

BMT/12/16

ORIGINAL: English DATE: May 5, 2010

INTERNATIONAL UNION FOR THE PROTECTION OF NEW VARIETIES OF PLANTS GENEVA

WORKING GROUP ON BIOCHEMICAL AND MOLECULAR TECHNIQUES AND DNA PROFILING IN PARTICULAR

Twelfth Session Ottawa, Canada, May 11 to 13, 2010

DEVELOPMENT OF AN INTERNATIONAL SEED TESTING ASSOCIATION (ISTA)
DNA-BASED APPROACH FOR TESTING VARIETY IDENTITY

Document prepared by experts from ISTA

The document prepared by the experts from ISTA is reproduced in the Annex to this document.

BMT/12/16

ANNEX



Secretariat, Zürichstrasse 50, 8303 Bassersdorf, CH-Switzerland - Phone: +41-44-838 60 00 - Fax: +41-44-838 60 01 - Email: ista.office@ista.ch - http://www.seedtest.org



Development of an ISTA DNA-Based Approach for Testing Variety Identity

DEVELOPMENT OF AN INTERNATIONAL SEED TESTING ASSOCIATION (ISTA) DNA-BASED APPROACH FOR TESTING VARIETY IDENTITY

Casarini E.1, Vicario A.2, Perry D.3, Dollard C.4, Zhang D.5 and Hwu K.6

- ¹.LaRAS (Laboratorio Ricerca ed Analisi Sementi). Facolta di Agraria. Universita di Bologna. Bologna . Italy. *emanuela.casarini@unibo.it*.
- ².Laboratorio de Marcadores Moleculares y Fitopatología. Instituto Nacional de Semillas. CABA. Argentina.
- ³.Grain Research Laboratory. Canadian Grain Commission. Winnipeg. Government of Canada.
- ⁴.Genotyping-Botany, Ottawa Plant Laboratory, Canadian Food Inspection Agency, Ottawa, Canada.
- ⁵.BIOGEVES, GEVES Le Magneraud. Surgeres. France.

Introduction

Many seed testing laboratories are receiving increasing numbers of requests to apply new technologies based on molecular markers for variety identification and genetic purity tests. For this reason, the International Seed Testing Association DNA working group was established to develop a common approach for DNA-based variety verification capable of providing repeatable and reproducible results among ISTA laboratories.

Importance Of Molecular Markers For Seed Testing

The ability of DNA-based markers to efficiently discriminate between closely related varieties has been reported. They are a step forward for variety characterization and verification compared to protein-based methods because they often reveal greater variation and are independent from the environment.

Among the different marker types, microsatellites were chosen as the most appropriate. The value of microsatellite markers for identification and genetic relationship studies in different crops arise from their multi-allelic nature, co-dominant transmission, relative abundance and extensive genome coverage. This methodology has been particularly useful for identification of genotypes and quantification of genetic diversity in a broad number of species such as potatoes, tomatoes, soybeans, rice, wheat, sunflower, sorghum, corn and many others.

Microsatellites, also known as Simple Sequence Repeats (SSR), are short tandemly repeated sequences of di-, tri-, tetra- or penta-nucleotide units, which are found scattered throughout the genomes of most eukaryotic species, making them an interesting tool for variety verification.

⁶.Department of Agronomy, Plant Breeding Laboratory, Taiwan Province of China*

[&]quot;Taiwan Province of China is considered as "Separate Customs Territory of Taiwan, Penghu, Kinmen and Matsu" under the rules of the International Seed Testing Association (ISTA).

The strategy for incorporating DNA-based tests into the ISTA Rules.

For each species, the strategy was to select a single set of markers that may be used worldwide. These sets of markers were selected for each crop from larger sets of markers proposed by crop experts and were evaluated on a diverse set of varieties from various regions through a series of collaborative tests (CTs). These CTs were performed initially by laboratories having experience with each crop and/or technique. Laboratories were free to choose the DNA extraction and PCR protocols and visualization system. This gave robustness to the study. Repeatability and reproducibility for each marker were assessed.

In the future, for their accreditation, laboratories will be required to perform a Proficiency Test. The way in which this will be carried out has not yet been decided in detail.

