
| 2. | Quarantine status | yes |
| 3. | Host species | *Phaseolus vulgaris*  |
| 4. | Source of inoculum | Vegetable Research Institute, Budapest (HU) |
| 5. | Isolate | Isolate 422 |
| 6. | Establishment isolate identity | - |
| 7. | Establishment pathogenicity | - |
| 8. | Multiplication inoculum |  |
| 8.1 | Multiplication medium | Yeast Glucose Agar (20 g yeast extract powder, 20 g glucose, 20 g CaCO3, 20 g agar/ 1000 ml distilled water) |
| 8.2 | Multiplication variety | - |
| 8.3 | Plant stage at inoculation | First leaf pair 2-3 cm long |
| 8.4 | Inoculation medium | - |
| 8.5 | Inoculation method | 100% relative humidity during 2 days after inoculation, later normal humidity |
| 8.6 | Harvest of inoculum | - |
| 8.7 | Check of harvested inoculum | - |
| 8.8 | Shelflife/viability inoculum | - |
| 9. | Format of the test |  |
| 9.1 | Number of plants per genotype | - |
| 9.2 | Number of replicates | - |
| 9.3 | Control varieties | - |
| 9.4 | Test design | - |
| 9.5 | Test facility |  |
| 9.6 | Temperature | 26/20°C day/night or 28/25°C day/night |
| 9.7 | Light | - |
| 9.8 | Season | - |
| 9.9 | Special measures | 100% relative humidity during 2 days after inoculation, later normal humidity |
| 10. | Inoculation |  |
| 10.1 | Preparation inoculum | - |
| 10.2 | Quantification inoculum | 108 cfu/ml |
| 10.3 | Plant stage at inoculation | - |
| 10.4 | Inoculation method | Mechanical, with camel hair brush or inoculation by spraying leaves with pressure (2 bars) until run-off. For this purpose several types of equipment may be used: atomizer or paint brush with a pressure supplier. |
| 10.5 | First observation | 7 days after inoculation |
| 10.6 | Second observation | 14 days after inoculation |
| 10.7 | Final observations | When infected leaves are fully developed |
| 11. | Observations |  |
| 11.1 | Method | - |
| 11.2 | Observation scale | Visual |
|  | Susceptible [1] | Extensive necrosis sometimes surrounded by an increasing ring of chlorotic tissue |
|  | Resistant [9] | Cell-sized brownish or red necrotic spots |
| 11.3 | Validation of test | - |
| 11.4 | Off-types | - |
| 12. | Interpretation of data in terms of UPOV characteristic states | 11.2 |
| 13. | Critical control points | - |

[End of Annex and of document]