Species selection:

During the ISTA Congress held in Foz do Iguaçu in 2007 the ISTA DNA experts gathered to decide upon the strategy to follow to incorporate DNA-based methods into the ISTA Rules. There, it was decided that crops representing the most important ones in terms of the cultivated area and production would be chosen for these tests. The selected crops were: maize, wheat, rice and soybean. An expert was chosen to lead each crop working group.

Marker selection:

The markers used during the first CT were selected by each crop leader based on their experience. The subsequent selection of markers per crop was based on the performance of the markers. SSRs that did not provide consistent results among laboratories during the first CT were withdrawn from the marker set and replacements were selected. During the second and third CTs the same set of SSR for some crops, and modified marker sets for others, were used on a larger number of varieties.

The aims of the Comparative Tests (CTs)

The aim of the first comparative test was to provide lists of DNA-based markers which could be used to discriminate varieties of *Zea mays* (maize), *Oryza sativa* (rice), *Triticum durum* and *Triticum aestivum* (wheat) as well as *Glycine max* (soybean). The aim was also to compare results between participant laboratories, and evaluate whether it is possible to obtain the same band patterns and allele sizes even when using different reactants, equipment and working protocols. Varieties and markers were the same for all participating laboratories for each crop. The aim of the second CT was to verify if the marker set was polymorphic enough to provide unique DNA-based patterns for a larger variety set, and also to select more suitable SSRs. A third CT was run to further expand the range of varieties tested for identification by the SSR markers validated during the first and second CTs; and to include additional laboratories that would run the SSRs and varieties tested during the second CT.

Summary of results for all 3 rounds for all species

Results of these CTs have been very good for all crops.

Table 1 summarizes the origin of the participating laboratories for the CTs, the crops included in this study and the different visualization methods used by different laboratories.

Table 1: list of participating laboratories and different visualization methods used by each laboratory over 3 test rounds.

| Crop group | Participating lab | Visualization methods used | | |
|---------------|-------------------|-------------------------------|-----------------|--------------------------|
| gr | CT1 | CT2 | CT3 | |
| Wheat | Canada x2 | Canada x2 | Austria | Licor 4200 and 4300, |
| | France | France | Argentina | ABI 3130xl and Silver |
| | Italy | Italy | Canada x3 | stain |
| | | | France | |
| | | | Italy | |
| Rice | Canada | Canada | Canada | ABI3100 and 3130, |
| | Italy | Italy | India | Licor 4300 and Agarose |
| | Taiwan Province | Taiwan Province | Italy | |
| | of China | of China | USA x2 | |
| | | | Taiwan Province | |
| | | | of China | |
| Soy | Argentina | Argentina | Argentina x3 | Silver stain and ABI |
| | Brazil | Brazil | Brazil | 3100 |
| | Canada | Canada | Canada | |
| | | USA | USA | |
| Maize | Argentina | Argentina | Brazil | ABI3130xl, Silver stain, |
| | Brazil | Brazil | Canada | ABI 3100 and Silver |
| | Canada | Canada | China | Stain |
| | France | France | France x2 | |
| | | | Italy | |
| | | | USA x2 | |

A summary of the results obtained for each is below:

Wheat

SSR markers assessed in collaborative trials

Table 2

| CT1 | CT2 | CT3 | CT4 (*) | Status (CT1 to 3) |
|---------|---------|---------|----------|---------------------------------------|
| DuPw004 | DuPw004 | DuPw004 | | Good |
| DuPw115 | DuPw115 | DuPw115 | | Good |
| DuPw167 | DuPw167 | DuPw167 | | Good |
| DuPw205 | DuPw205 | DuPw205 | | Good |
| DuPw217 | DuPw217 | DuPw217 | | Good |
| Xgwm003 | Xgwm003 | Xgwm003 | | Good |
| Xgwm099 | | | | May be complex in hexaploid varieties |
| Xgwm526 | | | | May be complex in hexaploid varieties |
| | Xgwm155 | Xgwm155 | | Good |
| | Xgwm413 | Xgwm413 | | Good |
| | | | Xbarc074 | Will be evaluated after CT4 |
| | | | Xbarc184 | Will be evaluated after CT4 |
| | | | Xbarc347 | Will be evaluated after CT4 |
| | | | Xgwm052 | Will be evaluated after CT4 |
| | | | Xgwm095 | Will be evaluated after CT4 |
| | | | Xgwm372 | Will be evaluated after CT4 |

(*) CT not yet completed

All markers assessed within a collaborative trial may be combined in a single multiplex PCR.

In CT1, eight varieties were included from Brazil (2), Canada (2) and Italy (4). In CT2, four varieties were added from each of Brazil, Canada, France and Italy, giving a total of 24 varieties. In CT3, new participants examined these same 24 varieties while the original participants each examined larger numbers of varieties specific to their countries; 24 each from France and Canada and 12 from Italy. In total 84 wheat varieties have been examined. Of these, all but one pair of Italian varieties and two pairs of Canadian varieties could be identified using the specified markers. The objective of CT4 is to test an additional set of six multiplexed markers which may enhance discrimination among varieties.

Rice

SSR markers tested and validated for all 3 rounds

Table 3

| CT1 | CT2 | CT3 | Status |
|-------|-------|-------|-----------------------------|
| | | RM1 | Will be evaluated after CT4 |
| | | RM19 | Will be evaluated after CT4 |
| RM70 | RM70 | RM70 | Will be evaluated after CT4 |
| | RM101 | | |
| RM105 | RM105 | | |
| | RM151 | | |
| | | RM154 | Will be evaluated after CT4 |
| RM159 | RM159 | | |
| | | RM171 | Will be evaluated after CT4 |
| RM215 | RM215 | | |
| | | RM237 | Will be evaluated after CT4 |
| | RM264 | | |
| RM266 | RM266 | | |
| RM276 | RM276 | | |
| | RM287 | RM287 | Will be evaluated after CT4 |
| | | RM307 | Will be evaluated after CT4 |
| | | RM316 | Will be evaluated after CT4 |
| RM333 | RM333 | RM333 | Will be evaluated after CT4 |
| | | RM334 | Will be evaluated after CT4 |
| | RM347 | | |
| | | RM413 | Will be evaluated after CT4 |
| | | RM447 | Will be evaluated after CT4 |
| | | RM510 | Will be evaluated after CT4 |
| | | RM514 | Will be evaluated after CT4 |
| RM567 | RM567 | | |

In CT 1 and 2, participating laboratories were able to demonstrate usefulness of microsatellite markers in rice variety testing and established DNA extraction, PCR setup and visualization approaches that could produce consistent results under variable conditions. The third CT was set up to evaluate 12 markers selected from a core set of rice SSRs previously shown to be discriminatory on a large set of Asian varieties. Six laboratories participated, including laboratories from Taiwan Province of China, Canada, Italy, India and the USA on varieties from Taiwan Province of China, India and Italy. Based on these results, some minor adjustments to the conditions have been made to improve performance and a forth CT is in preparation, which will determine if these markers are

suitable to identify varieties from more countries. An additional 26 Italian varieties have been obtained for this work, and the group is working to establish contacts with other Asian countries to try to secure additional varieties. The countries of key interest include China, India, Indonesia, Japan, Republic of Korea, Philippines, Thailand, and Viet Nam.

Soybean

SSR markers tested and validated for all 3 rounds

Table 4

| CT1 | CT2 | CT3 | CT4 (*) | Status (CT1 to 3) |
|---------|---------|---------|---------|-----------------------------|
| ATT-177 | ATT-177 | ATT-177 | | Very good |
| ATT-094 | ATT-094 | ATT-094 | | Good |
| ATT-233 | ATT-233 | ATT-233 | | Good |
| ATT-307 | ATT-307 | ATT-307 | | Good |
| ATT-105 | ATT-105 | | | Not good |
| ATT-114 | ATT-114 | | | Not good |
| ATT-216 | ATT-216 | | | Not good |
| ATT-231 | ATT-231 | | | Not good |
| AT001 | | | | Not good |
| ATT-353 | | | | Not good |
| ATT-446 | | | | Not good |
| ATT-534 | | | | Not good |
| ATT-577 | | | | Not good |
| | ATT-030 | ATT-030 | | Very good |
| | ATT-181 | ATT-181 | | Very good |
| | ATT-311 | ATT-311 | | Good |
| | ATT-449 | ATT-449 | | Good |
| | | ATT-070 | | Not good |
| | | ATT-147 | ATT-147 | Will be evaluated after CT4 |
| | | ATT-180 | ATT-180 | Will be evaluated after CT4 |
| | | ATT-191 | | |
| | | ATT-373 | ATT-373 | Will be evaluated after CT4 |
| | | ATT-703 | | |
| | | | ATT-080 | Will be evaluated after CT4 |
| | | | ATT-352 | Will be evaluated after CT4 |
| | | | ATT-540 | Will be evaluated after CT4 |
| | | | ATT-600 | Will be evaluated after CT4 |
| | | | ATT-728 | Will be evaluated after CT4 |

(*) not finished yet.

The status "good" and "very good" of the marker means that repeatable and reproducible results were obtained for that marker during 2-3 rounds of comparative tests.

For the first CT, 8 soybean varieties, 4 from Brazil and 4 from Argentina were analyzed using 12 SSR markers (see Table 4). For the second CT, 16 additional soybean varieties, 4 from Brazil, 4 from Canada and 8 from Argentina were analyzed making a total of 24 varieties. Twelve SSRs were tested, 4 new and 8 from the previous CT (see Table 4). During the third round, 14 SSRs were tested: 4 were tested for the third time, 4 for the second time and 6 were new ones. The 24 varieties used for this analysis were from: Brazil (8), Argentina (12) and Canada (4). A forth CT is in progress to validate new SSR markers by comparing results (reproducibility and repeatability of the markers) among and within participating laboratories. Each laboratory will use their own

methodology for obtaining the results. Three SSR markers were already run during the third comparative test and 5 are new ones. These markers are being tested on a set of 24 varieties from Brazil (8), Canada (4) and Argentina (12).

Maize

The maize group has established an SSR set that is ready to be applied for the organization of a Performance Test. For the Performance Test it is necessary to have a core set of varieties to be used as reference material (RM). These varieties will be tested using the markers already selected during the previous comparative tests.

| CT1 | CT2 | CT3 | Status (CT1 to 3) |
|------------|------------|------------|-------------------|
| Phi 109275 | Phi 109275 | Phi 109275 | Very good |
| Phi083 | Phi083 | Phi083 | Good |
| Phi 102228 | Phi 102228 | Phi 102228 | Very good |
| Phi015 | Phi015 | Phi015 | Good |
| Umc 1545 | Umc 1545 | Umc 1545 | Very good |
| Umc 1061 | Umc 1061 | Umc 1061 | Very good |
| Phi 032 | Phi 032 | | Good |
| Umc 1122 | | | Good |
| Phi 093 | | | Good |
| Phi 452693 | | | Not good |
| Umc 1153 | | | Not good |
| Umc 1152 | | | Not good |
| | Umc 1448 | Umc 1448 | Good |
| | Umc 1117 | Umc 1117 | Good |
| | Umc 1133 | Umc1133 | Very good |
| | Bnlg 1782 | | Good |
| | Umc 1792 | | Not good |
| | | Phi 233376 | Good |
| | | Bnlg 1129 | Not good |
| | | Umc 1478 | Good |

Further steps

- Once the marker sets have been established, a defined set of "reference varieties" will be identified for each crop that can used by laboratories that wish to establish protocols for variety verification.
- Following this, the group will work to define a strategy together with ISTA's Statistics Committee for the accreditation of laboratories performing DNA-based variety verification tests. This means specification of how the proficiency testings (PTs) will be carried out and consideration of reference materials required for such tests.
- Finally, a proficiency testing (PT) program will be established in the future to assist laboratories
 develop testing capacity for varietal verification for these crops using molecular markers and to
 promote use of these internationally standardized methods.

Acknowledgement

We would like to acknowledge to all private and public institutions that kindly collaborated with these tests and to ISTA who gave technical and administrative support.